

# Understanding Quality Control for Infectious Disease Testing

Wayne Dimech  
ASLM Quality Control Workshop  
Abuja, Nigeria  
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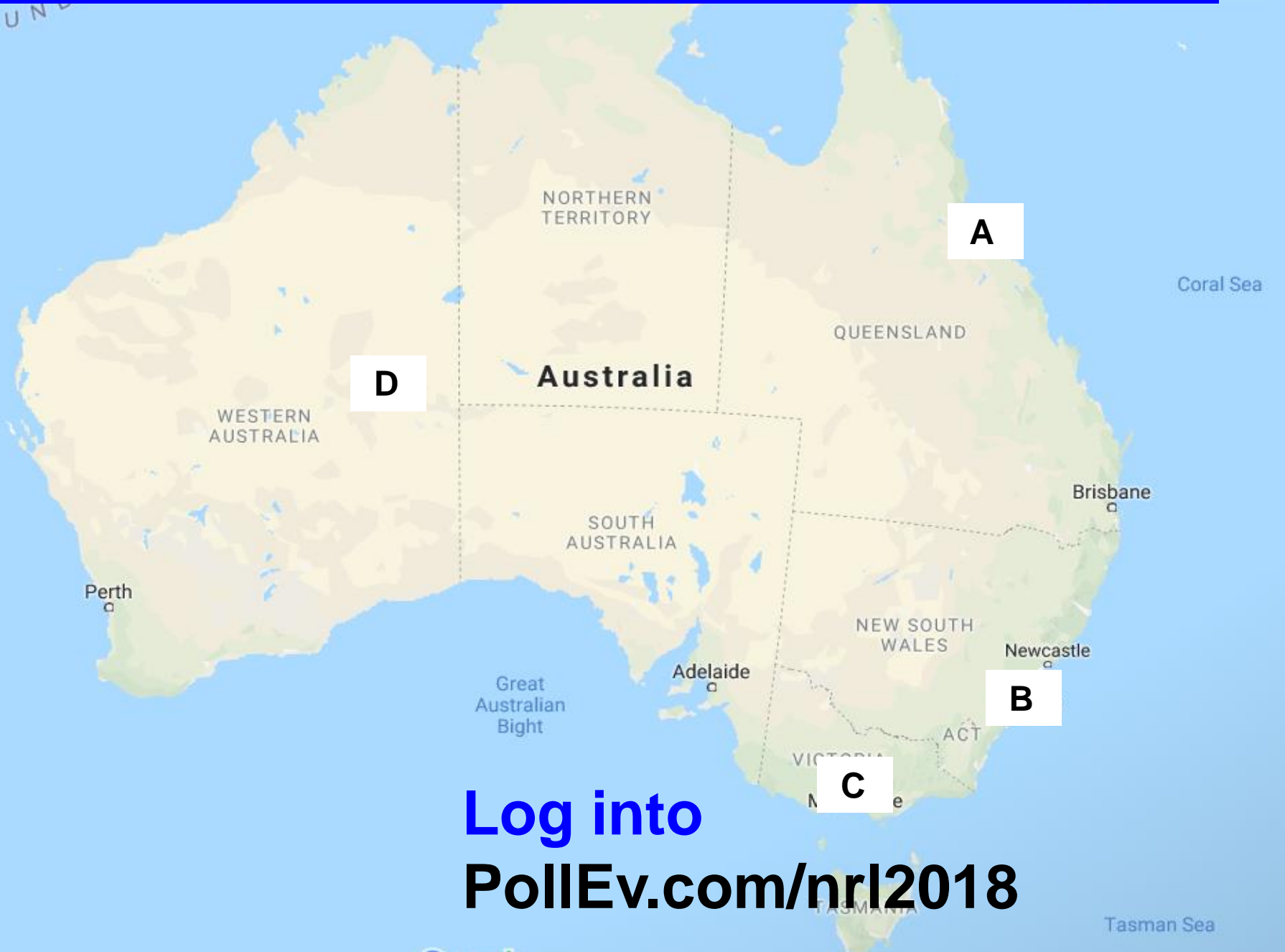
# Disclosure

- Attendance at ASLM part-sponsored by CDC
- No personal financial gain

# Acknowledgements

- Joe Vincini – NRL
- Katy Yao – CDC
- Anna Murphy

Which selection represents the location of Melbourne?



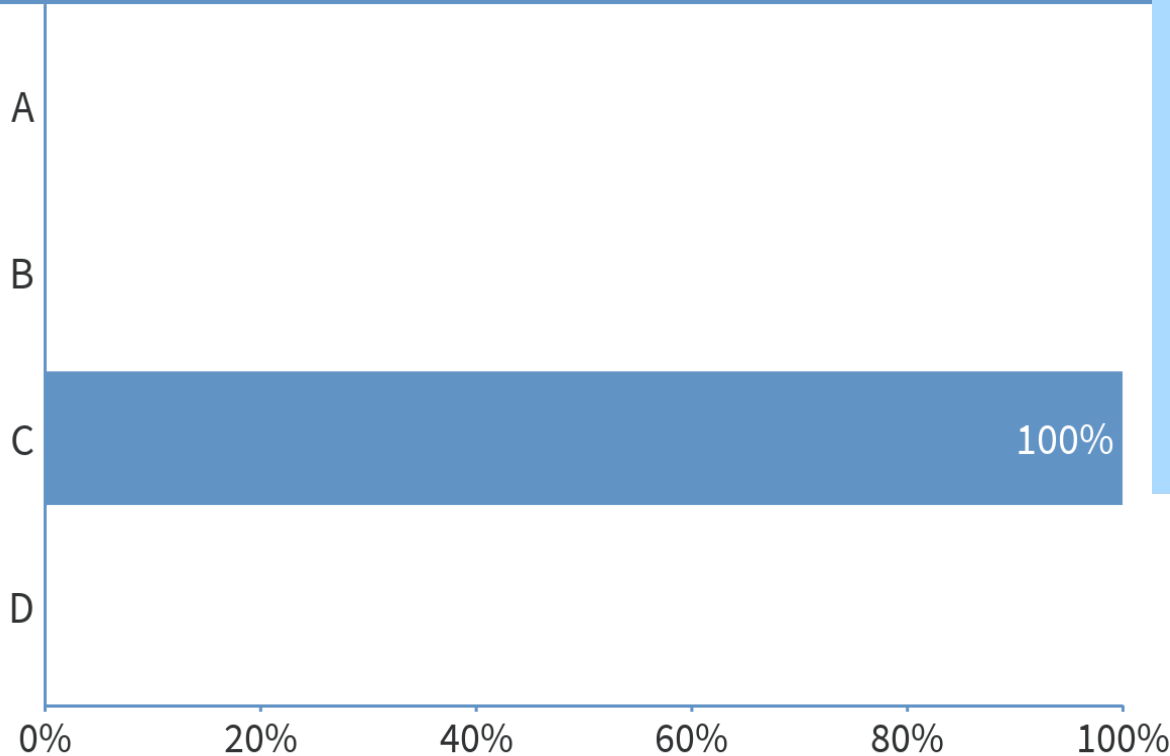
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[PollEv.com/nrl2018](https://www.PollEv.com/nrl2018)

# Which selection represents the location of Melbourne?

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## Which selection represents the location of Melbourne, Australia?





# NRL

- Established in 1985
- Not-for-profit organisation that exists to support laboratories that perform testing for the diagnosis and management of human infectious disease
- Funded partially by the Australian Government
- Mission

*To promote the quality of tests and testing for infectious diseases globally.*

# NRL

- Our objectives are achieved by providing:
  - comprehensive and innovative quality assurance services;
  - evaluations of tests and test algorithms;
  - specialised laboratory testing services;
  - training with sustainable outcomes;
  - consultation and advice on policy relating to laboratory testing.



# NRL

- Major Stakeholders:
  - Australian Government (DoH, DFAT, TGA)
  - WHO, US CDC, Global Fund, UNDP
  - Australian Red Cross Blood Service
  - Test kit manufacturers
  - Laboratories (blood screening and clinical)

# NRL

## ● Credentials:

- WHO Collaborating Centre
- Certified to Management Standard AS/NZS ISO 9001:2008
- Accreditation as a Medical Testing Laboratory; Compliant with ISO/IEC 15189: 2007
- Accredited as EQAS Provider to ISO 17043:2010
- Licensed by TGA under the *Code of Good Manufacturing Practice - Human Blood and Tissue:2000*

# Major Activities

- NRL Evaluations
- NRL Training
- NRL Testing
- NRL Workshop
- Quality Assurance
  - EQAS
  - QC

# Who am I?

- Medical scientist
  - Fellow AIMS
  - Fellow RCPA (faculty of science)
  - MBA
- Microbiology scientist
- Laboratory auditor
- WHO consultant
- Standards Australia committee member
- Reviewer of NRL's QC program results



# What I am not

- Clinician
- Clinical biochemist
- Statistician
- Salesman
- Solver of all things QC

# History of Quality Control

- Walter A. Shewhart of the Bell Telephone Laboratories (1924) introduced the first control chart
- W. Edwards Deming of US Bureau of the Census (1940s) used statistical sampling techniques
- Post WWII emergence in Japanese manufacturing
- Deming Cycle of quality control in 1950s
- Adoption of control charts by Levey and Jennings (1950) in clinical chemistry

# History of Quality Control

- Invention of control rules by Westgard et al (1981)
- Six Sigma was pioneered by Motorola in 1980s
- Operational Process Specifications charts (OPSspecs) in 1994 by Westgard
- Six Sigma as applied to clinical chemistry introduced by Westgard in 2001
- CLSI Guidelines 3rd ed. Vol. C24-A3 (2006)
- CLSI Guidelines 4th ed. Vol. C24 (2016)

# History of Quality Control

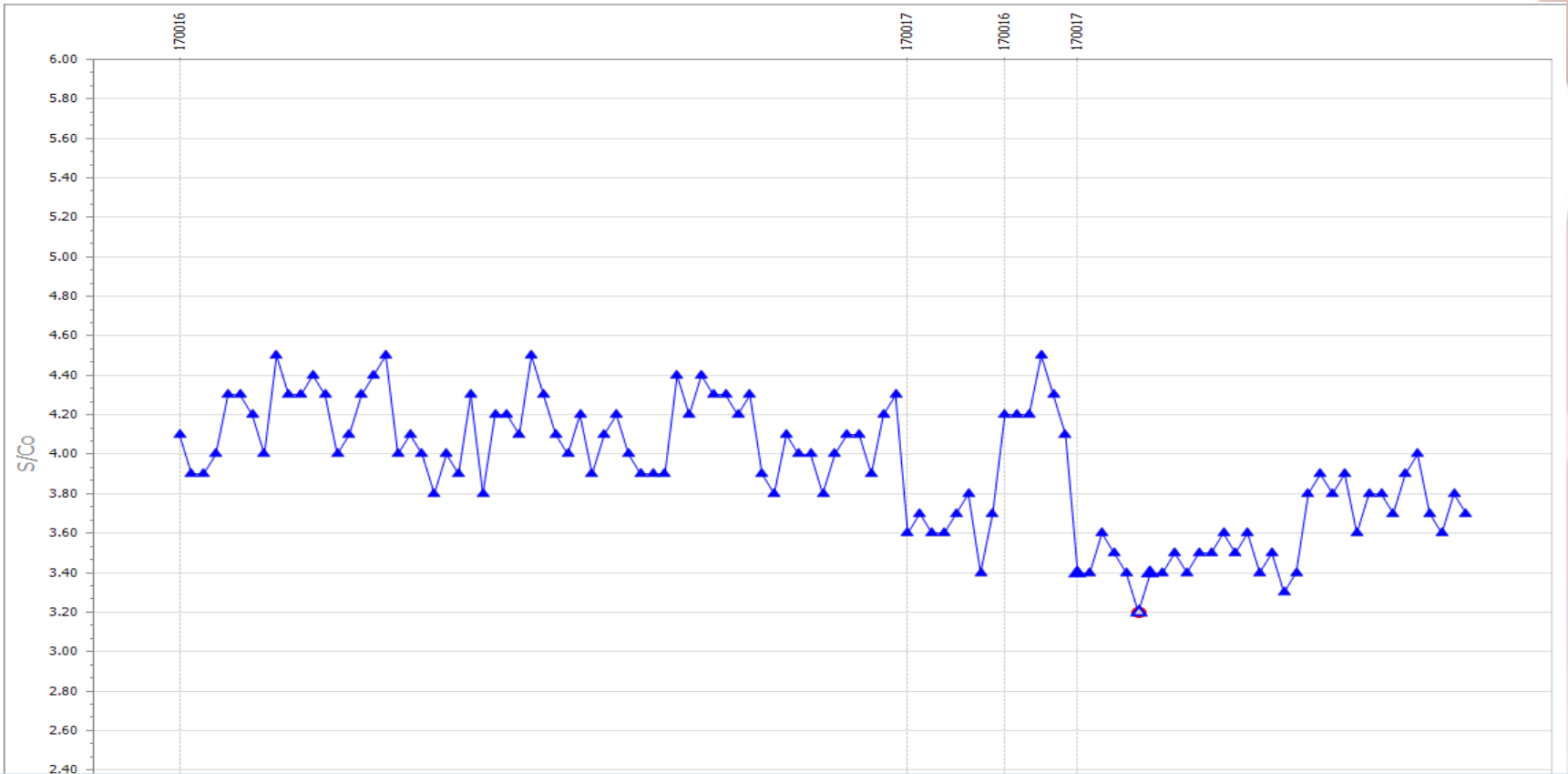
- Since 1990s only three non-NRL published papers on QC applied to infectious disease serology
  - One on HBsAg testing
  - Two reviewing antibody testing – limited reagent lot changes
- No systematic studies on QC for viral load testing





Point 1:  
Measuring systems must be fit for purpose

# Variation



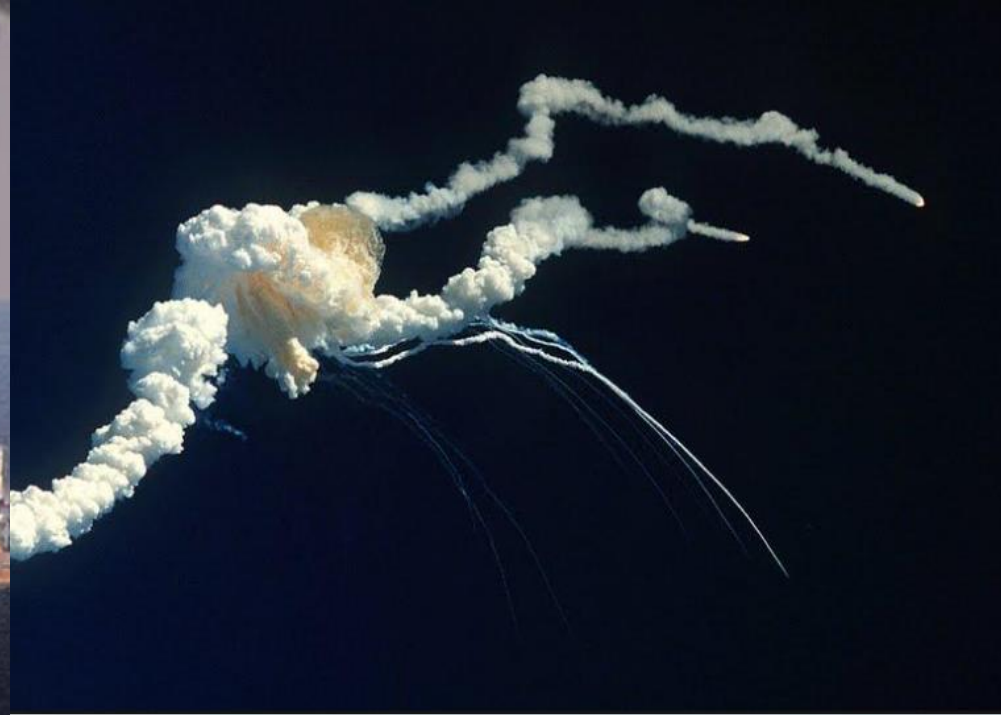
Point 2:

Measurement systems have *normal* variation



# Titanic

- Captain Smith ignored seven iceberg warnings from his crew and other ships
- Speeding to make crossing in 6 days
- Rivets made of sub-standard iron
- Water-tight compartments compromised to allow more 1<sup>st</sup> class room
- There were too few lifeboats
- Captain of **SS Californian** took no action



- People
- Processes
- Components
- Equipment



# Variation

- Reagent lots
- Instrument and equipment
- Calibrations and maintenance
- Operators
- Storage and transport conditions
- Environmental conditions

# Why Run Quality Control?

To monitor

- People
- Processes
- Components
- Equipment

Point 3:

Variation is derived from people, process, components and equipment

# Common terms

## ● Accuracy

- the ability to measure the true value correctly on average

## ● Precision

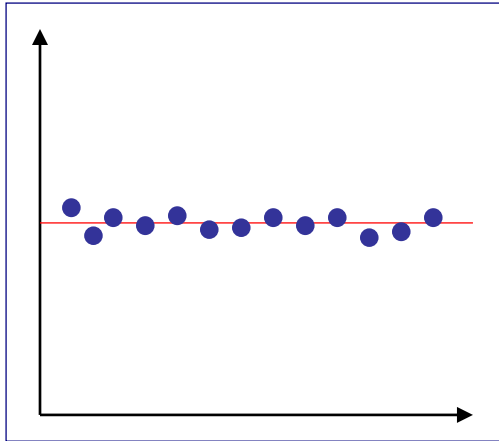
- a measure of inherent variability in the measurement (the repeatability of a result)

## ● Bias

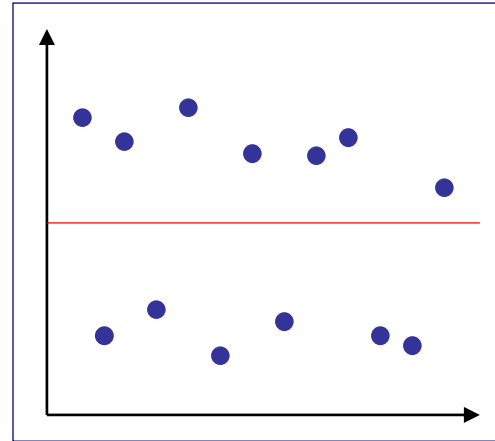
- the difference between the observed value and the expected/target value



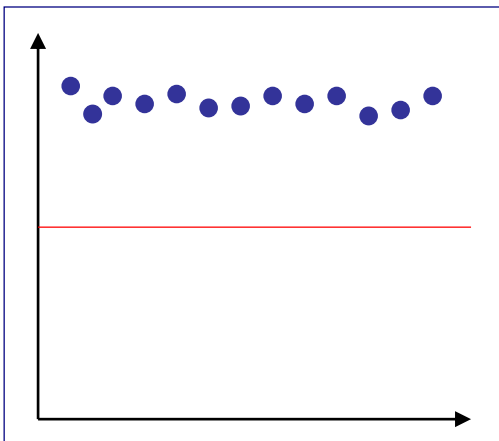
# Common terms



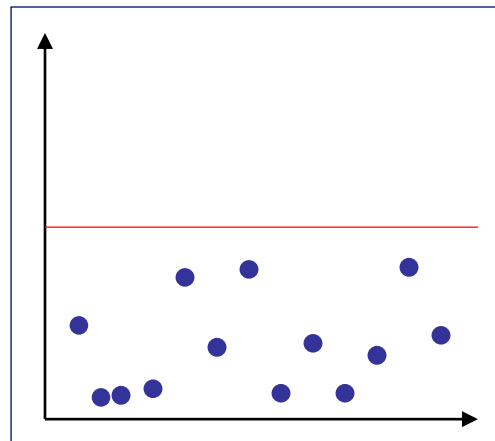
**A.**  
Accurate  
and  
Precise



**B.**  
Accurate  
(on  
average)  
but not  
Precise



**C.**  
Precise  
but not  
Accurate



**D.**  
Neither  
Precise nor  
Accurate

# Quality Control

## EQAS

- Monitors integrity of entire testing process
- Snapshot in time
- Identifies systematic and random errors
- Inter-lab comparisons identify assay and/or lab problems

Pre-analytical:  
Samples processed



Analytical:  
Samples tested



Post-analytical:  
Results reported

## QC

- Monitors analytical process only
- On-going
- Identifies systematic and random errors
- Estimates precision within lab
- Inter-lab comparisons for estimation of accuracy
- Uncertainty of Measurement

# **Caveat – This talk is about:**

**Quality control for  
*infectious disease testing***

**Quality control means  
*Run control (IQC or EQC)***

# The Quality Control Process

**QC Samples**

**Testing Frequency**

**Data Management**

**Data Analyses**

**Determine Control Limits**

**Monitor Variation**

**Investigate Variation**

**Always test the manufacturer's kit controls as these are used to validate the assay**



# QC Samples

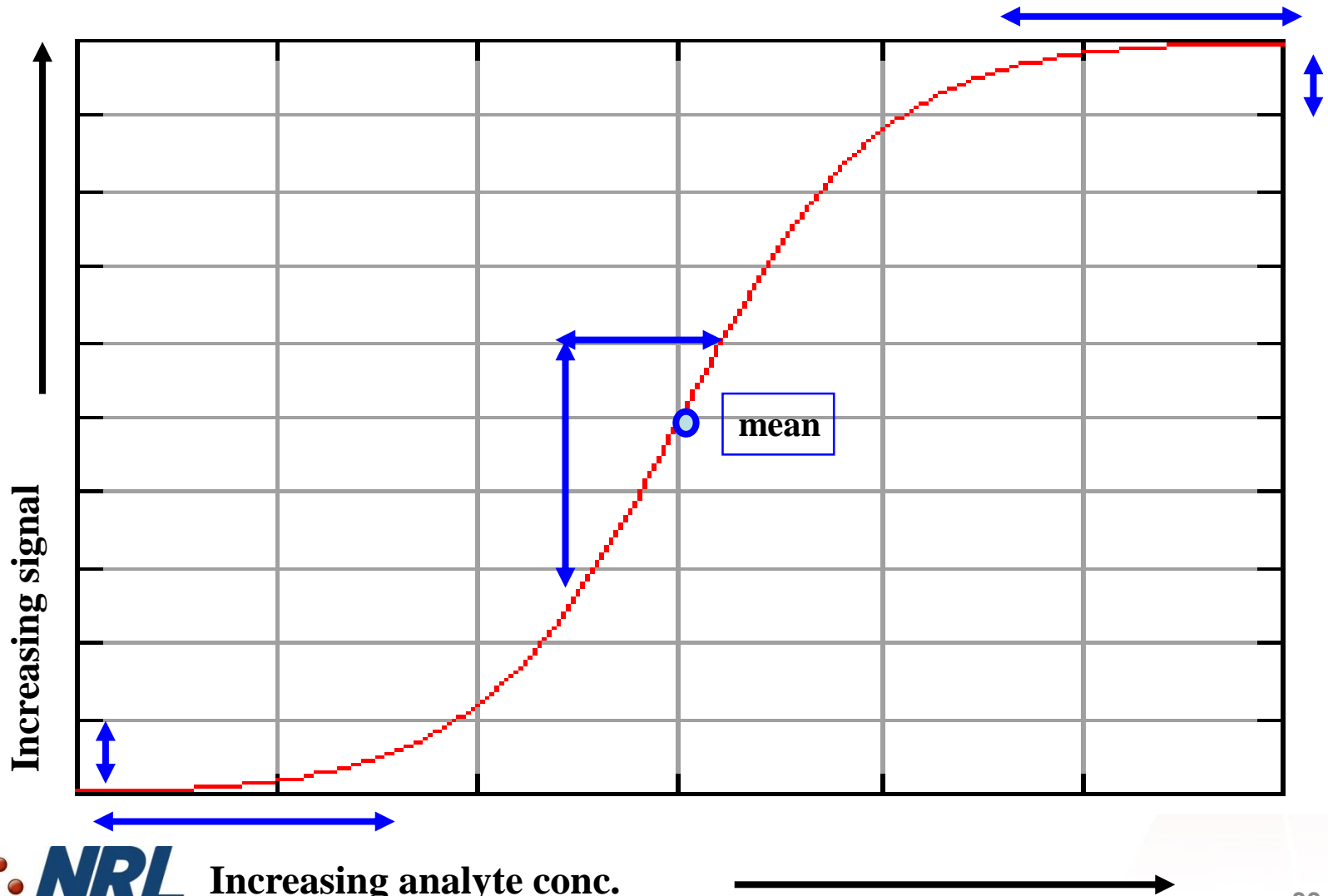
# Quality Control Sample

- Sufficient volume for extended use
- Stable over a long period
- Minimal lot-to-lot variation
- Composition similar to patient sample
- Results within the clinically significant range
- Must not “saturate” the assay
- Must be on the linear part of the curve
- Colour coded; Bar coded
- Liquid stable - no reconstitution

# Ideal QC/Assay Combination



# Serology Dose Response



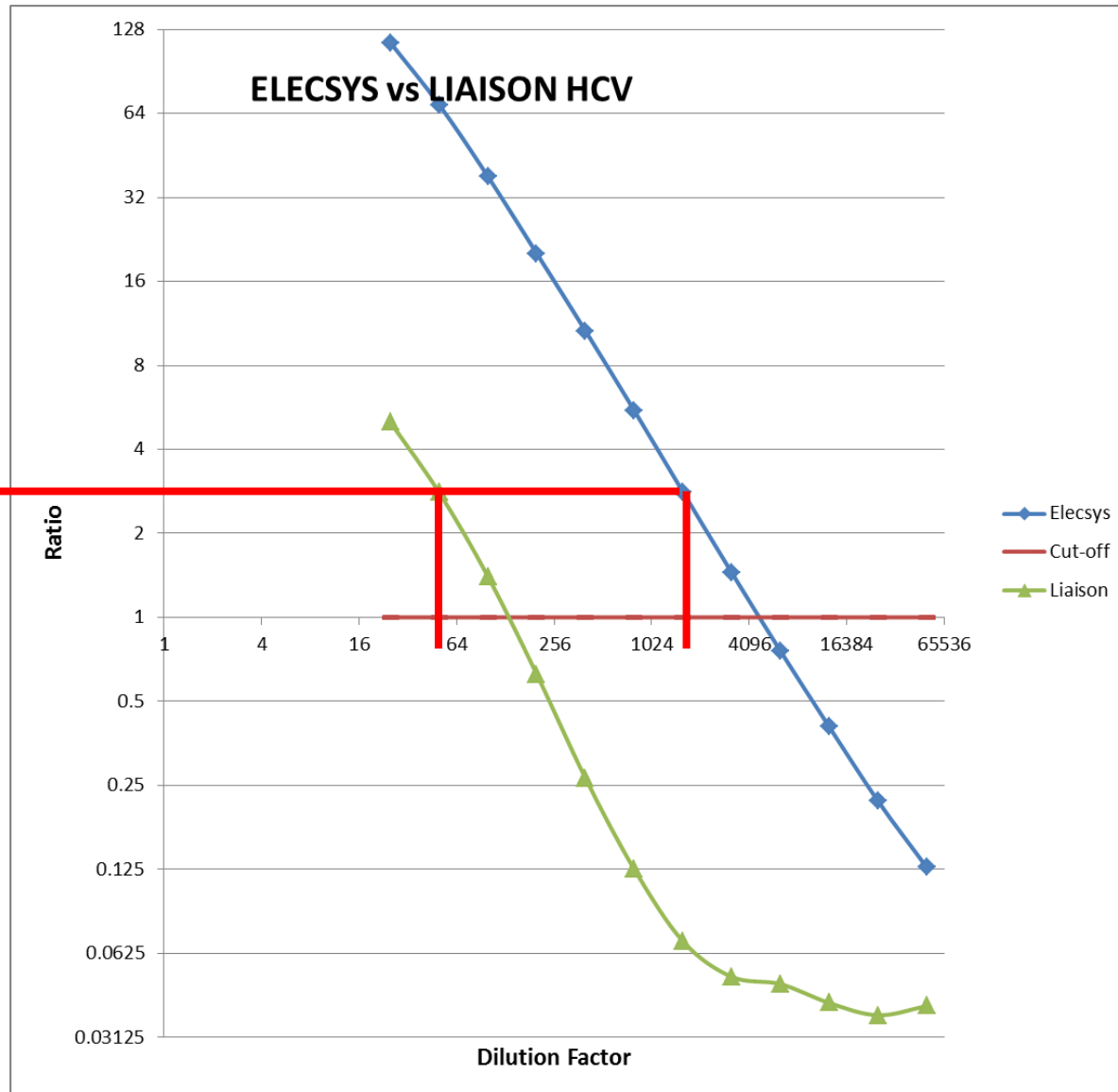


# Serology Dose Response

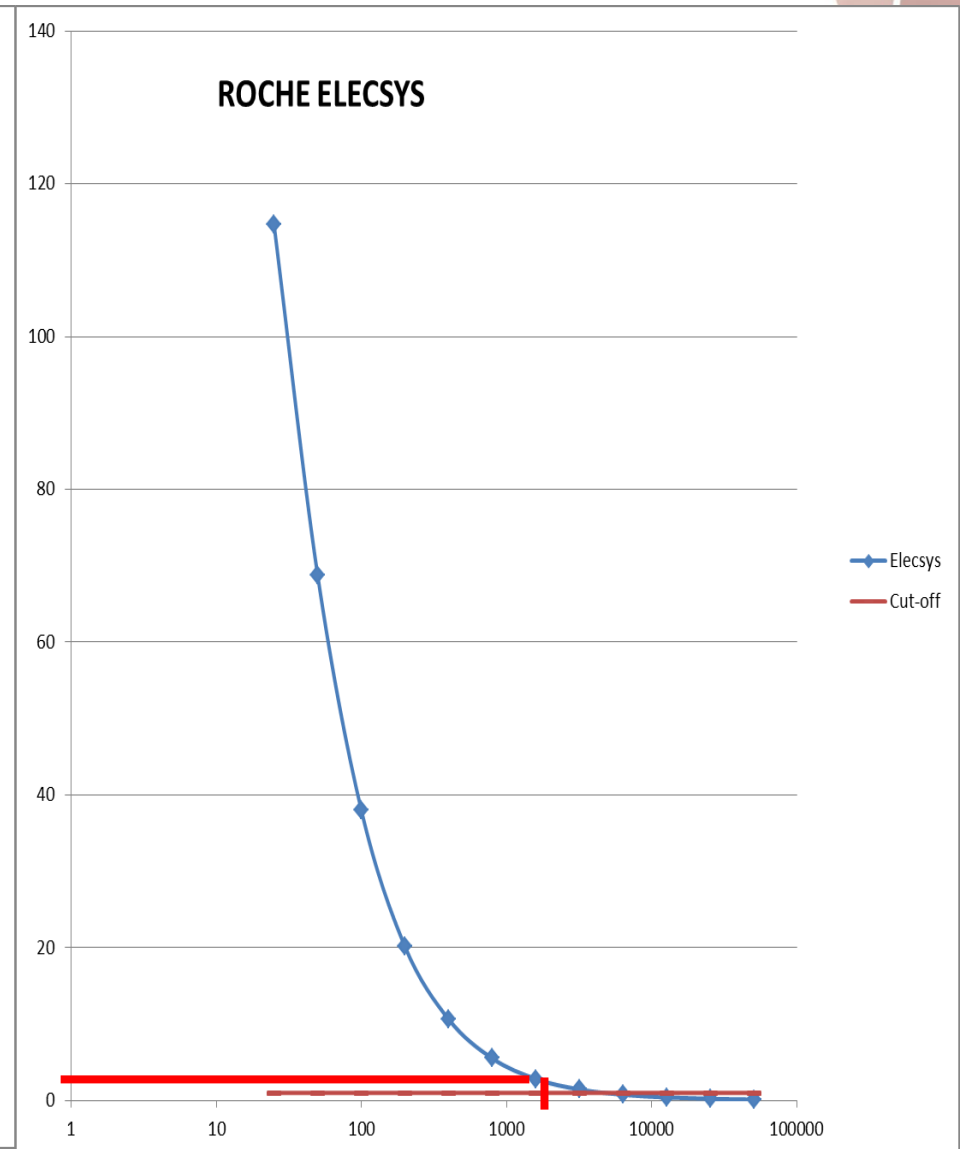
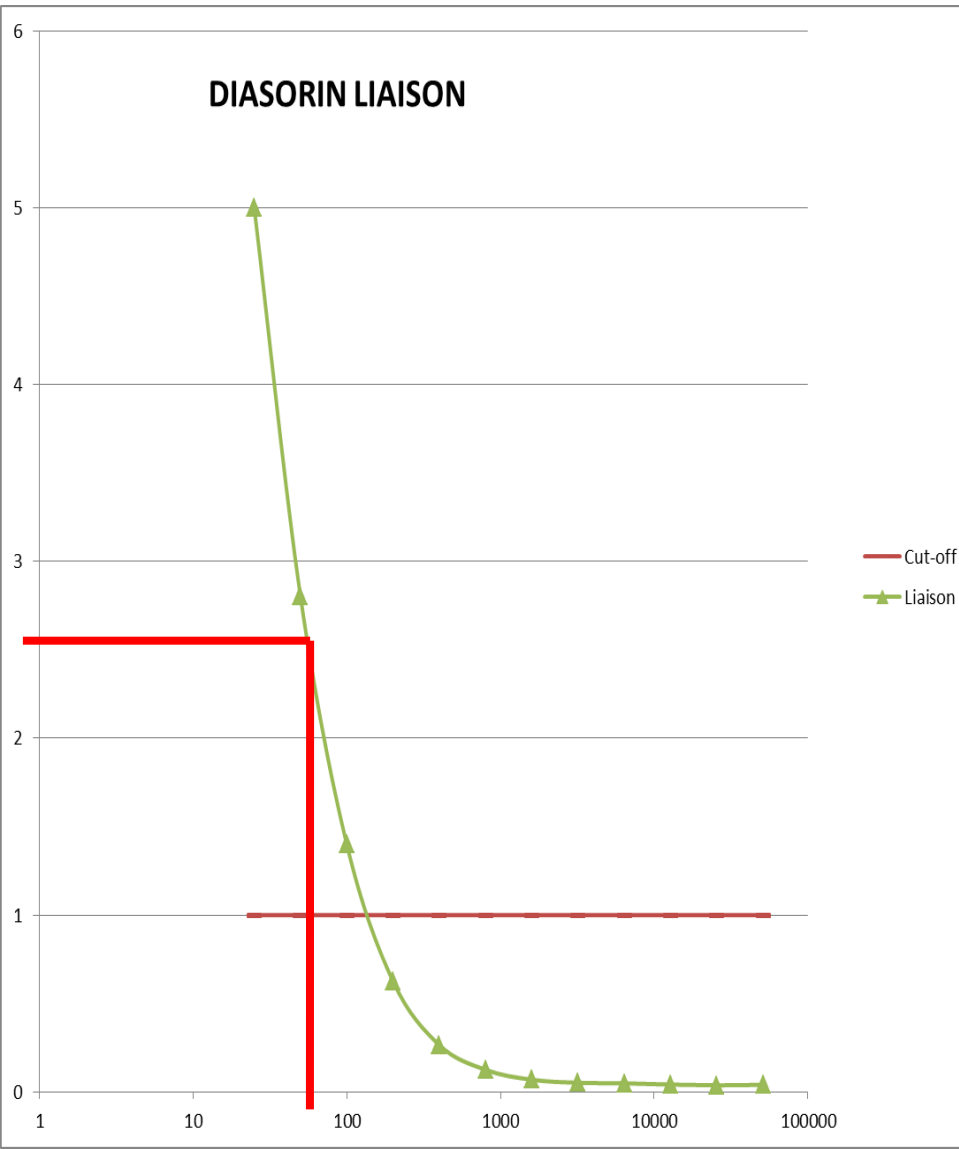
Traditional  
“obsession”  
with  
2-3x cut-off

Ideal ratio?

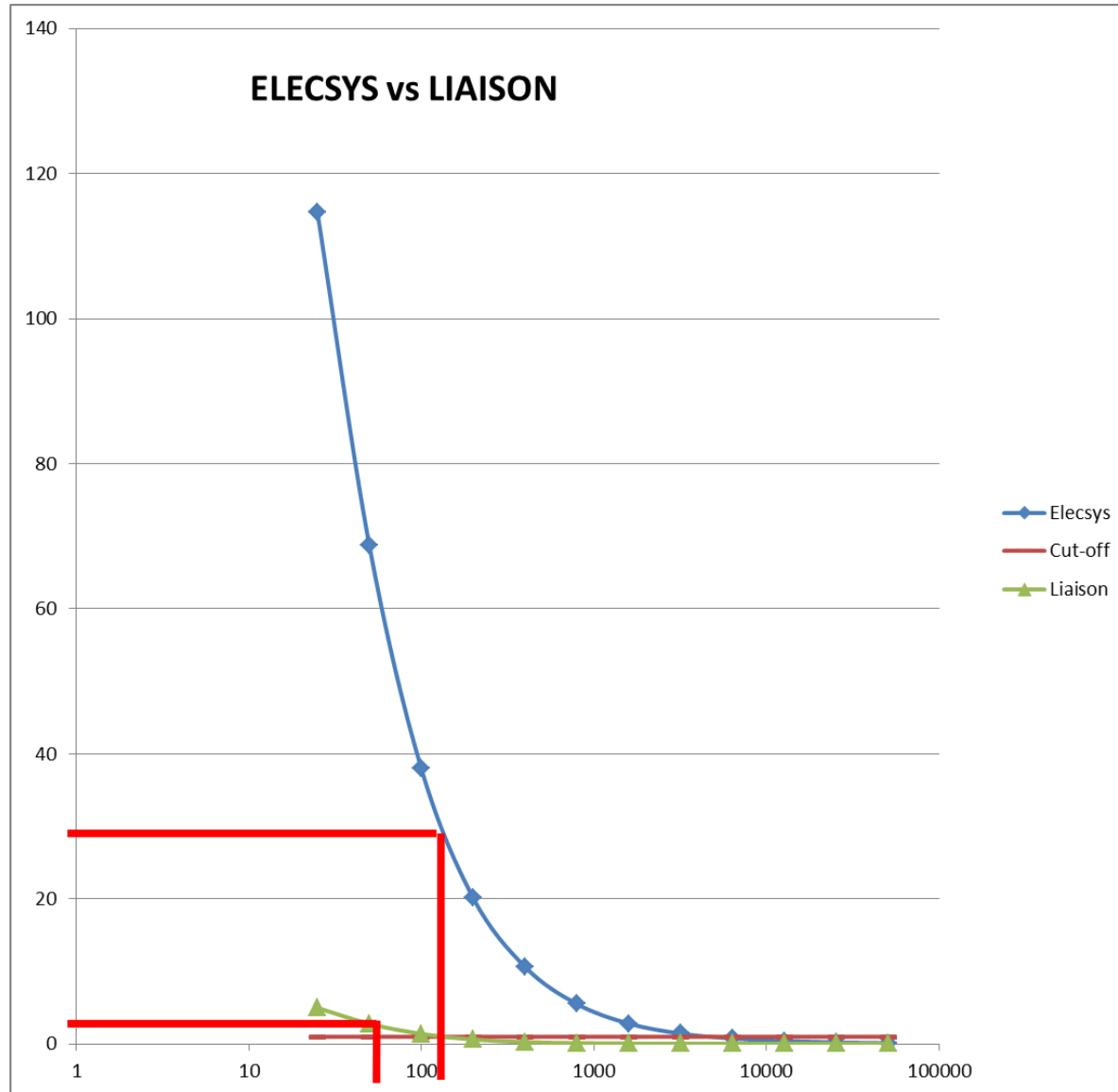
Appearances  
can be  
deceptive!!



# Serology Dose Response



# Examples of Dilution Series



Point 4:

Dynamic range of serology assays is not always linear

Point 5:

Choose QC sample that reacts at the linear portion of the dilution curve

# Differences between Clinical Chemistry and ID Assays

Clinical Chemistry	Infectious Disease Serology
Linear	Non-linear
Inert analyte	Functional biological analyte
Quantitative	Qualitative
Adjust for bias	No adjustment for bias
Lower level of regulation	Highly regulated
Several medical decision points	Single decision point
Adjust for lots variation	No adjustment for lot variation

# Differences between Clinical Chemistry and ID Assays

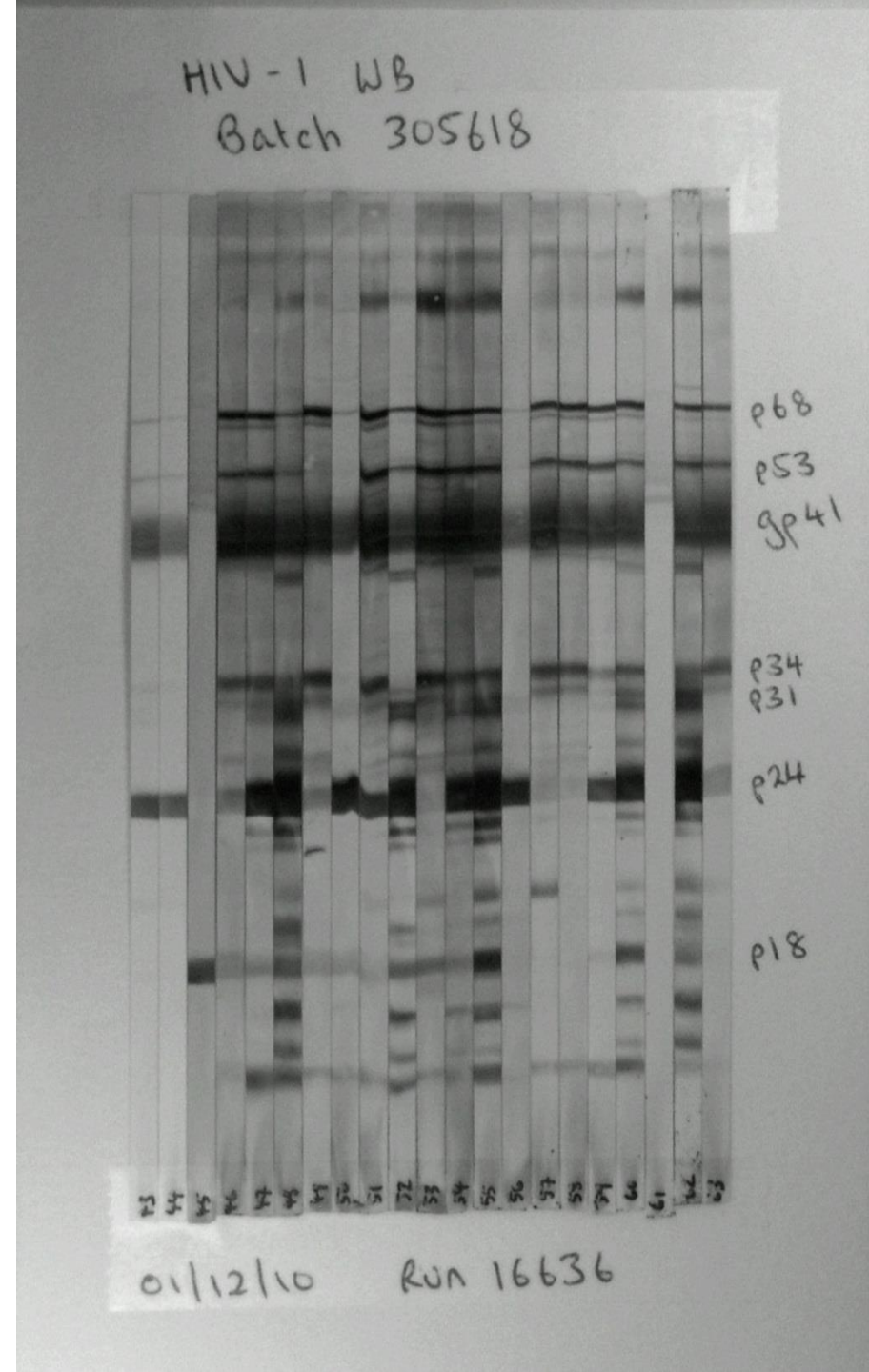
Clinical Chemistry	Infectious Disease Serology
International standards	Poor or no standards
Certified reference methods	No CRMs
Available in a pure form	Different forms
Single target	Multiple and varying targets
TEa	?? TEa

## Point 6:

There are fundamental differences between testing for an inert chemical and a functional, biological analyte

# HIV western blot

- Different Ab responses in different people
- Assay response depends on what the manufacturer has used in design





# Testing Frequency



# Testing Frequency

- No correct answer
- Cost vs risk
- Knowledge of assay
- Local and international regulation
- Minimum vs optimum

## Point 7: Recommendation

Daily at start of day for automated platforms  
and or every microtiter plate



# Data Management

# Data Management

- QC samples are a tool, not the end point
- Results collected after each test run
- Displayed graphically
- Have acceptance rules
- React immediately if unexpected results are detected

Point 8:

Monitoring QC results without reference to a peer-group only monitors precision

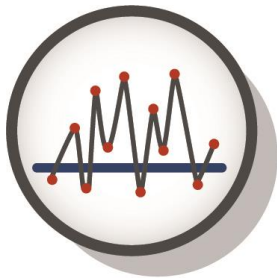
# **What We (Think) We Know About Quality Control**

# Interpretation of QC Results

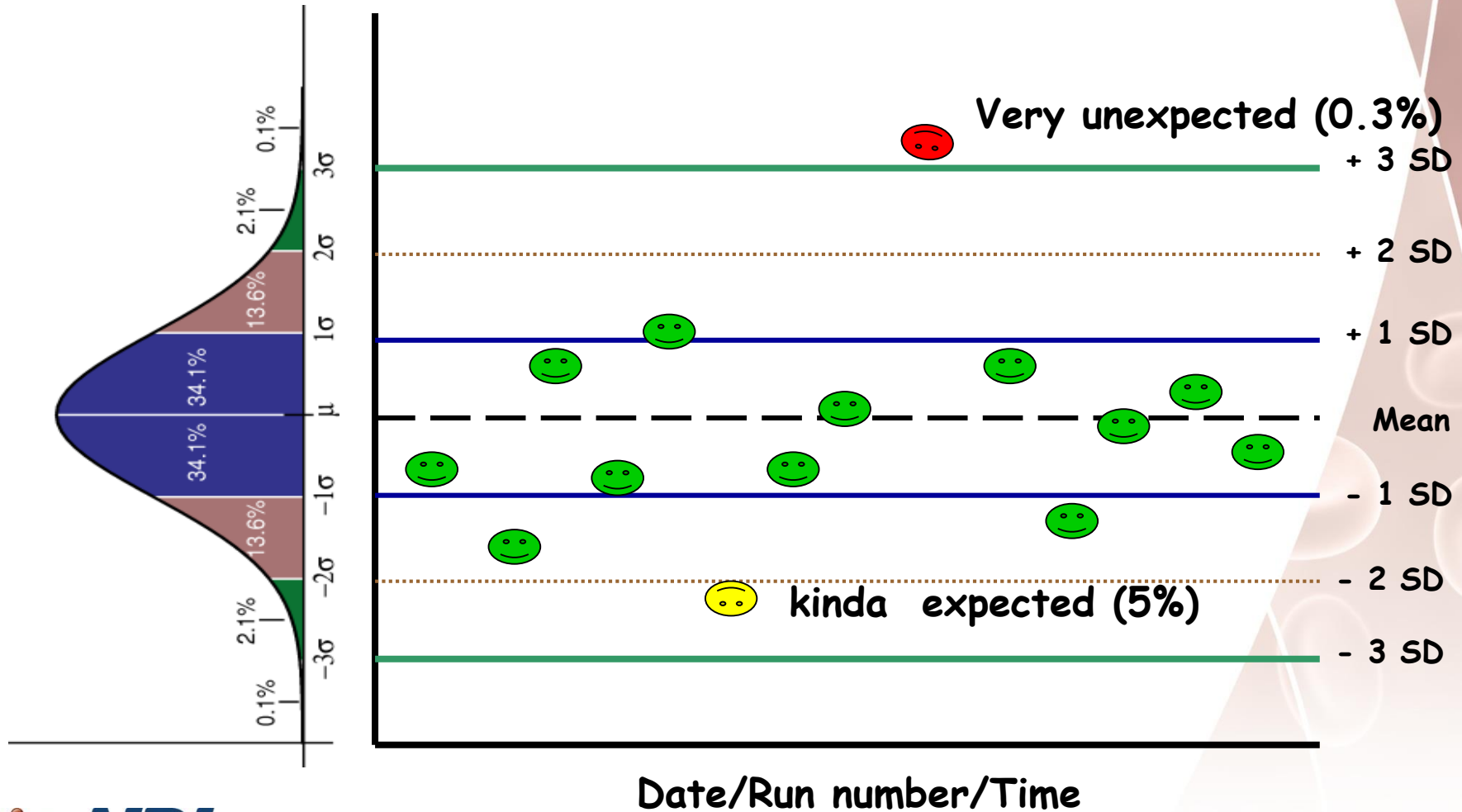


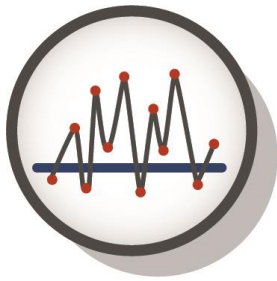
## TEA LEAF MEANINGS





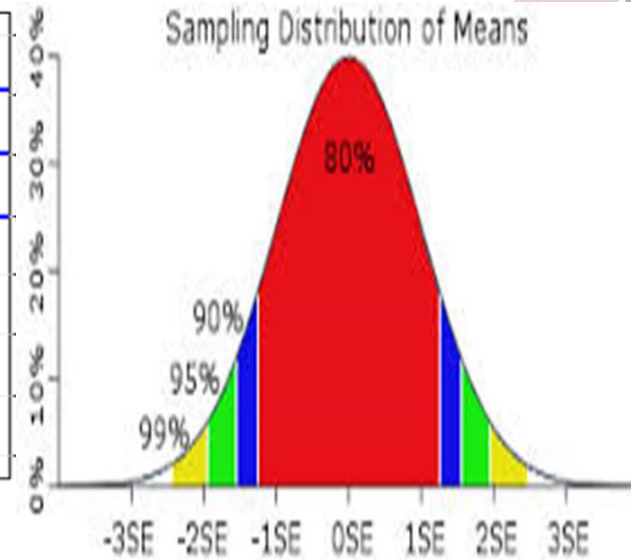
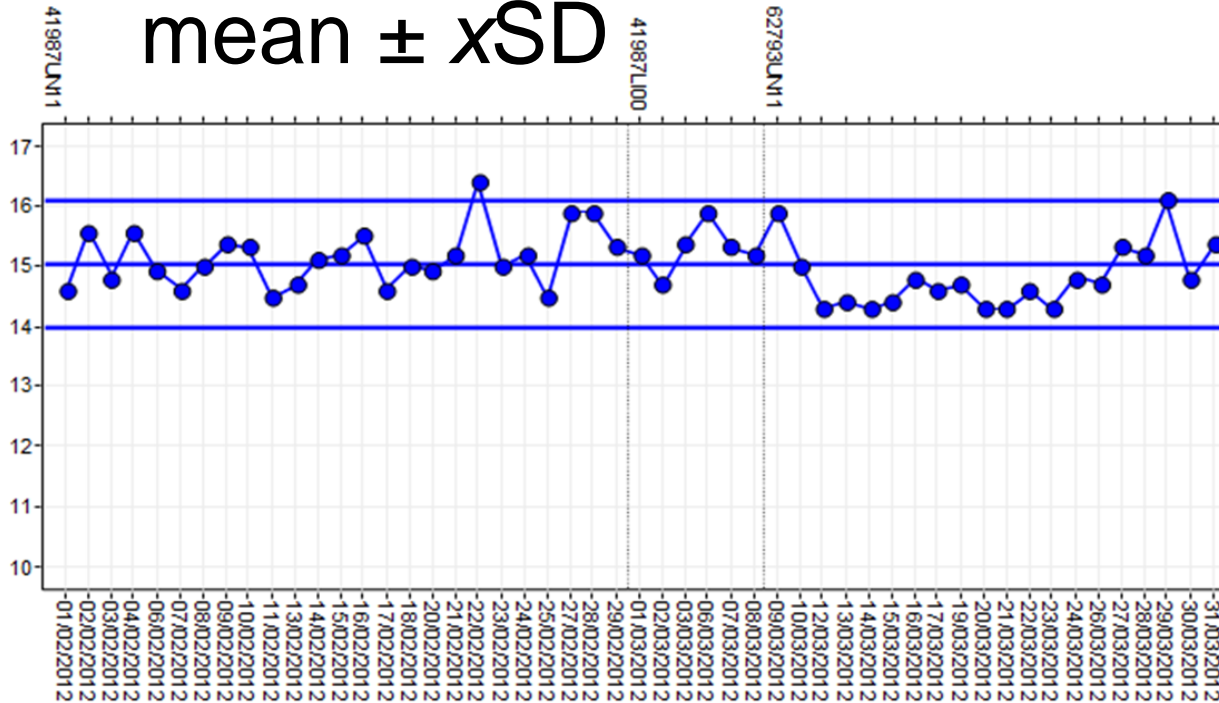
# Control Limits



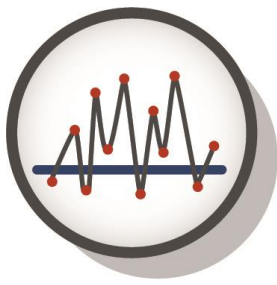


# Control Limits

- Traditional methods for setting QC limits rely on mean  $\pm$  xSD



...of what data set?



# Control Limits

- RiliBÄK standard (2015)  
**15**
- Public Health England (2015)  
**20**
- CLSI Guideline third edition C24-A3 (2006)  
**20**
- CLSI Guideline fourth edition C24 (2016)  
**20 ( recalculate periodically)**



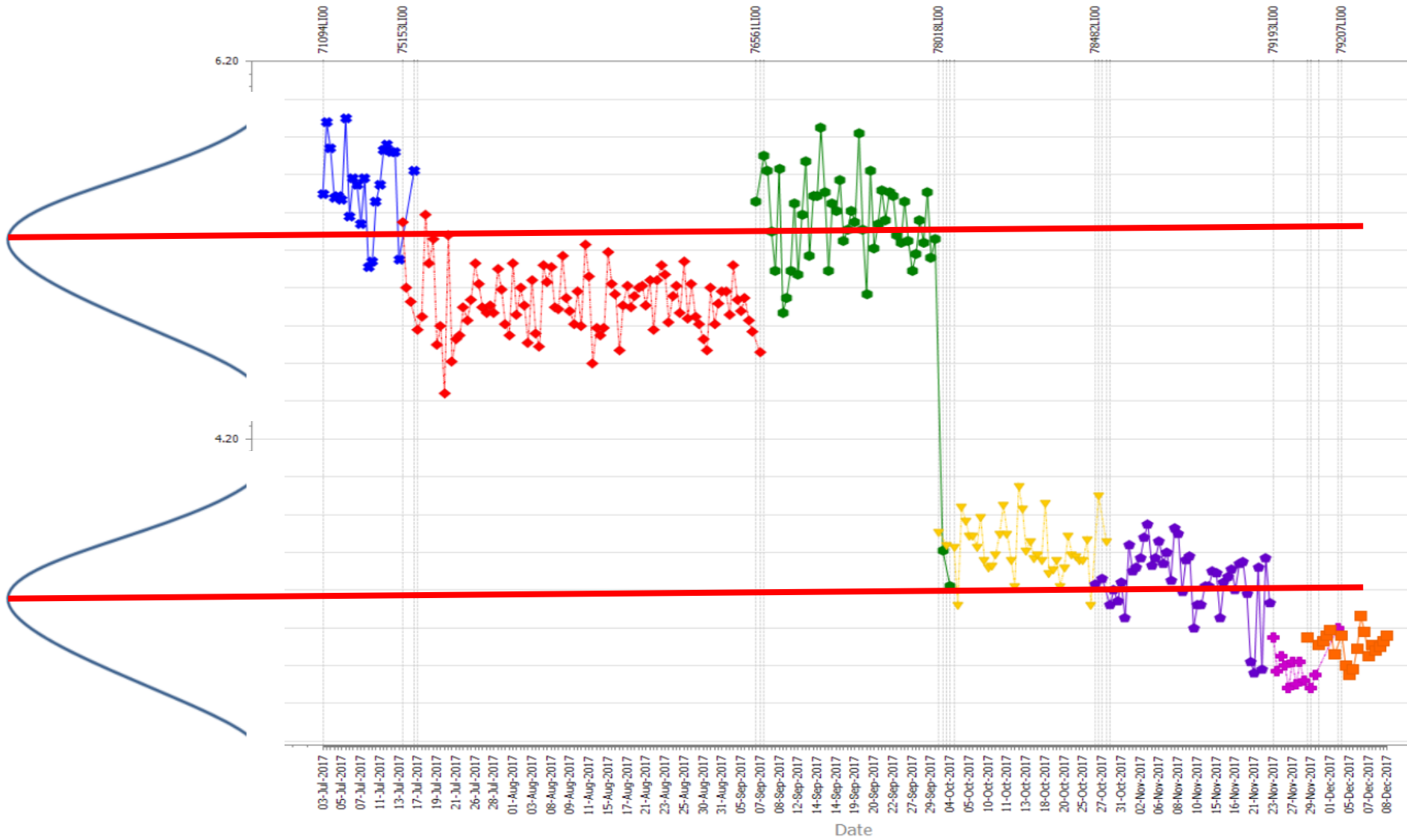
# Traditional Approaches to QC

- Assumes normal distribution of QC results
- Patient and QC sample results change proportionally (Commutability)
- Data set used to establish limits are representative of future results

Point 9:

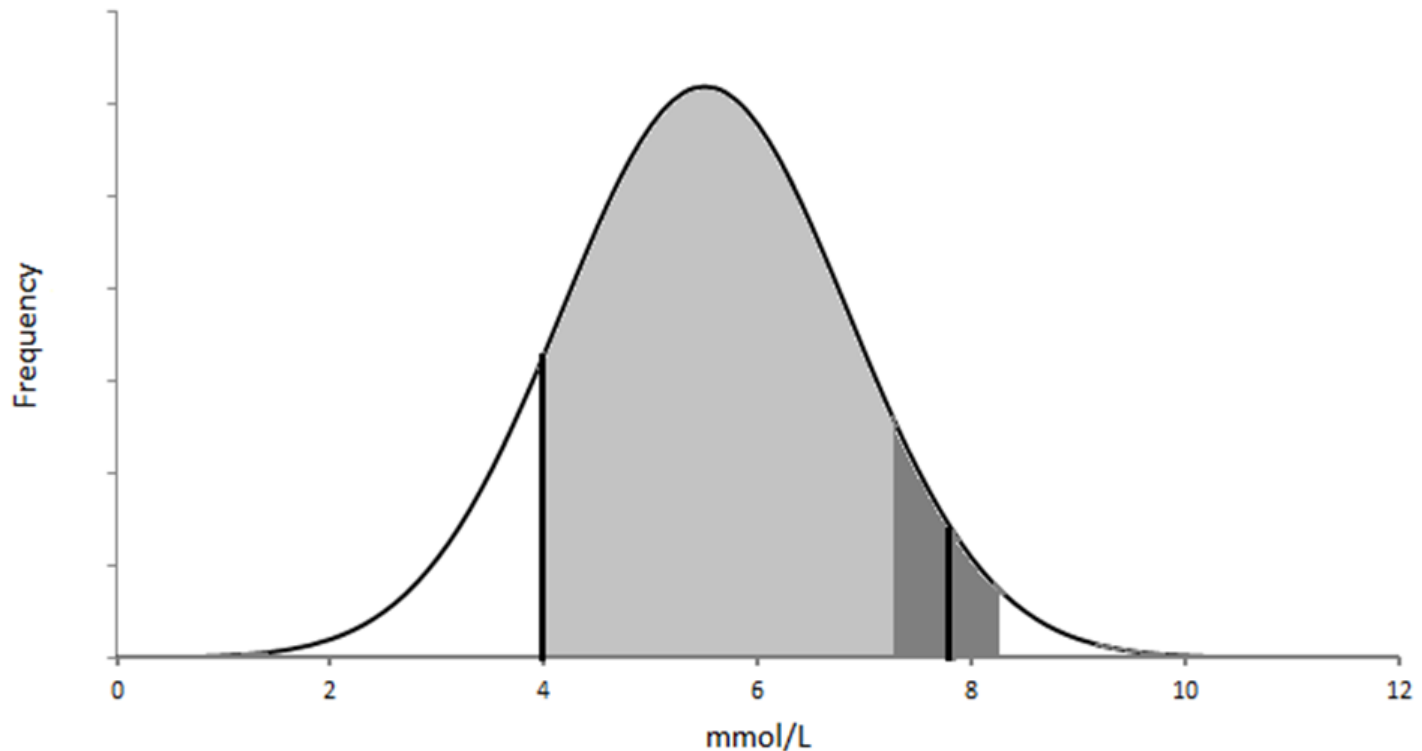
These assumptions are not true for infectious disease serology

# Data are Normally Distributed



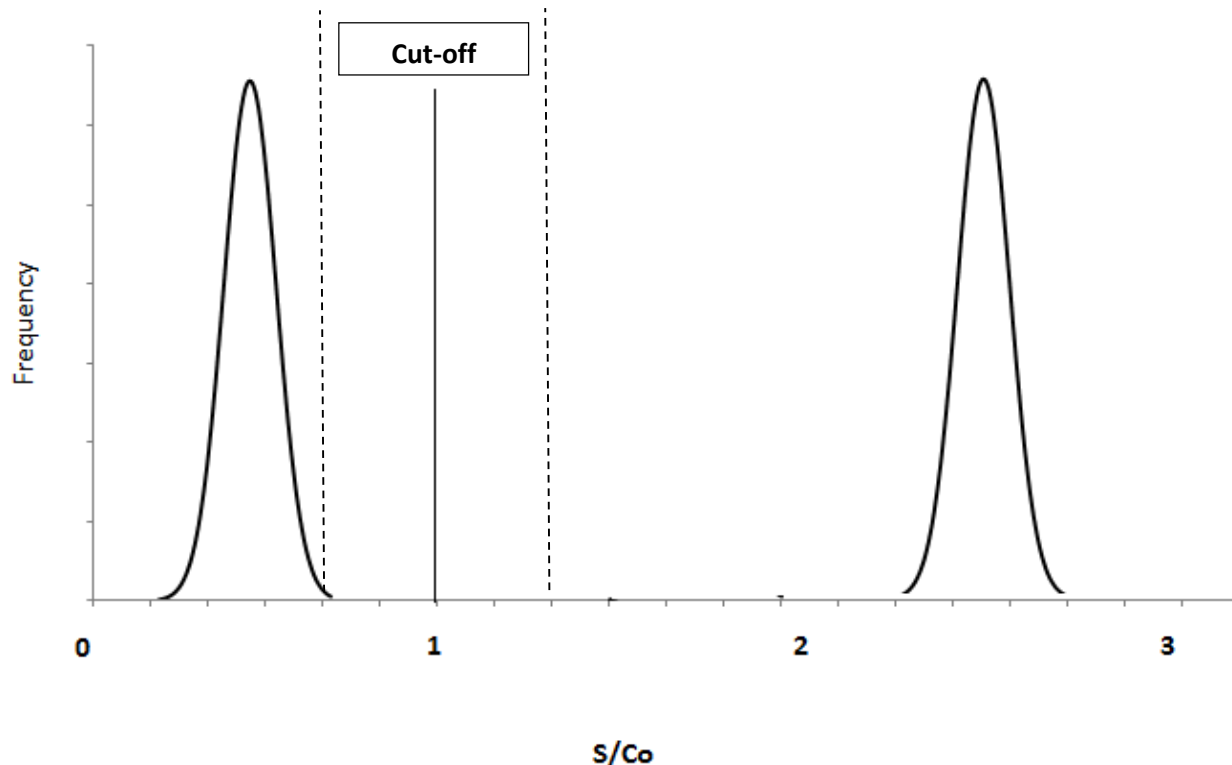
# QC Commutability

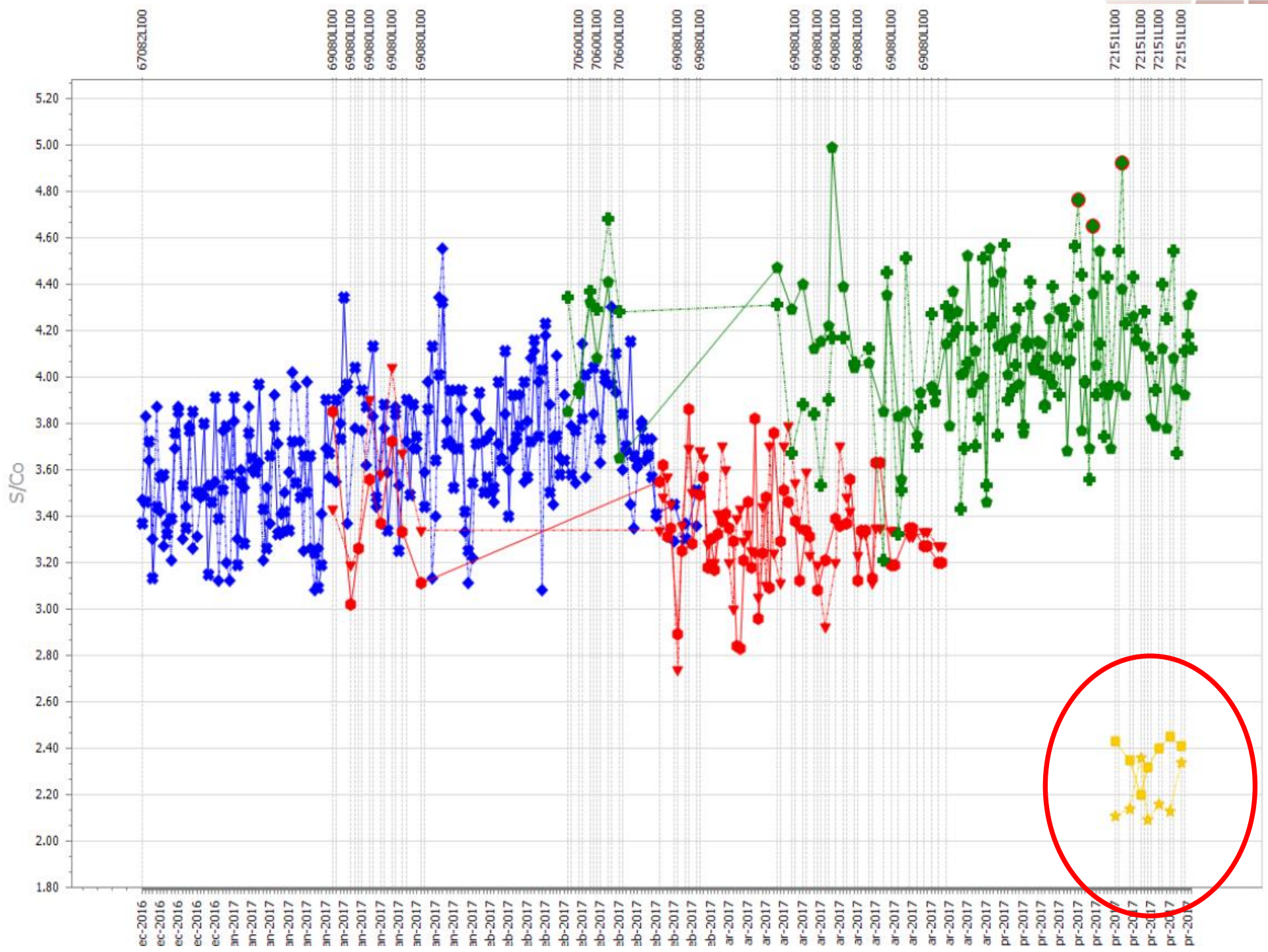
- Assuming QC commutability, percentage misinterpretation of chemistry results can be determined



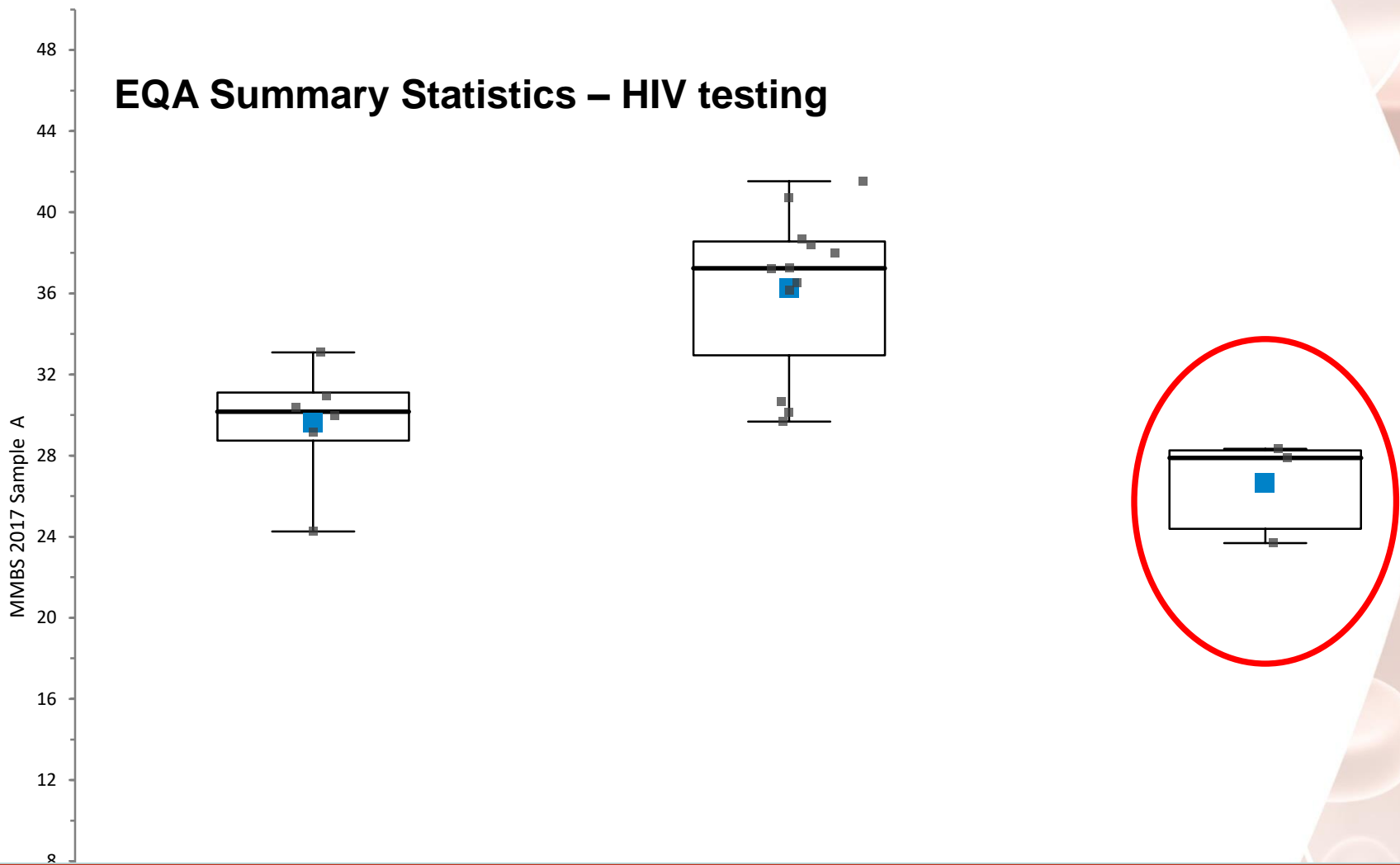
# QC Commutability

- However, in serology, very few results will be misinterpreted
- How do you estimate TE<sub>a</sub> for serology?





## EQA Summary Statistics – HIV testing



**Point 10:**  
Commutability of QC samples and patient samples should not be assumed

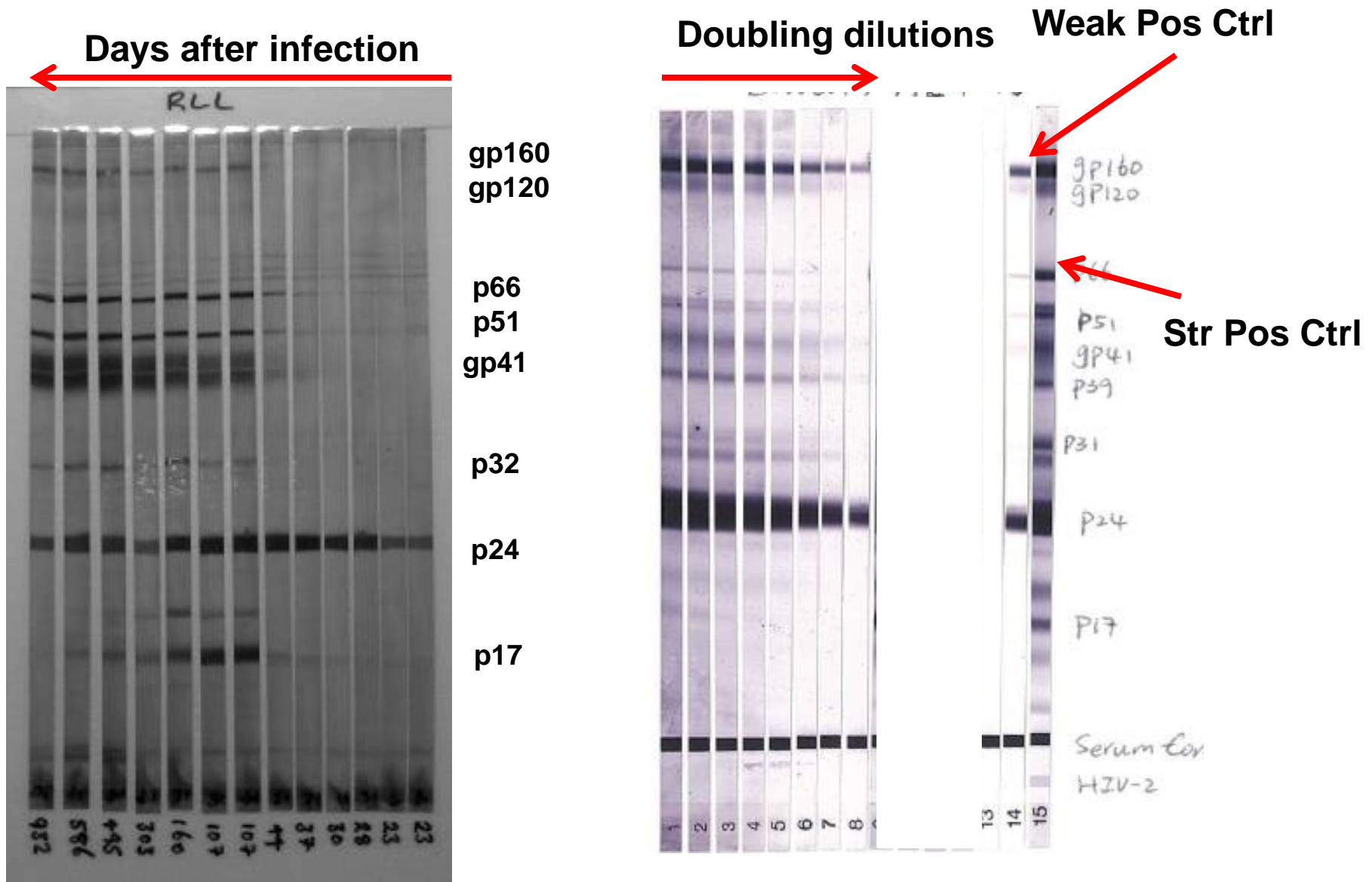
# Low levels QC mimics seroconversion

- Low positive, diluted QC mimics a seroconverter
- Diluted samples  $\neq$  early infection
- Antibodies ramp up very quickly
- Antibody profiles different to early infection (seroconversion)

Point 11:

Using a diluted sample to mimic early infection is a flawed concept

# WB Seroconversion vs Dilution

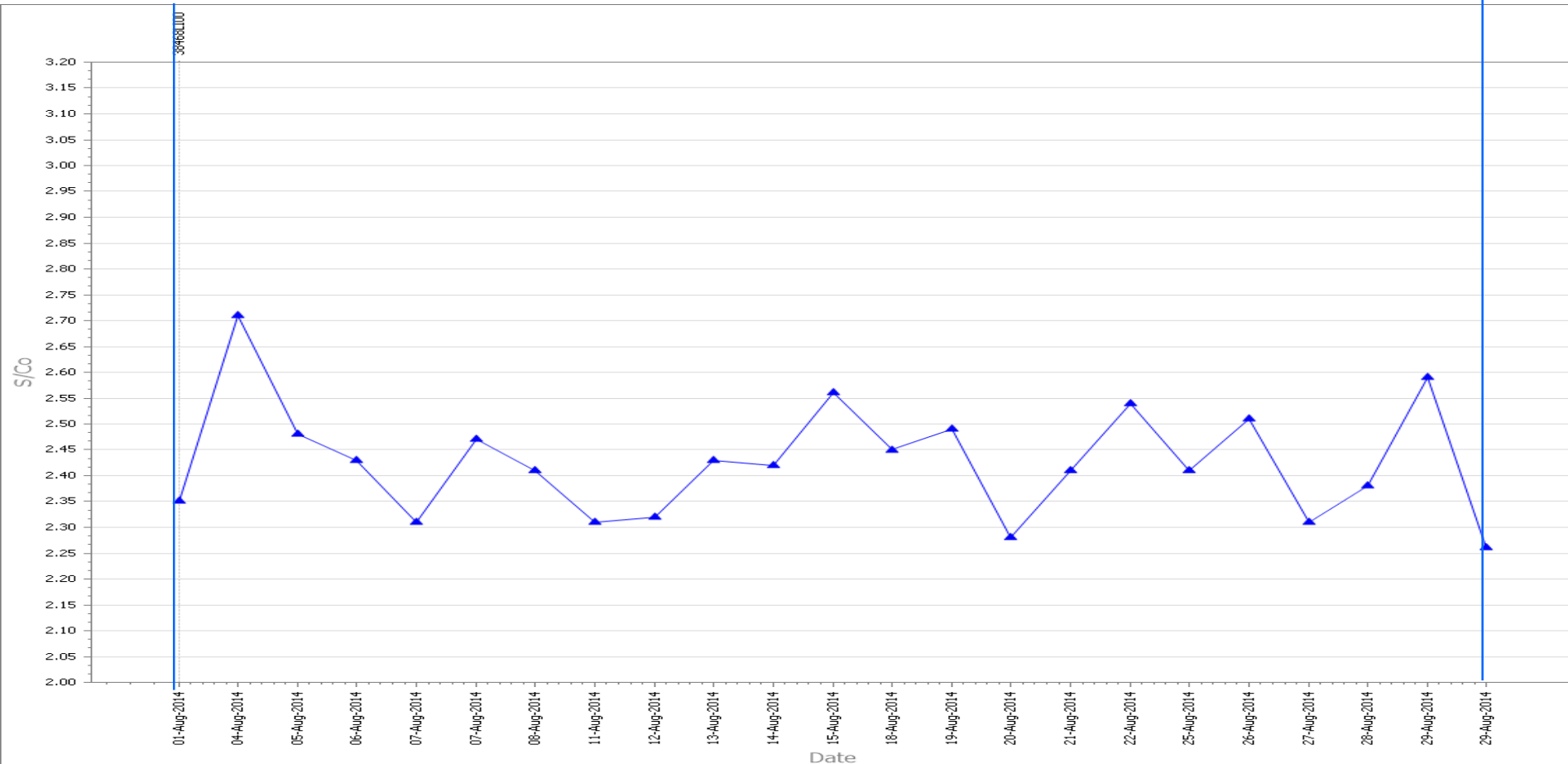




# Data are Representative

August

Sept



Laboratory:50

From:01-Aug-2014 to 31-Aug-2014

Assays:Abbott ARCHITECT Anti-HCV CMIA;

Analyte:anti-HCV

EQC:QConnect Blue

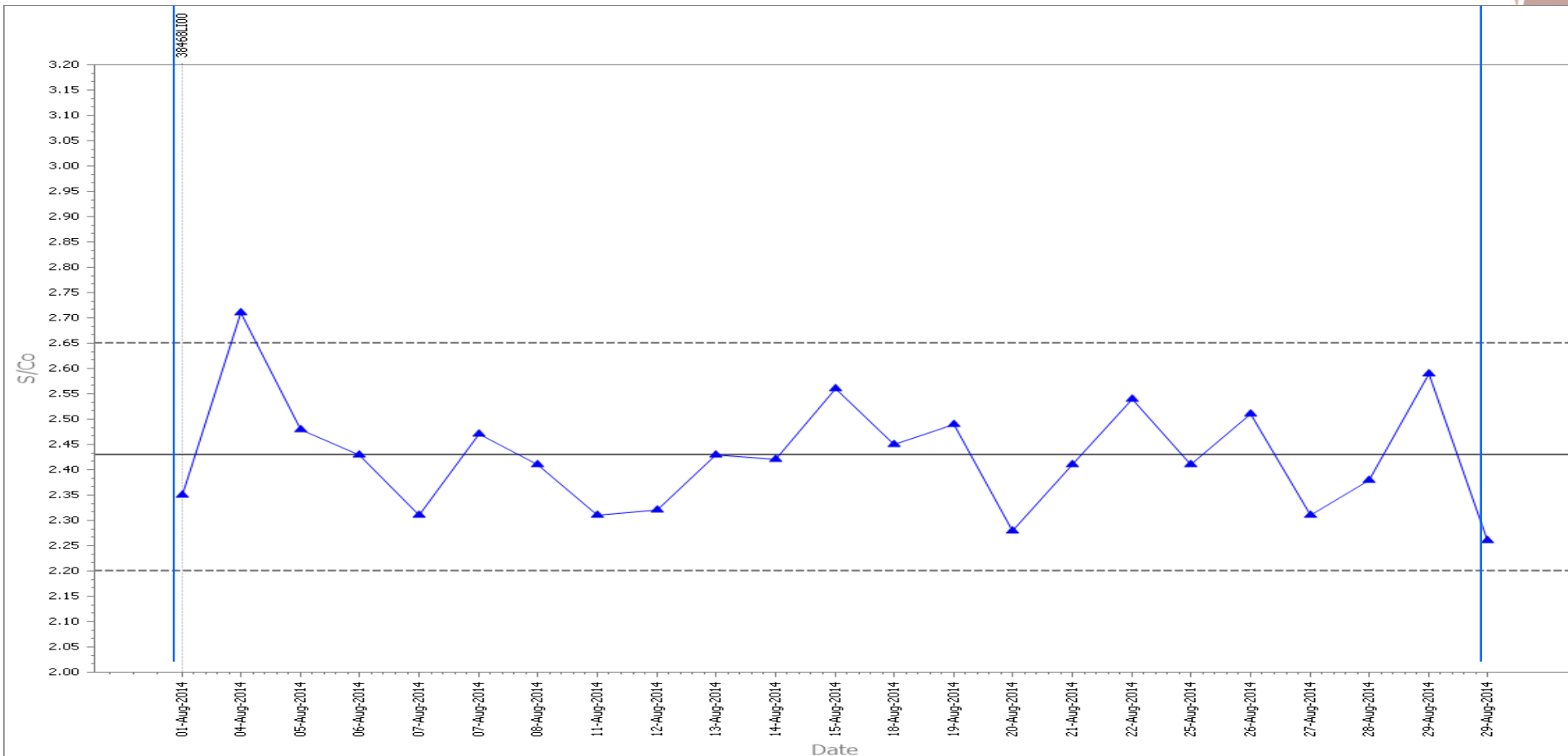
EQC Lot Number:413301;

Note: Custom Y Axis range used.

# Data are representative

Sept

August



Laboratory: 50    From: 01-Aug-2014 to 31-Aug-2014    Assays: Abbott ARCHITECT Anti-HCV CMIA;    Analyte: anti-HCV    EQC: QConnect Blue    EQC Lot Number: 413301;

— Your Laboratory - Visible Data; Mean=2.43    - - - - Laboratory Range: 2.20 to 2.65

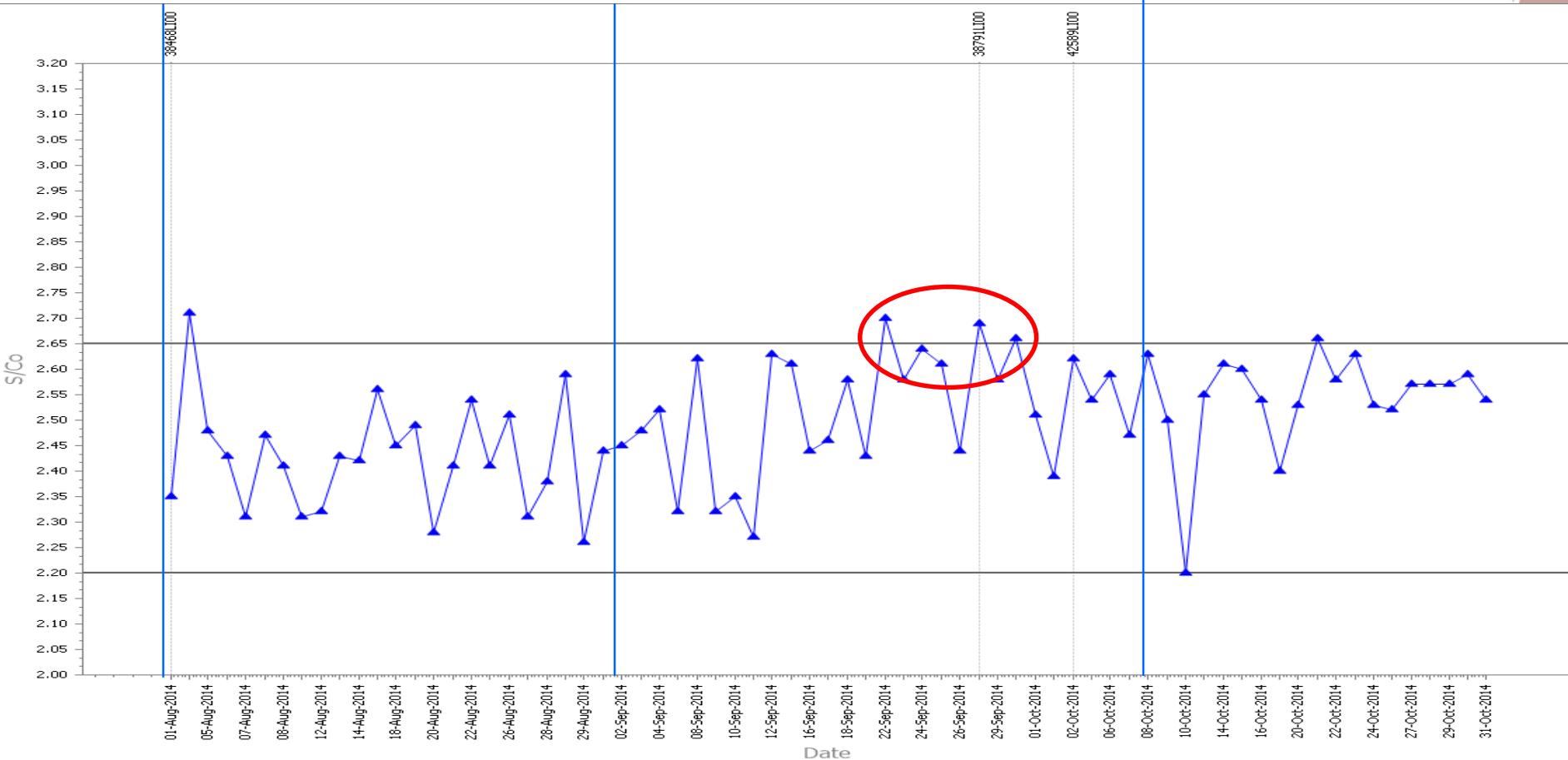
Note: Custom Y Axis range used.

# Data are representative

Aug

Sept

Oct



Laboratory:50 From:01-Aug-2014 to 31-Oct-2014

Assays:Abbott ARCHITECT Anti-HCV CMIA;

Analyte:anti-HCV

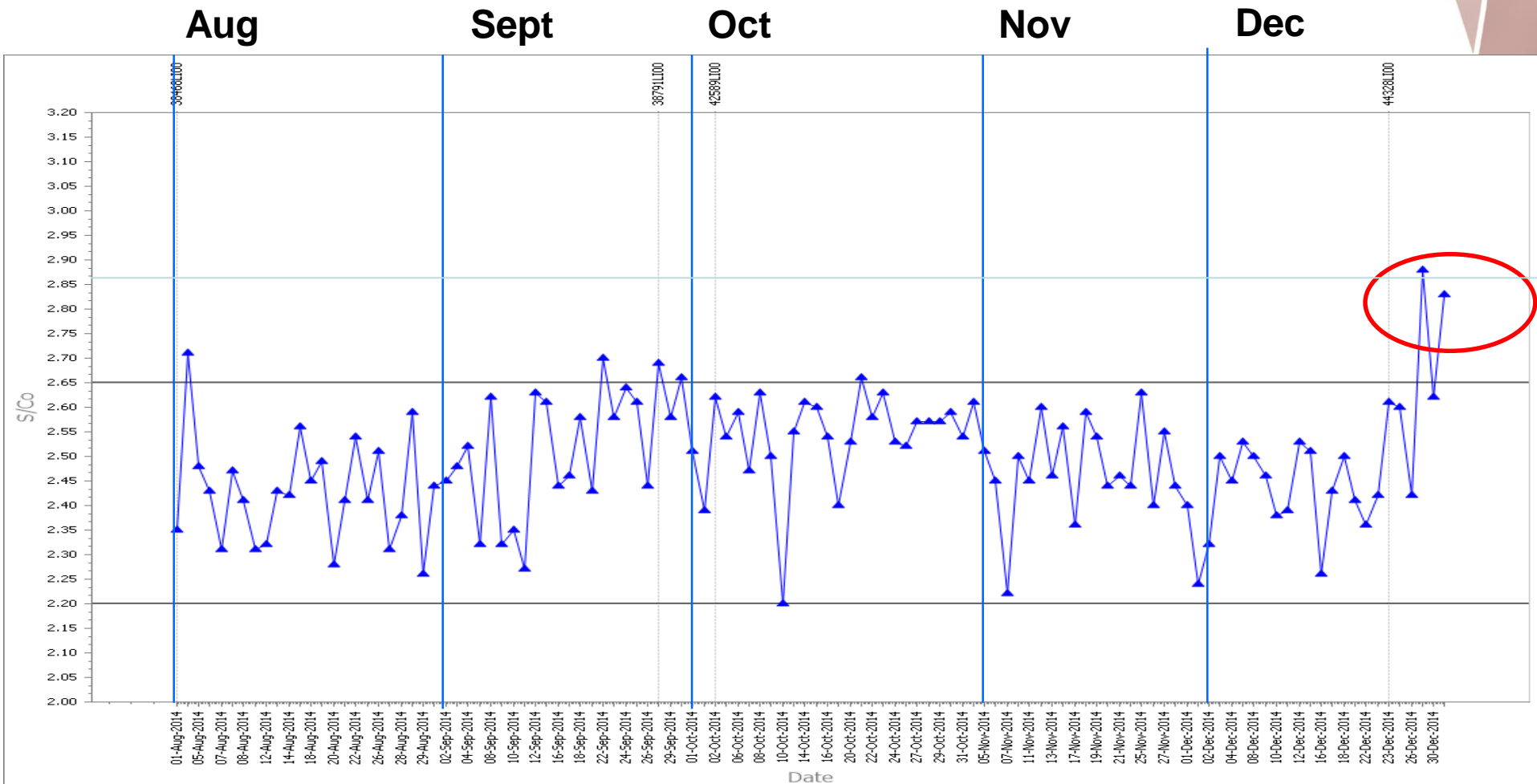
EQC:QConnect Blue

EQC Lot Number:413301;

Laboratory Range:2.20 to 2.65

Note: Custom Y Axis range used.

# Data are representative



Laboratory:50 From:01-Aug-2014 to 31-Dec-2014

Assays:Abbott ARCHITECT Anti-HCV CMIA;

Analyte:anti-HCV

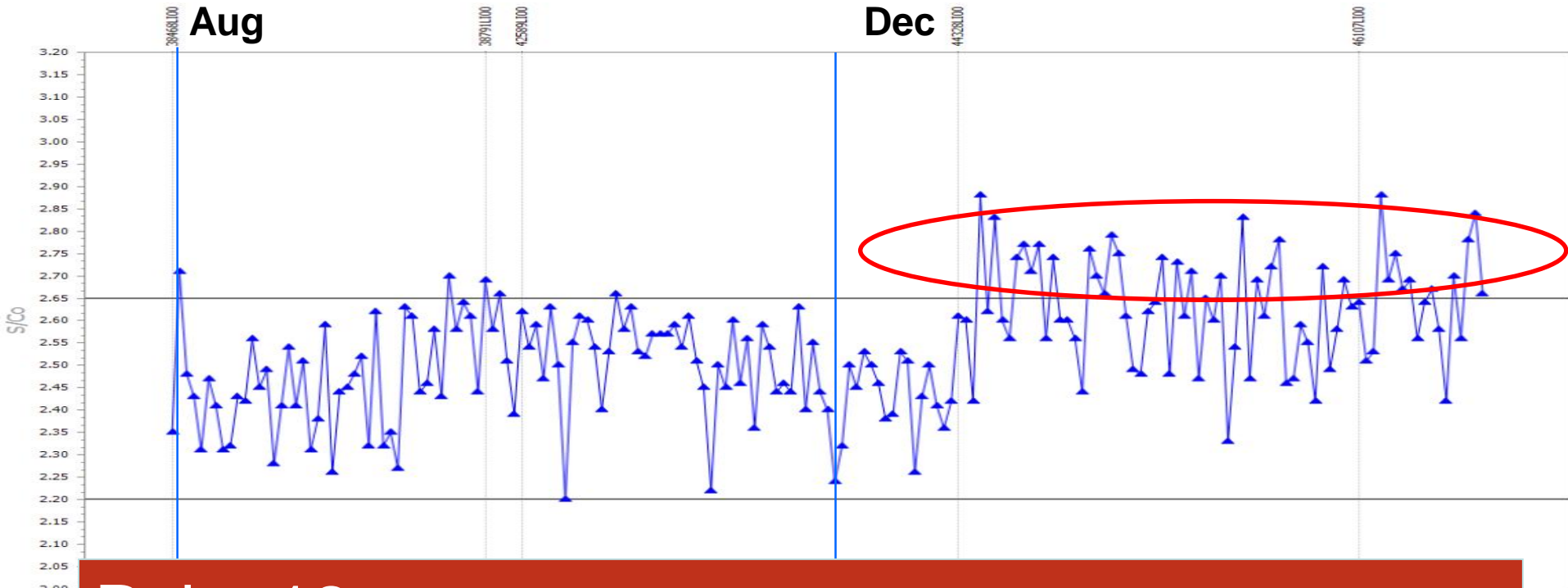
EQC:QConnect Blue

EQC Lot Number:413301;

Laboratory Range:2.20 to 2.65

Note: Custom Y Axis range used.

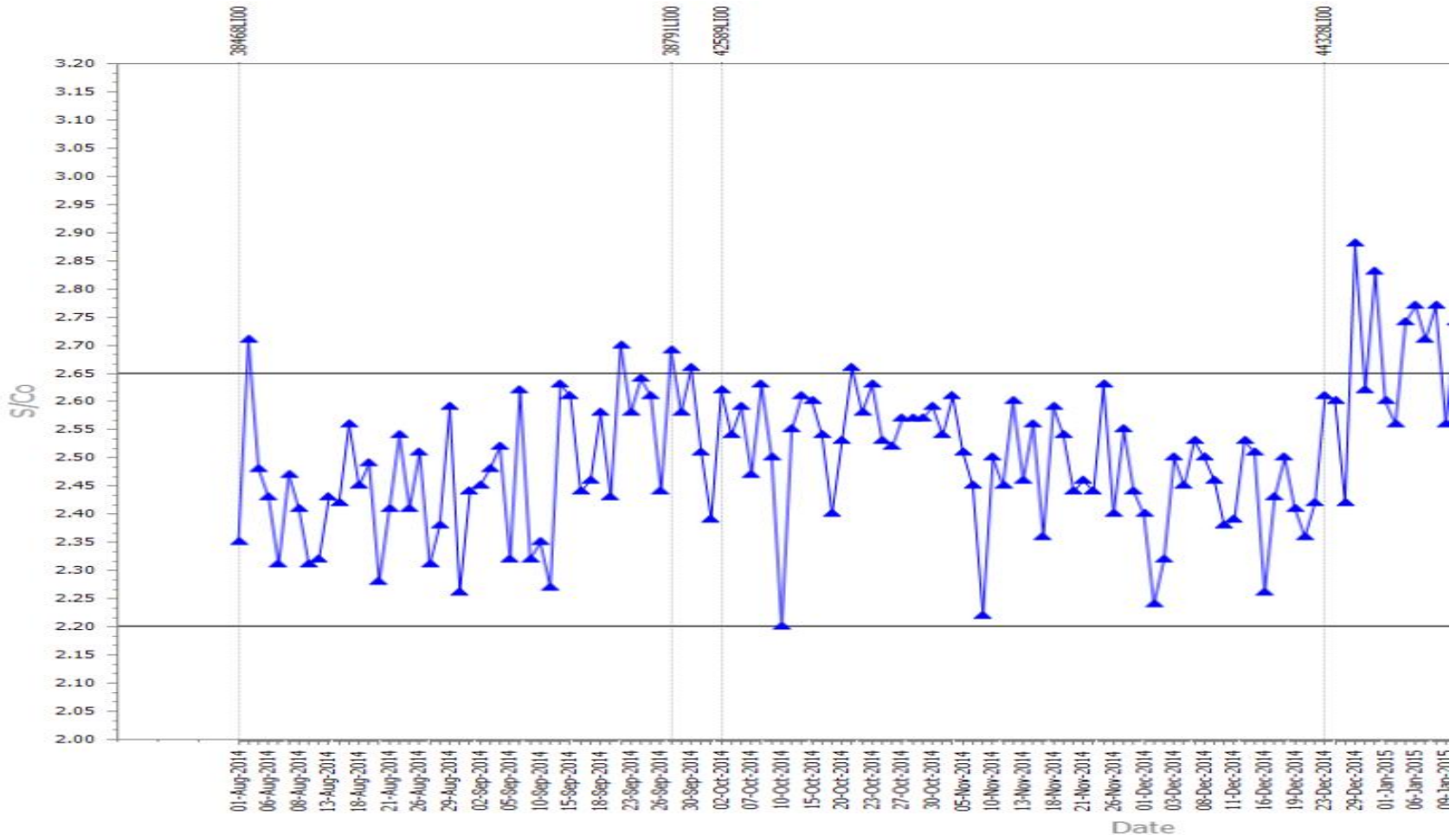
# Data are representative



Point 12:

Reagent lot variation is the major contributor to normal variation of serology assays

# What would you do?



Laboratory:50 From:01-Aug-2014 to 01-Apr-2015 Assays:Abbott ARCHITECT Anti-HCV CMIA; Analyte:anti-HCV EQC:QConn  
 Laboratory Range:2.20 to 2.65  
 Note: Custom YAxis range used.

# Possible actions when reagent lot changes

A - Ignore

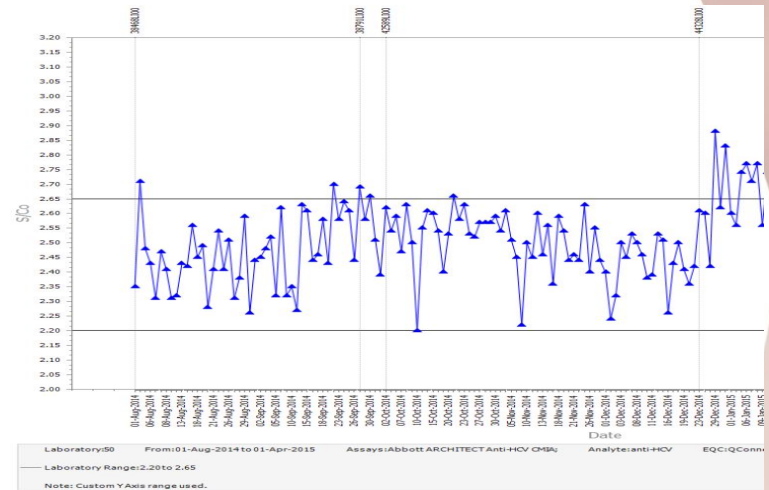
B - Re-test QC

C - Re-calibrate instrument

D - Re-set limits

E – Contact manufacturer

F – Reject reagent lot



# What do you do?

A - Ignore

B - Re-test QC

C - Re-calibrate  
instrument

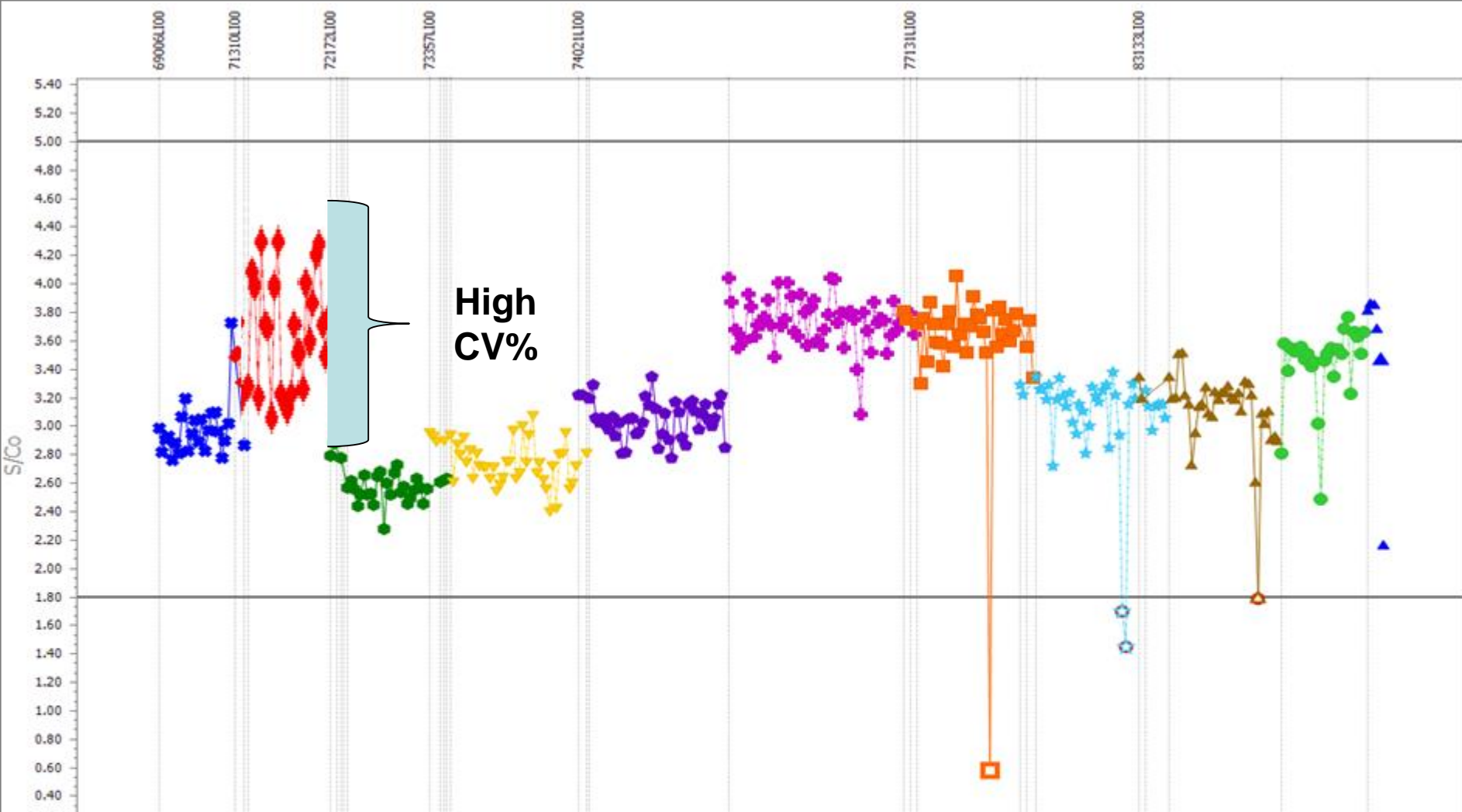
D - Re-set limits

E - Contact manufacturer

F - Reject reagent lot

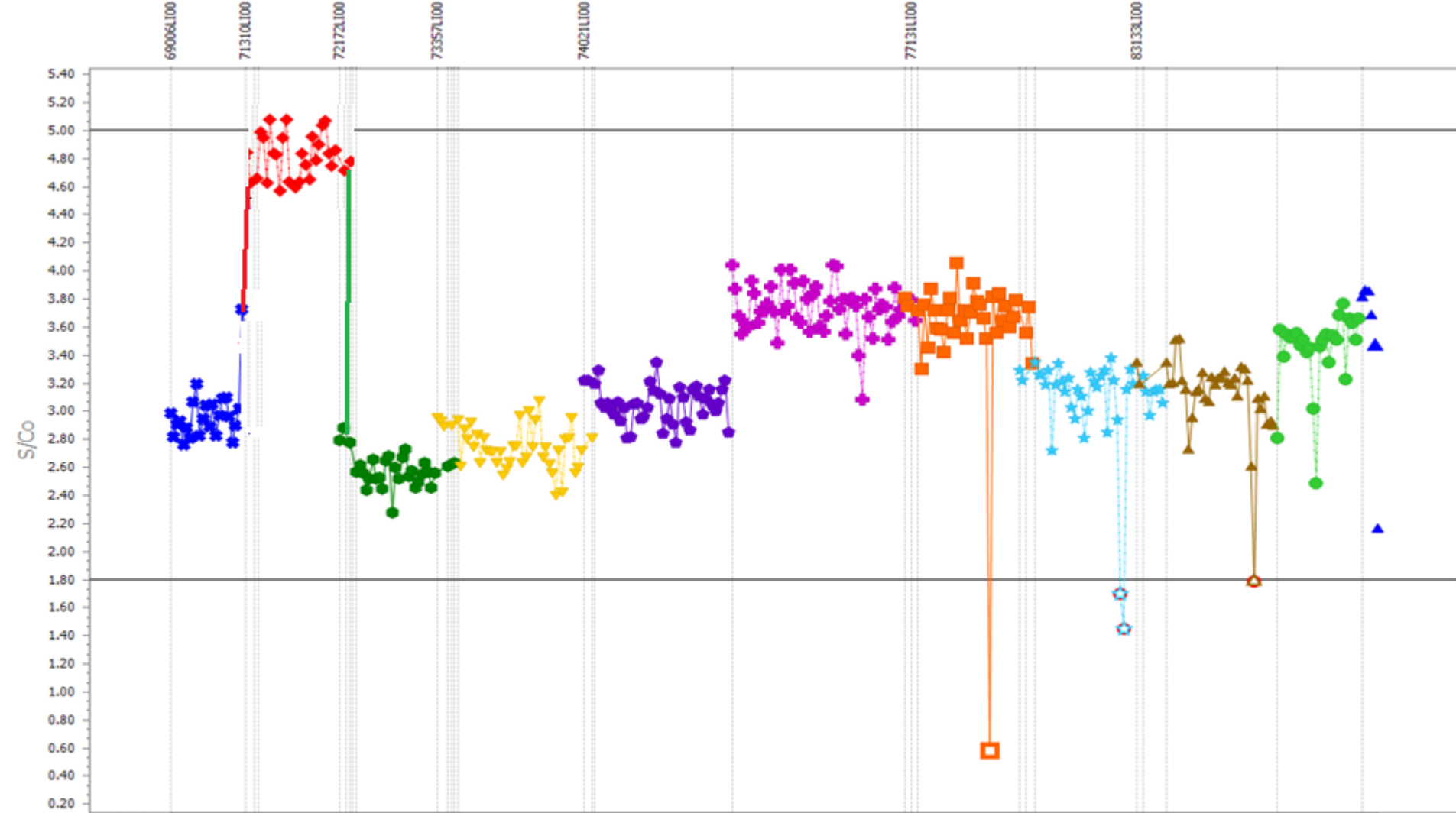
Start the presentation to see live content. Still no live content? Install the app or get help at [PollEv.com/app](https://PollEv.com/app)





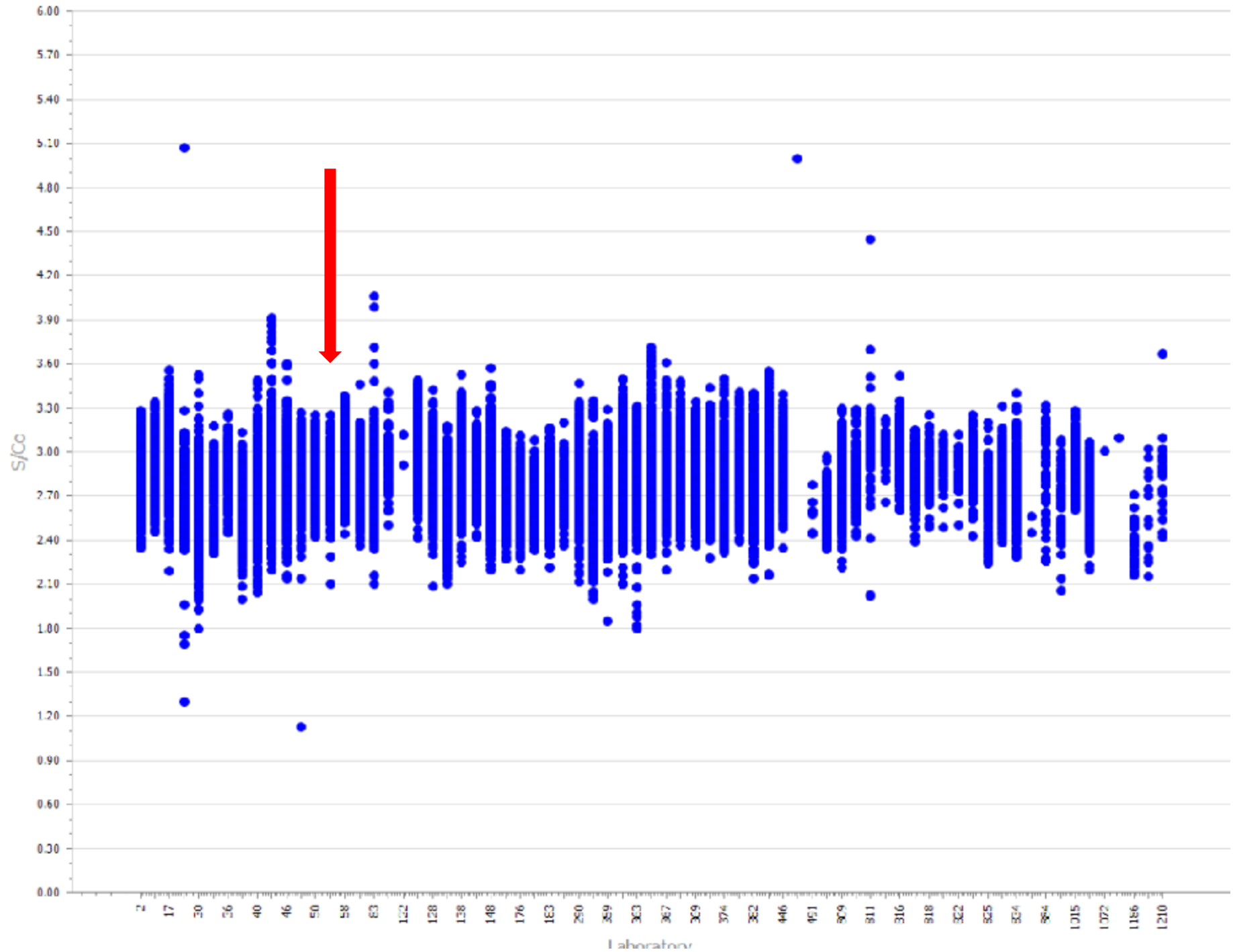
## Point 13:

Usually there are insufficient runs to re-set limits before reagent lot is exhausted



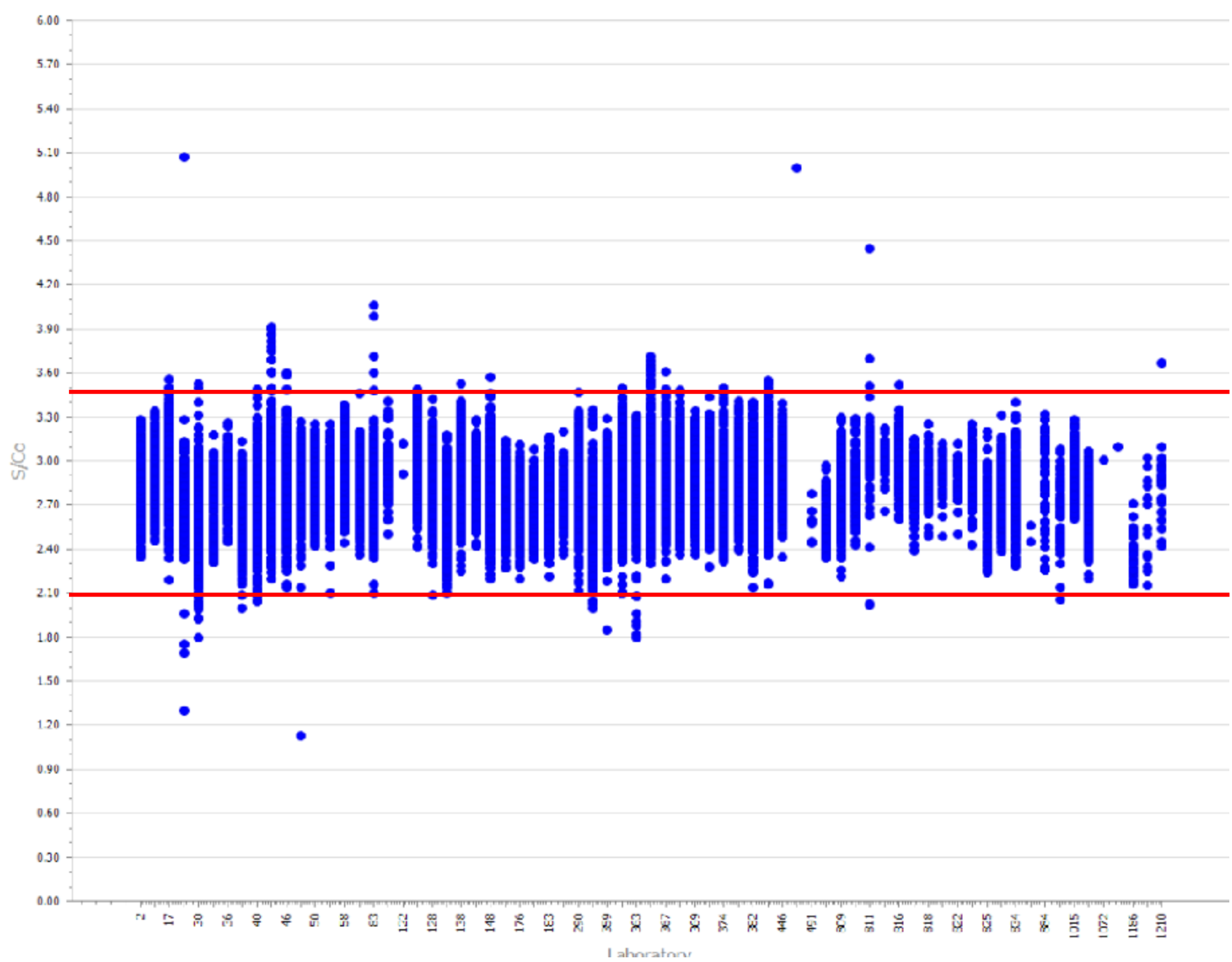
Point 14:

By re-setting limits, you are accepting that the variation is normal



# Normal Assay Variation

- One years results for Abbott Architect HCV
- 71 laboratories
- 94 instruments
- 77 reagent lot numbers
- 7 QC lot numbers
- 18,234 results
- CV% range: 3.74 to 10.86%
- CV% average 7.24%
- NRL Range: 2.1 to 3.5 (S/Co)



# Setting Acceptance Limits for Serology

- 20 data points is not representative
- Should include all normal variation
- QConnect limits uses historical data
- Laboratories can use their own data over time
- Determine acceptable range and acceptable CV%

# Quality Control for Viral Load Testing

- Three processes
  - Extraction
  - Amplification
  - Detection
- Possibly use different reagents and instruments for each process

# Differences between Clinical Chemistry and Viral Load Testing

Clinical Chemistry	Serology	Viral Load
Linear	Non-linear	Linear
Inert analyte	Functional biological analyte	Biological, not functional
Quantitative	Qualitative	Quantitative
Adjust for bias	No adjustment for bias	No adjustment for bias
Lower level of regulation	Highly regulated	Highly regulated
Several medical decision points	Single decision point	Several medical decision points
Adjust for lots variation	No adjustment for lot variation	No adjustment for lot variation
International standards	Poor or no standards	International standards
Certified reference methods	No CRMs	Certified reference methods
Available in a pure form	Different forms	Some mutations or variation
Single target	Multiple and varying targets	Single target
TEa	?? TEa	?? TEa



# Hepatitis B DNA

Assay	No Labs	No QC Lots	n	Mean (log <sup>10</sup> )	SD	CV(%)
Abbott RealTime HBV (0.5 mL)	2	3	186	2.4	0.12	4.99
Abbott RealTime HBV (0.2 mL)	4	3	385	2.5	0.23	8.88
Roche COBAS AmpliPrep/TaqMan HBV Test v2	16	3	604	2.3	0.10	4.40
Roche COBAS 4800 HBV	1	2	73	2.3	0.08	3.52
Roche cobas HBV Quantitative (6800/8800)	6	2	208	2.3	0.16	6.83

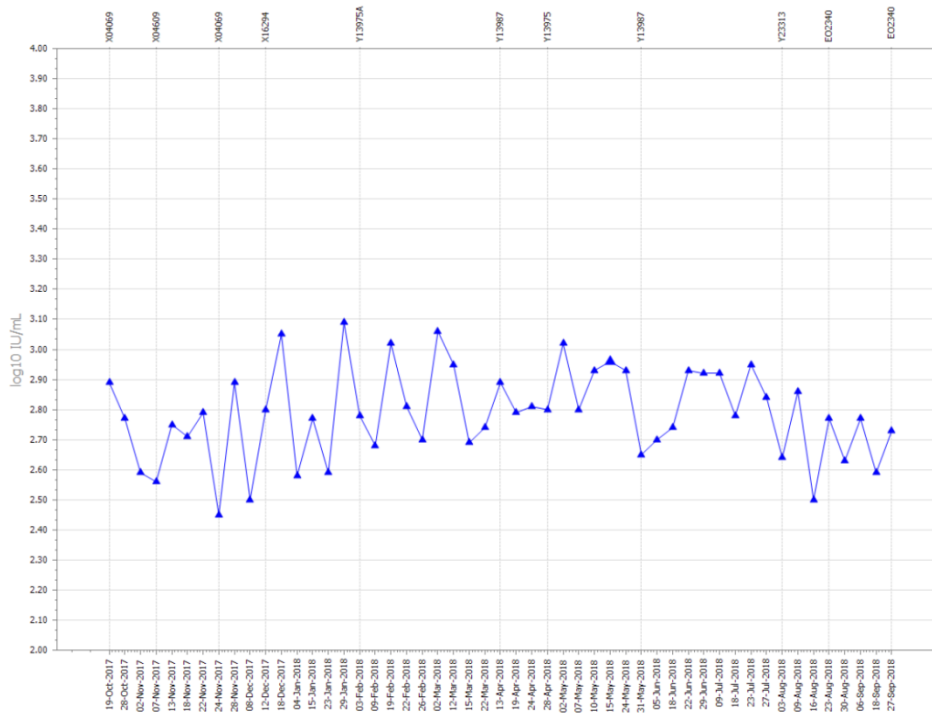
# Hepatitis C RNA

Assay	No Labs	No QC Lots	n	Mean (log <sup>10</sup> )	SD	CV(%)
Abbott RealTime HCV (0.2 mL)	2	2	79	2.3	0.25	10.56
Abbott RealTime HCV (0.5 mL)	4	2	196	2.4	0.24	9.52
Roche COBAS AmpliPrep/COBAS TaqMan HCV v2	16	2	724	2.7	0.25	8.94
Roche cobas 4800 HCV	2	2	92	2.5	0.18	7.24
Roche cobas HCV Quantitative (6800/8800)	6	2	299	2.4	0.19	7.78

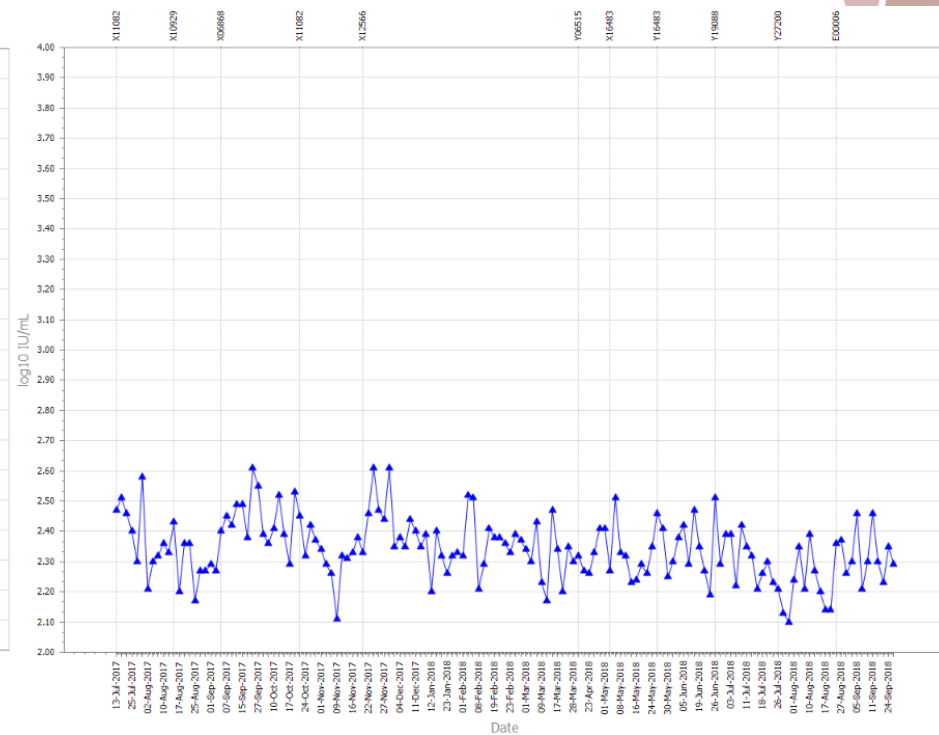
# HIV RNA

Assay	No Labs	No QC Lots	n	Mean (log <sup>10</sup> )	SD	CV(%)
Abbott RealTime HIV-1 (0.2 mL)	1	2	22	2.3	0.16	6.85
Abbott RealTime HIV-1 (0.6 mL)	4	2	217	2.2	0.16	6.91
Cepheid Xpert HIV-1 Viral Load	1	2	13	2.1	0.10	4.57
Roche cobas 4800 HIV-1	1	2	33	2.2	0.13	5.63
Roche cobas HIV-1 Quantitative (6800/8800)	3	2	139	2.2	0.14	6.28
Roche COBAS AmpliPrep/COBAS TaqMan HIV-1 Test v2	16	2	902	2.4	0.15	6.10

# LJ Chart for Selected Laboratory

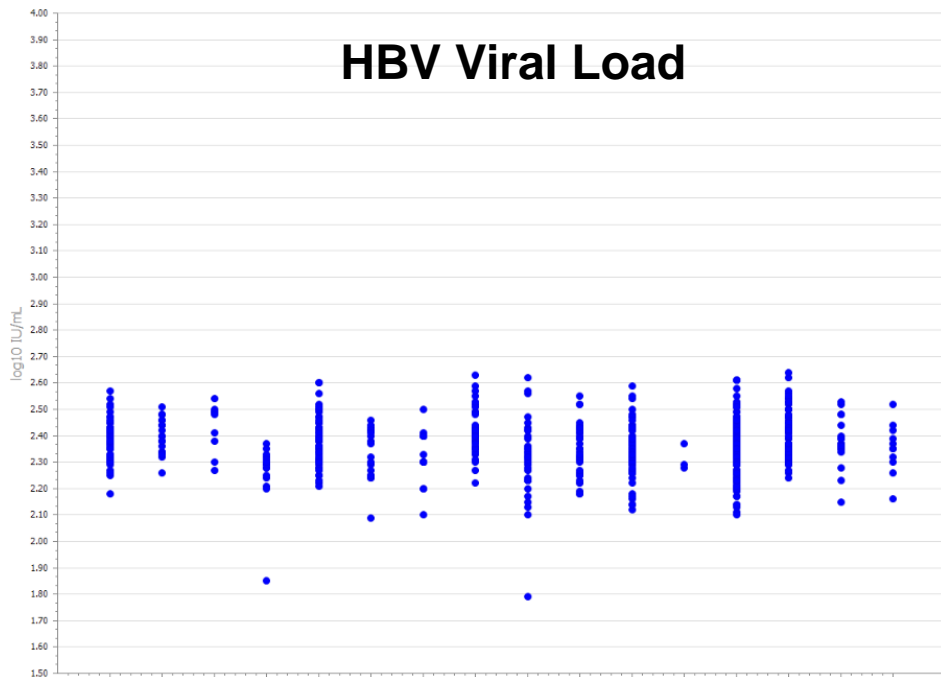


**HCV Viral Load**

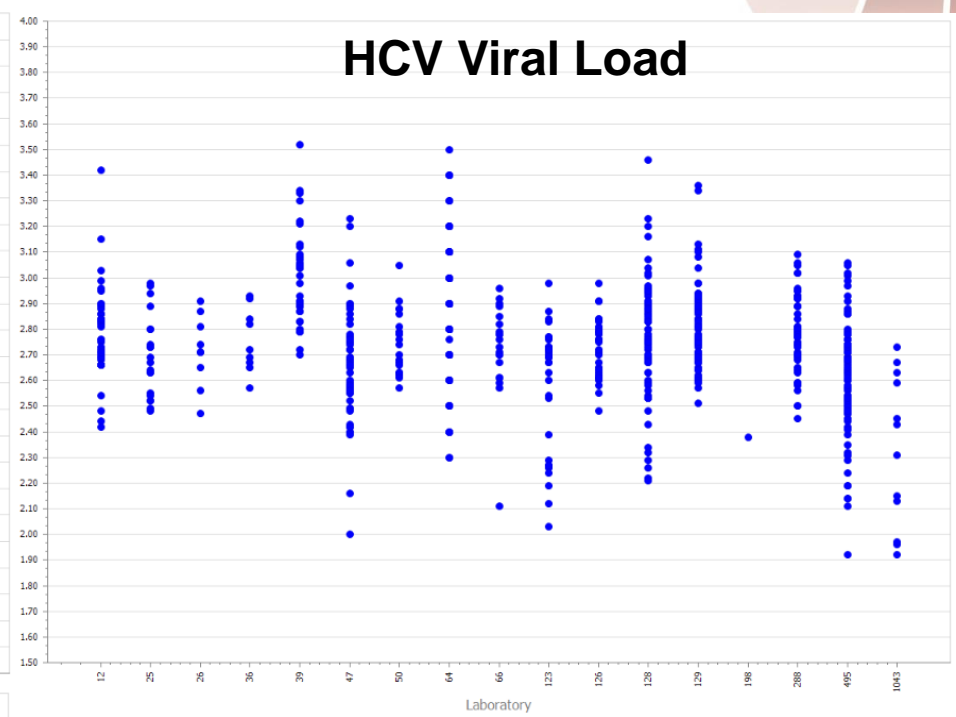


**HBV Viral Load**

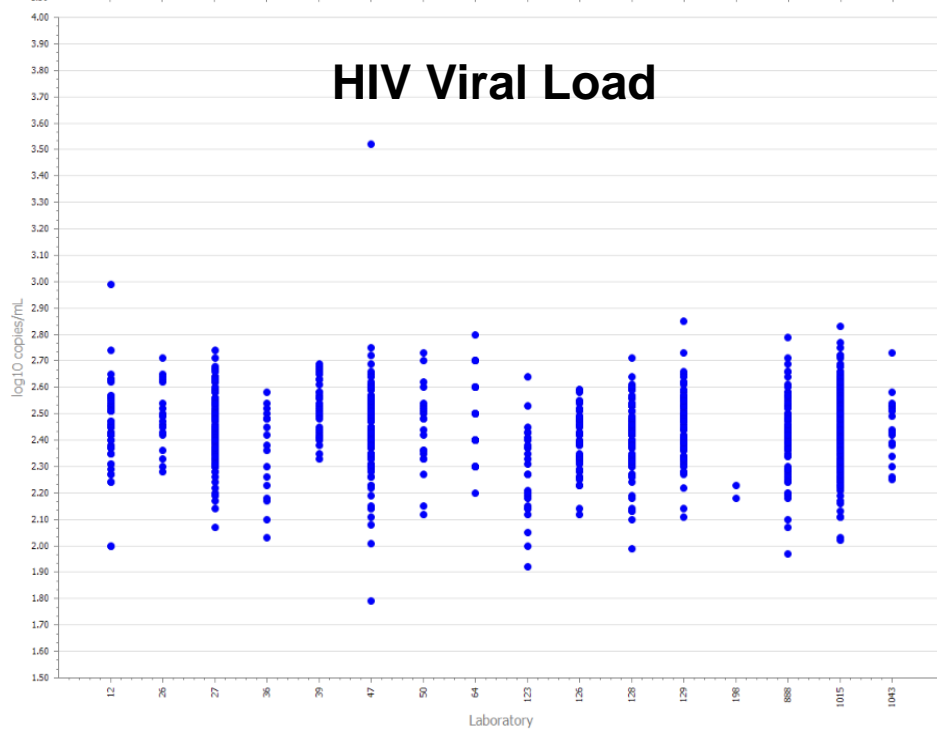
## HBV Viral Load



## HCV Viral Load



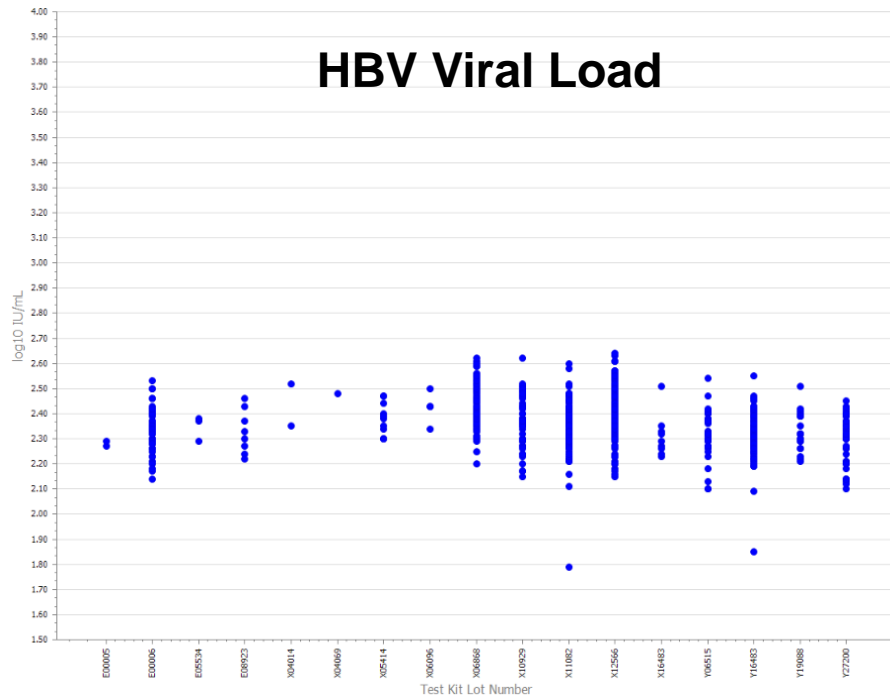
## HIV Viral Load



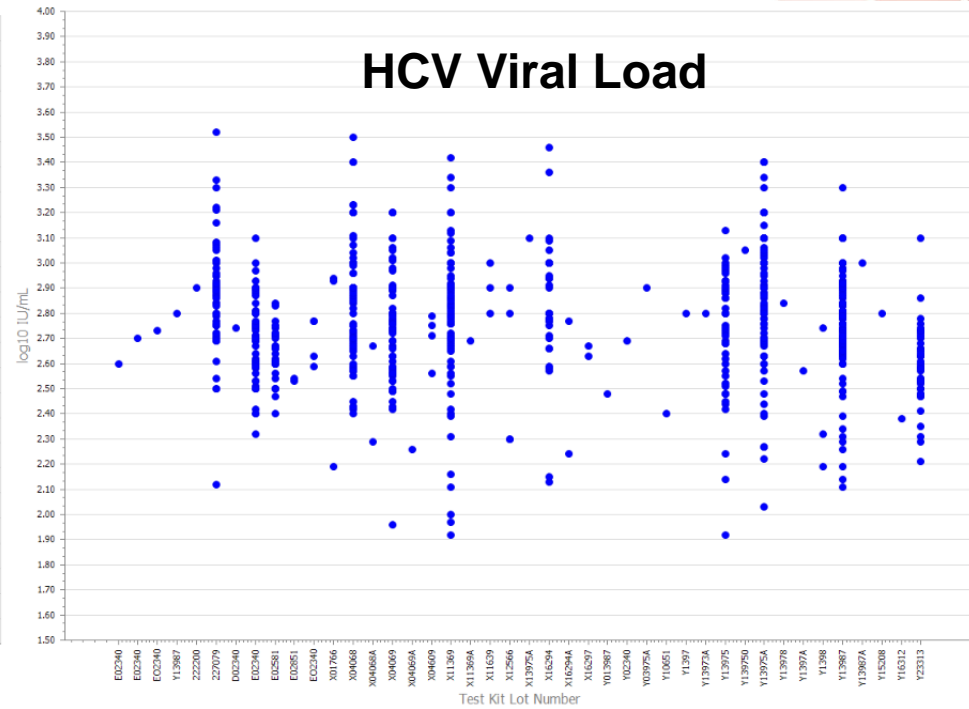
**Roche COBAS AmpliPrep/COBAS  
TaqMan Assays**

**(Laboratory Results)**

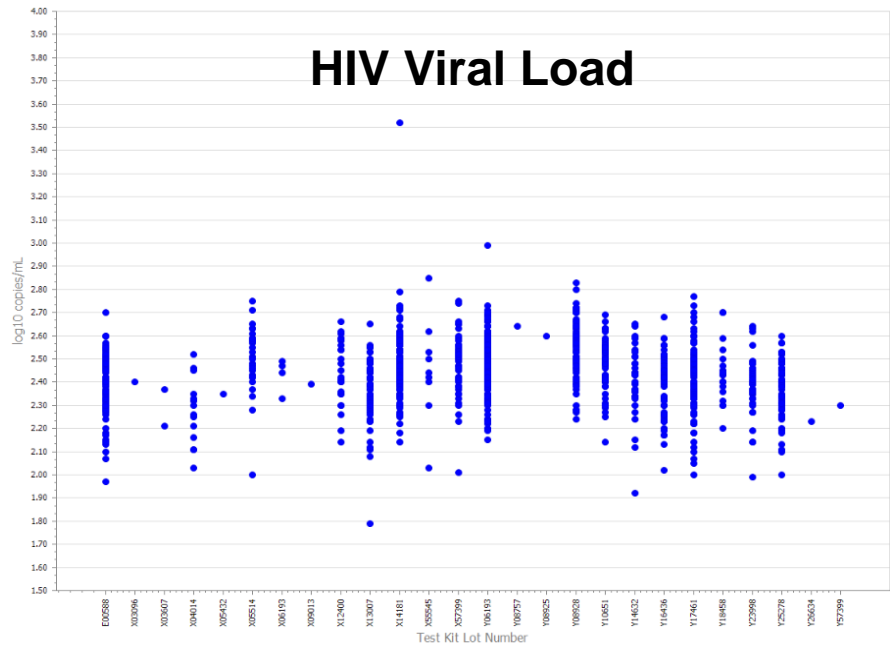
## HBV Viral Load



## HCV Viral Load



## HIV Viral Load



**Roche COBAS AmpliPrep/COBAS  
TaqMan Assays**

**(Reagent Lot Number Results)**

# Quality Control for Viral Load Testing

- Similarities with clinical chemistry and serology
- Dose response is linear
- Reagent lot variation not as great as serology
- Standardisation between assays is good
- Clinically significant change is acknowledged
- Precision is predictable

# Setting Acceptance Limits for Viral Load Testing

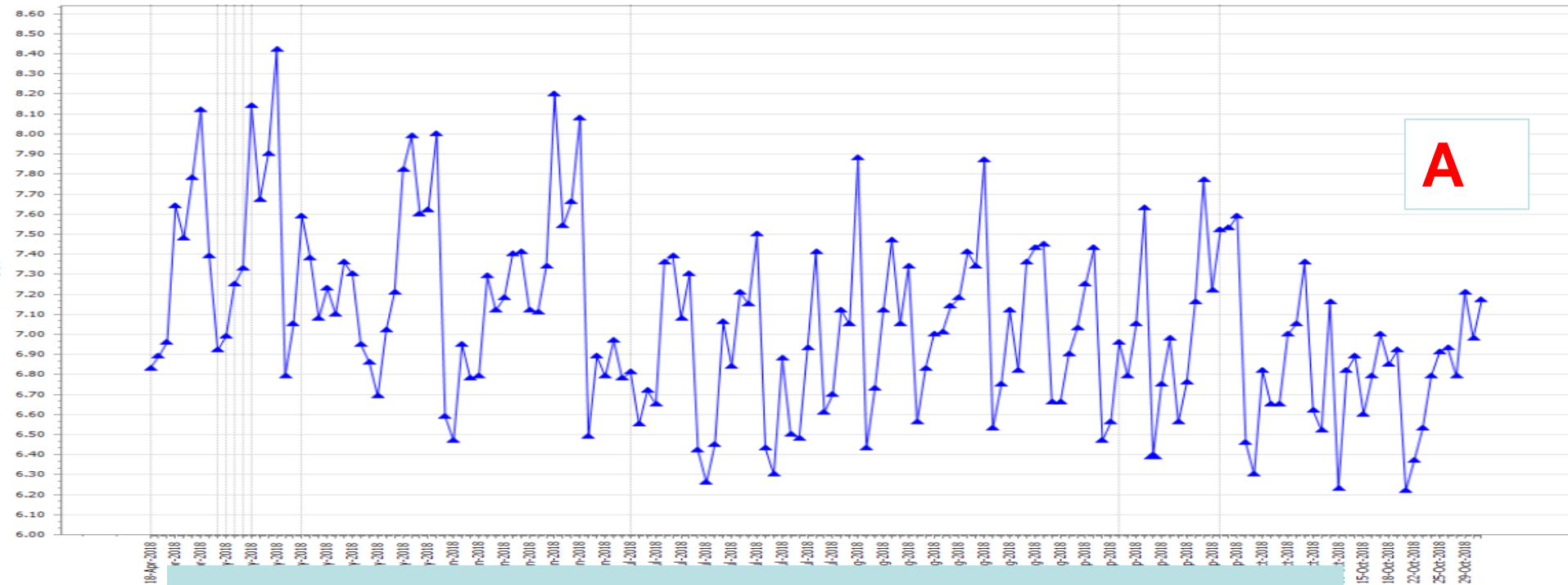
- Traditional Methods?
- QConnect Limits?
- Alternative based on:
  - clinical significant change (TEa)
  - expected variation (QC and EQA)
  - Assay specific or general rules
- Cannot assume VL is similar to chemistry or serology
- Requires investigation using true QC results; not modelling





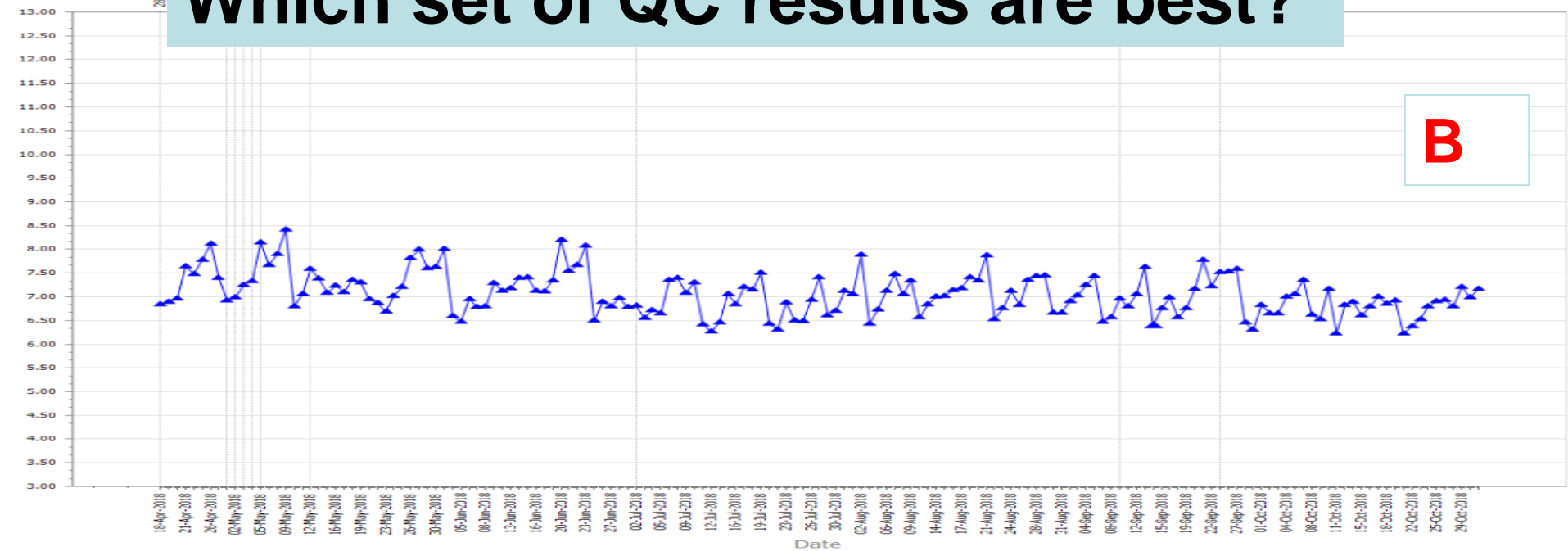
## Quiz Time

Let's have  
some fun!



**A**

**Which set of QC results are best?**



**B**

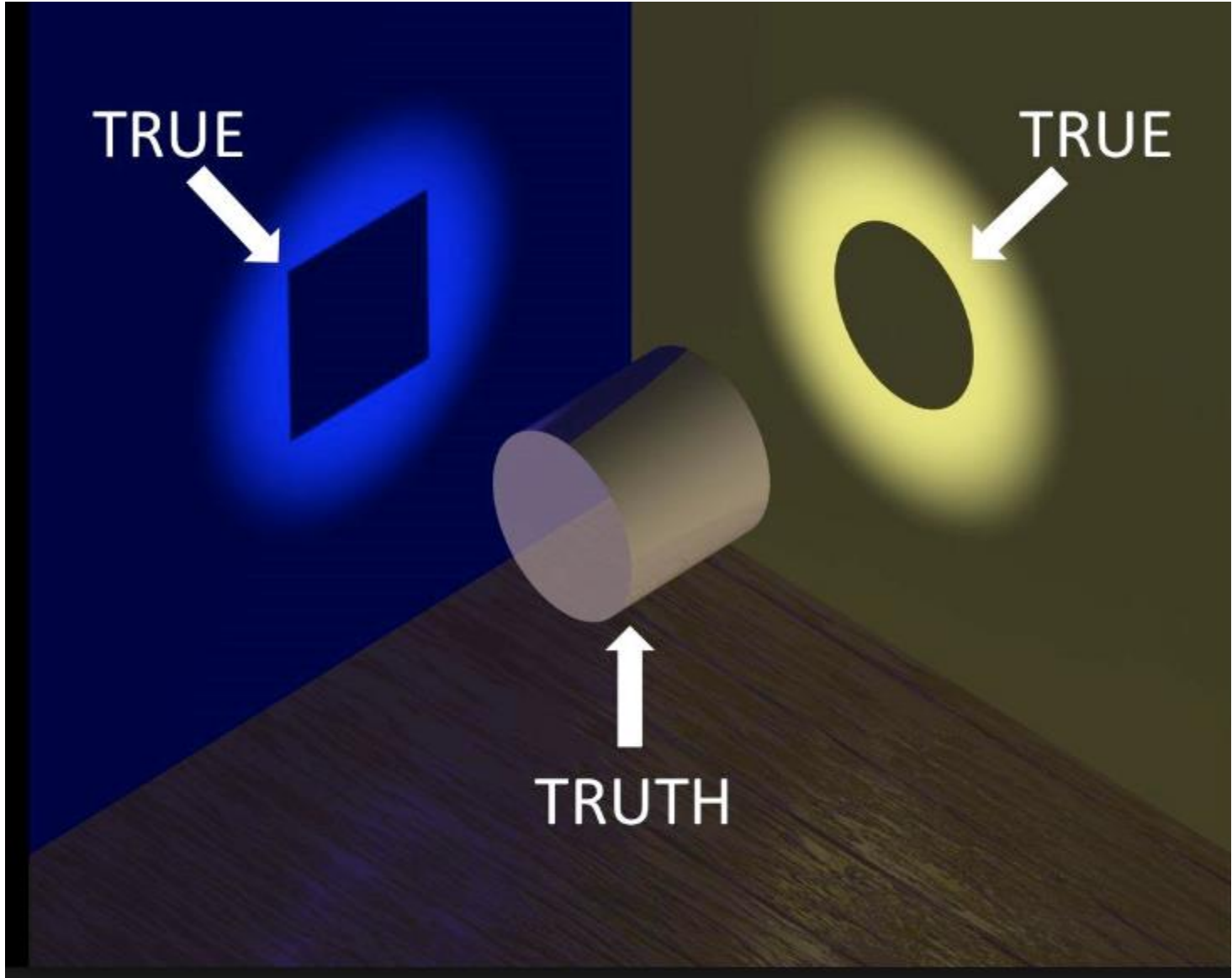
# Which set of QC results are bst?

Graph A

Graph B

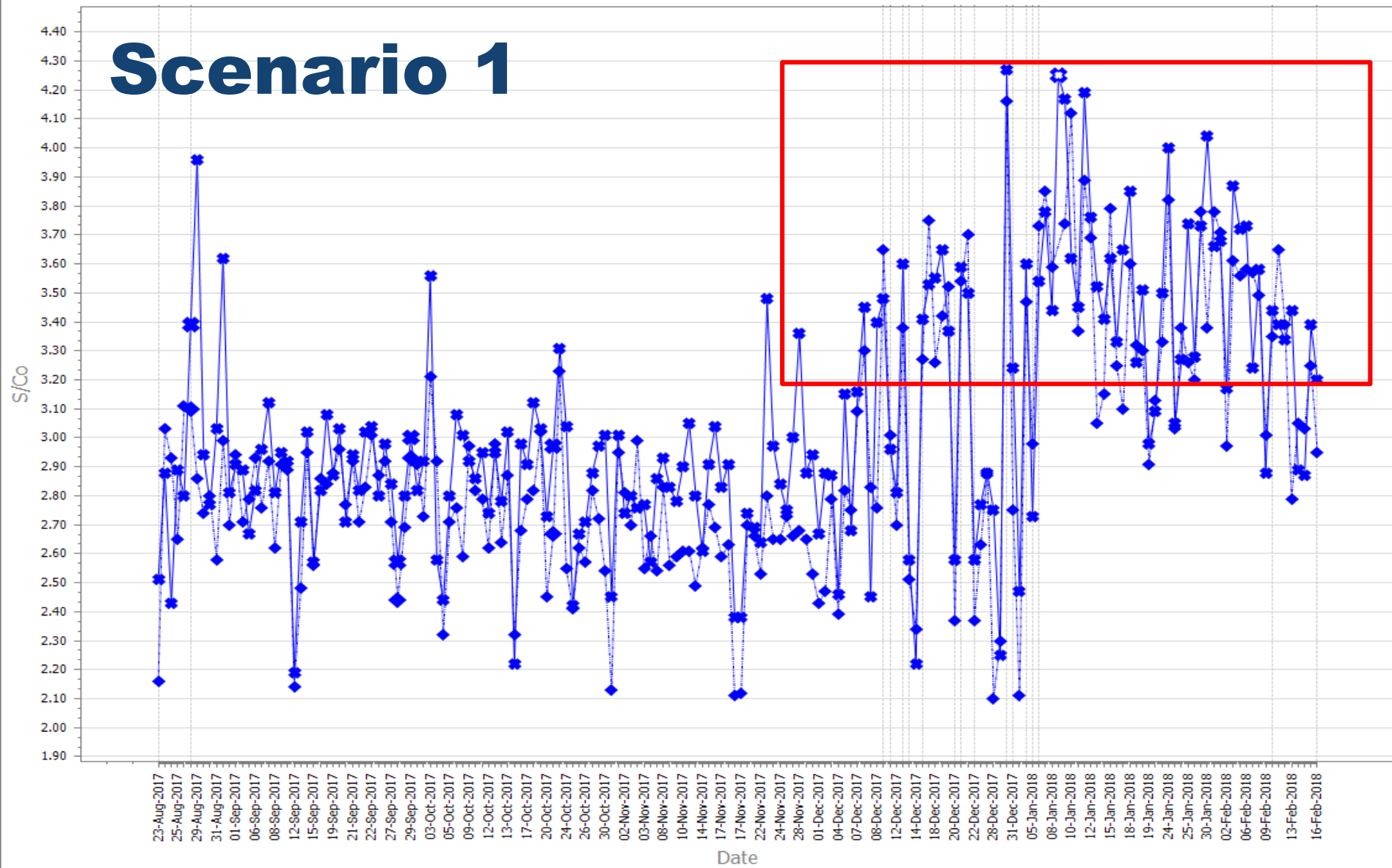
Graphs are  
the same

Both are  
bad



# Scenario 1

# Scenario 1



Laboratory:3 From:01-Feb-2017 to 20-Feb-2018 Assays:Abbott PRISM HIV Ag/Ab Combo ChLIA; Analyte:HIV p24 Ag/Anti-HIV EQC:QConnect Purple EQC Lot Number:DM17076;

▲ Subchannel A ▼ Subchannel B

Trend: Assay -●- Abbott PRISM HIV Ag/Ab Combo ChLIA/Subchannel A -◆- Abbott PRISM HIV Ag/Ab Combo ChLIA/Subchannel B

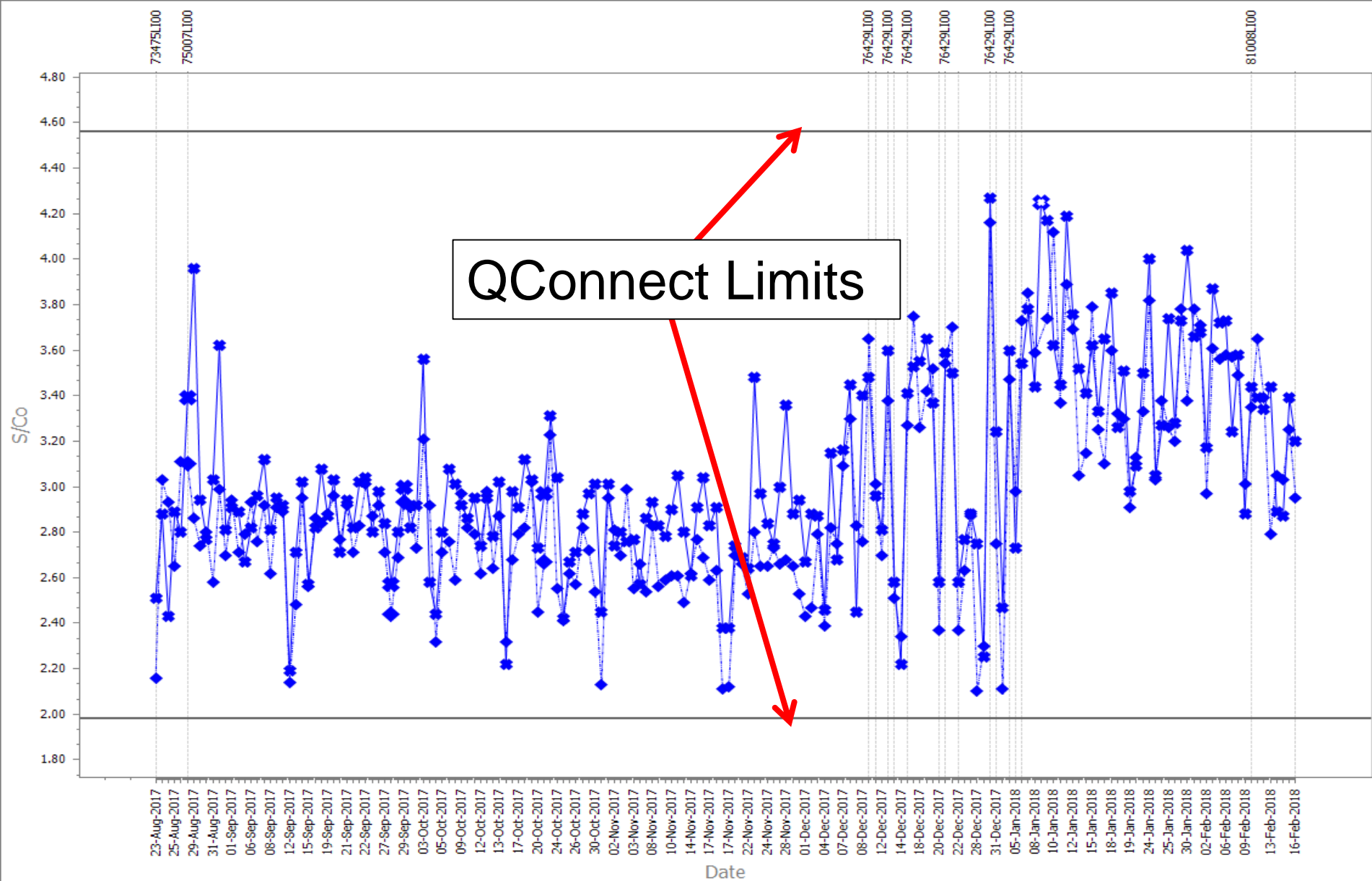
# What is the cause of the change

A - Change not significant

B - Reagent lot number

C - Instrument

D - Process/People



QConnect Limits

Laboratory:3    From:01-Feb-2017 to 20-Feb-2018    Assays:Abbott PRISM HIV Ag/Ab Combo ChLIA;    Analyte:HIV p24 Ag/Anti-HIV    EQC:QConnect Purple    EQC LotNumber:DM17076;

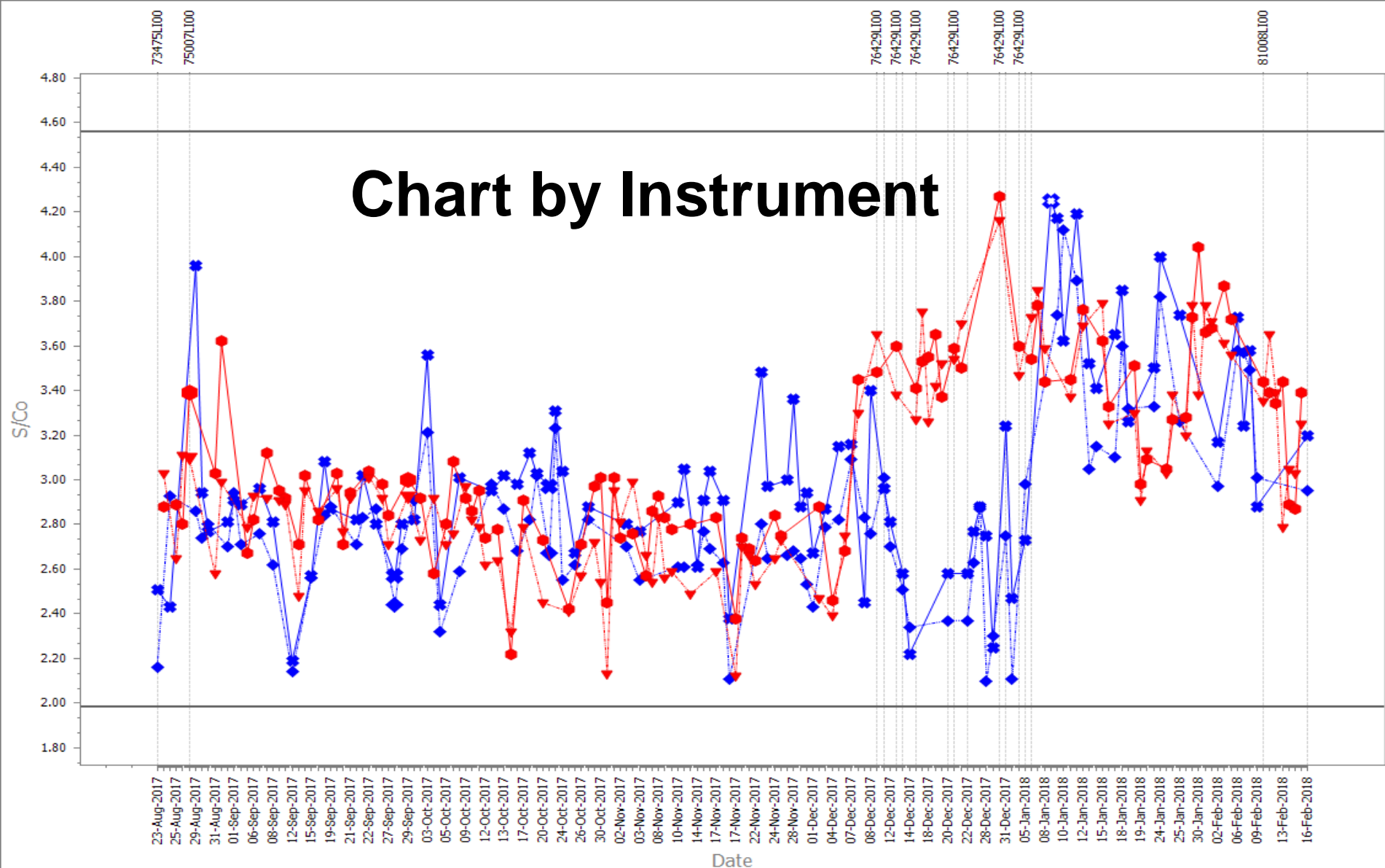
▲ Subchannel A    ▼ Subchannel B

Trend: Assay    ● Abbott PRISM HIV Ag/Ab Combo ChLIA/Subchannel A    ◆ Abbott PRISM HIV Ag/Ab Combo ChLIA/Subchannel B

— NRL Range:1.98 to 4.56



# Chart by Instrument



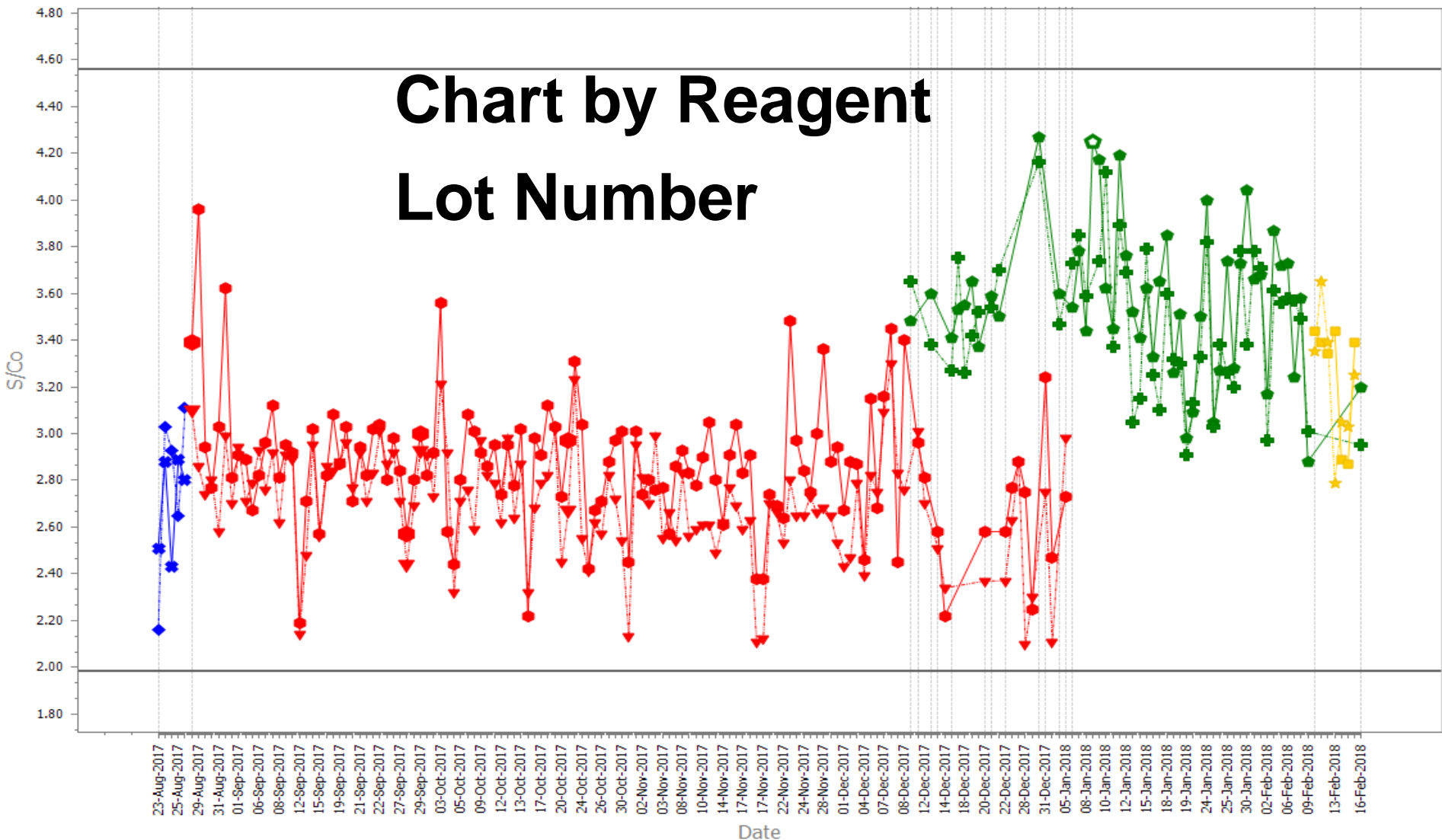
Laboratory:3 From:01-Feb-2017 to 20-Feb-2018 Assays:Abbott PRISM HIV Ag/Ab Combo ChLIA; Analyte:HIV p24 Ag/Anti-HIV EQC:QConnect Purple EQC LotNumber:DM17076;

▲ Subchannel A ▼ Subchannel B

Trend: Instrument; Test Process: Detection —●— PRISM/Channel 3 (1045)/Subchannel A —◆— PRISM/Channel 3 (1045)/Subchannel B —●— PRISM/Channel 3 (1528)/Subchannel A  
—▼— PRISM/Channel 3 (1528)/Subchannel B

— NRL Range:1.98 to 4.56

# Chart by Reagent Lot Number



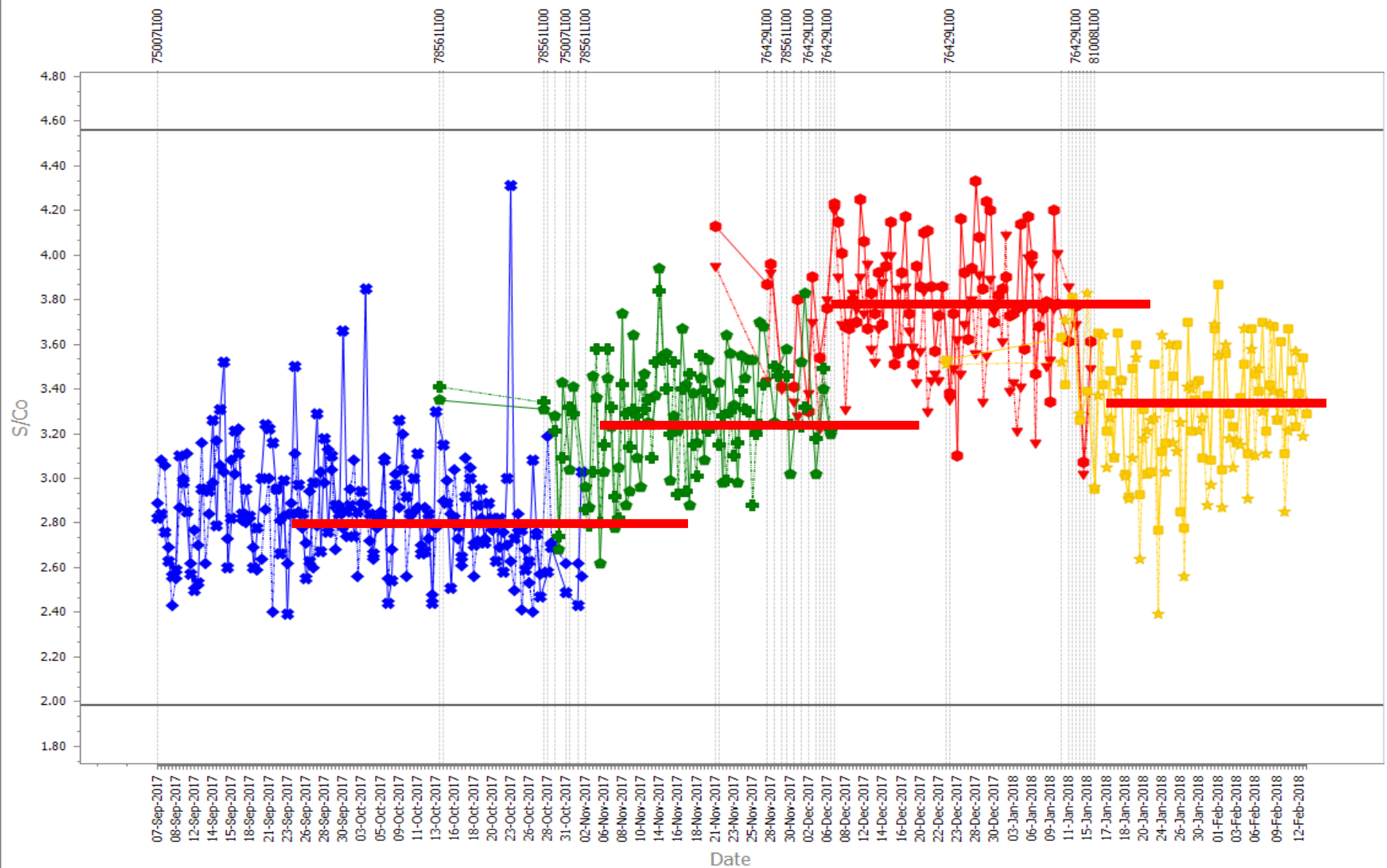
Laboratory:3    From:01-Feb-2017 to 20-Feb-2018    Assays:Abbott PRISM HIV Ag/Ab Combo ChLIA;    Analyte:HIV p24 Ag/Anti-HIV    EQC:QConnect Purple    EQC LotNumber:DM17076;

▲ Subchannel A    ▼ Subchannel B

Trend: KitLot Number; Test Process: Detection

● 73475LI00/Subchannel A    ◆ 73475LI00/Subchannel B    ● 75007LI00/Subchannel A    ▼ 75007LI00/Subchannel B    ● 76429LI00/Subchannel A  
◆ 76429LI00/Subchannel B    ■ 81008LI00/Subchannel A    ★ 81008LI00/Subchannel B

— NRL Range:1.98 to 4.56



Laboratory:11    From:01-Feb-2017 to 20-Feb-2018    Assays:Abbott PRISM HIV Ag/Ab Combo ChLIA;    Analyte:HIV p24 Ag/Anti-HIV    EQC:QConnect Purple    EQC LotNumber:DM17076;

▲ Subchannel A    ▼ Subchannel B

Trend: KitLot Number; Test Process: Detection    ◆ 75007LI00/Subchannel A    ◆ 75007LI00/Subchannel B    ● 76429LI00/Subchannel A    ▼ 76429LI00/Subchannel B    ◆ 78561LI00/Subchannel A

◆ 78561LI00/Subchannel B    ■ 81008LI00/Subchannel A    ★ 81008LI00/Subchannel B

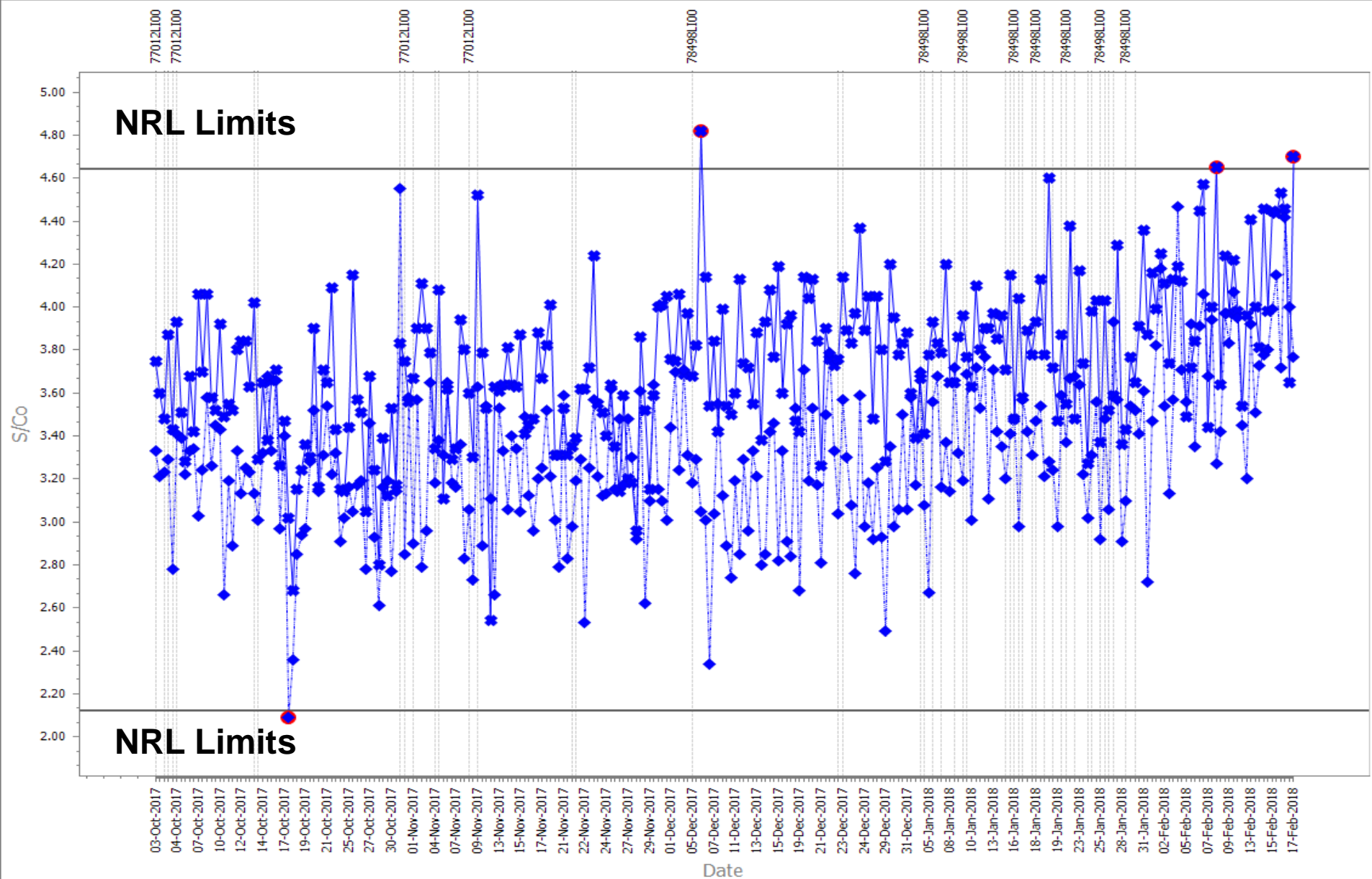
— NRL Range:1.98 to 4.56

# Scenario 1

***Answer:***

Classic reagent lot variation

# Scenario 2



Laboratory:11

From:03-Oct-2017 to 22-Feb-2018

Assays:Abbott PRISM HCV ChLIA;

Analyte:anti-HCV

EQC:QConnect Purple

EQC LotNumber:DM17076;

▲ Subchannel A ▼ Subchannel B

Trend: Assay —●— Abbott PRISM HCV ChLIA/Subchannel A —◆— Abbott PRISM HCV ChLIA/Subchannel B

— NRL Range:2.12 to 4.64

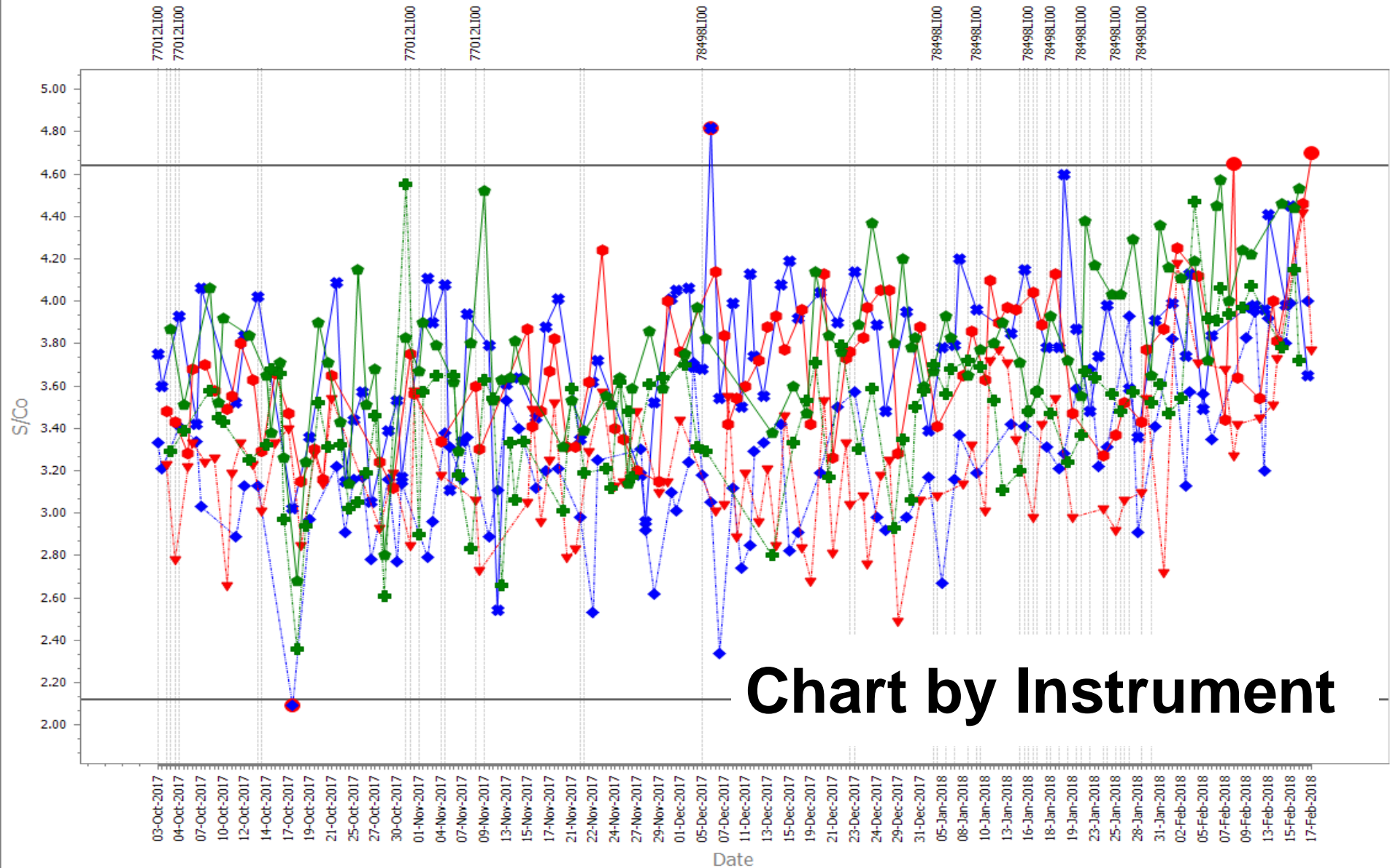
# What is the cause of the change

A - Change not significant

B - Reagent lot number

C - Instrument

D - Process/People



Laboratory:11

From:03-Oct-2017 to 22-Feb-2018

Assays:Abbott PRISM HCV ChLIA;

Analyte:anti-HCV

EQC:QConnect Purple

EQCLotNumber:DM17076;

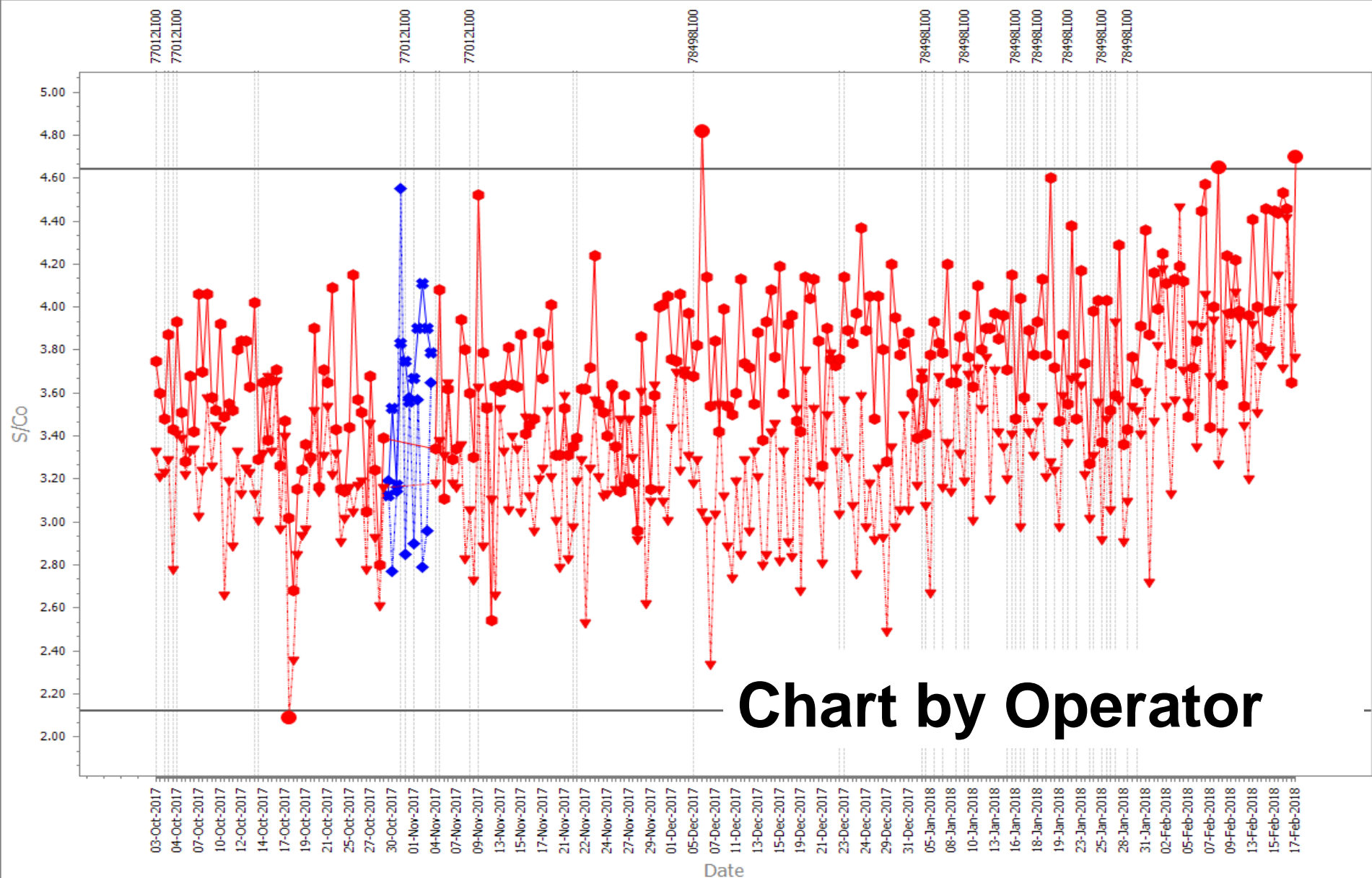
▲ Subchannel A ▼ Subchannel B

Trend: Instrument; Test Process: Detection

● PRISM/Channel 2 (1048)/Subchannel A  
 ● PRISM/Channel 2 (1048)/Subchannel B  
 ● PRISM/Channel 2 (1092)/Subchannel A  
 ● PRISM/Channel 2 (1092)/Subchannel B  
 ● PRISM/Channel 2 (1118)/Subchannel A  
 ● PRISM/Channel 2 (1118)/Subchannel B

— NRL Range:2.12 to 4.64





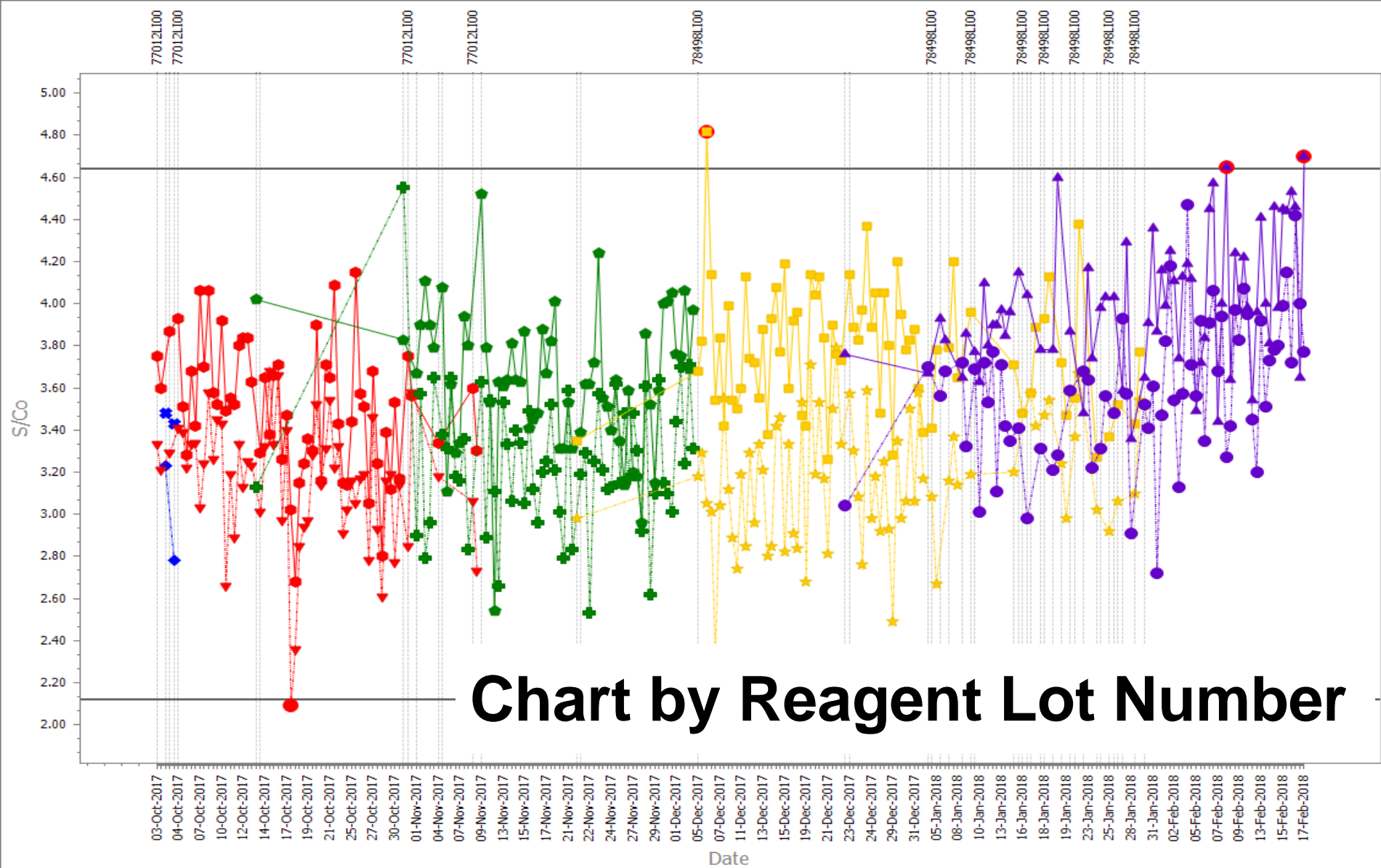
# Chart by Operator

Laboratory:11    From:03-Oct-2017 to 22-Feb-2018    Assays:Abbott PRISM HCV ChLIA;    Analyte:anti-HCV    EQC:QConnect Purple    EQC LotNumber:DM17076;

▲ Subchannel A    ▼ Subchannel B

Trend: Operator; Test Process: Detedion    ● DV/Subchannel A    ◆ DV/Subchannel B    ● SH/Subchannel A    ▼ SH/Subchannel B

— NRL Range:2.12 to 4.64



# Chart by Reagent Lot Number

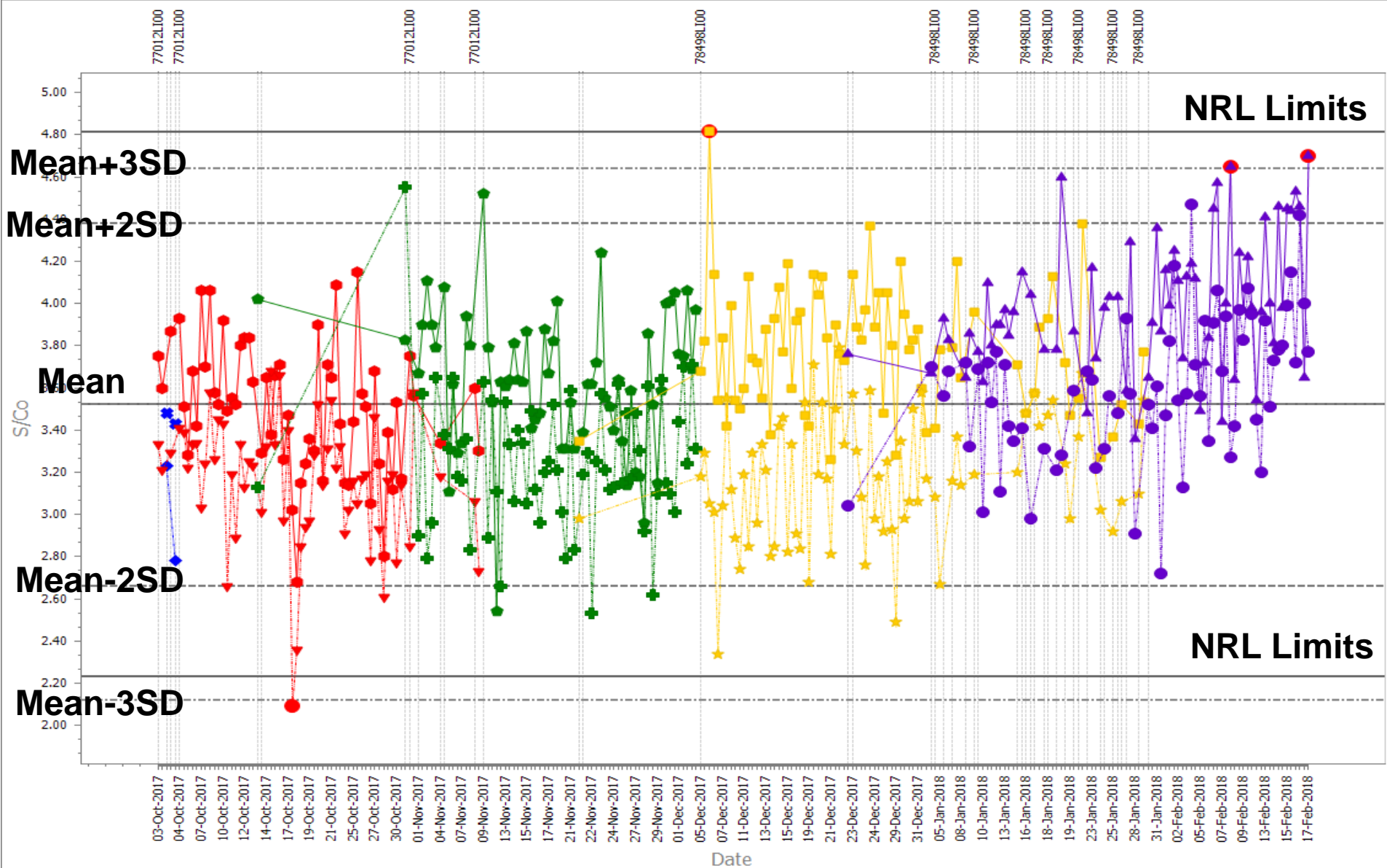
Laboratory:11    From:03-Oct-2017 to 22-Feb-2018    Assays:Abbott PRISM HCV ChIA;    Analyte:anti-HCV    EQC:QConnect Purple    EQC LotNumber:DM17076;

▲ Subchannel A    ▼ Subchannel B

Trend: KitLot Number; Test Process: Detection

- 75017L100/Subchannel A (Blue circle)
- 75017L100/Subchannel B (Blue diamond)
- 77012L100/Subchannel A (Red circle)
- 77012L100/Subchannel B (Red inverted triangle)
- 77308L100/Subchannel A (Green diamond)
- 77308L100/Subchannel B (Green circle)
- 78498L100/Subchannel A (Yellow square)
- 78498L100/Subchannel B (Yellow star)
- 79546L100/Subchannel A (Purple triangle)
- 79546L100/Subchannel B (Purple circle)

— NRL Range:2.12 to 4.64



Laboratory:11

From:03-Oct-2017 to 22-Feb-2018

Assays:Abbott PRISM HCV ChLIA;

Analyte:anti-HCV

EQC:QConnect Purple

EQCLotNumber:DM17076;

▲ Subchannel A ▼ Subchannel B

Trend: KitLot Number; Test Process: Detection

● 75017LI00/Subchannel A   ● 75017LI00/Subchannel B   ● 77012LI00/Subchannel A   ● 77012LI00/Subchannel B   ● 77308LI00/Subchannel A  
 ● 77308LI00/Subchannel B   ● 78498LI00/Subchannel A   ● 78498LI00/Subchannel B   ● 79546LI00/Subchannel A   ● 79546LI00/Subchannel B

— Your Laboratory - Visible Data; Mean=3.52; +3SD=4.81; -3SD=2.23   - - - - Your Laboratory - Visible Data; Mean=3.52; +2SD=4.38; -2SD=2.66   - - - - NRL Range: 2.12 to 4.64

# Scenario 2

- **Normal QC performance**
- $n = 593$
- Data within NRL Limits = 99.3% (n=589)
- Number of points within  
 $\pm 2SD$  (n=569) and  $\pm 3SD$  (n=591)
- Compare this with statistical likelihood  
 $\pm 2SD = 95\%$  (actual 95.95%)  
 $\pm 3SD = 99.7\%$  (actual 99.66%)

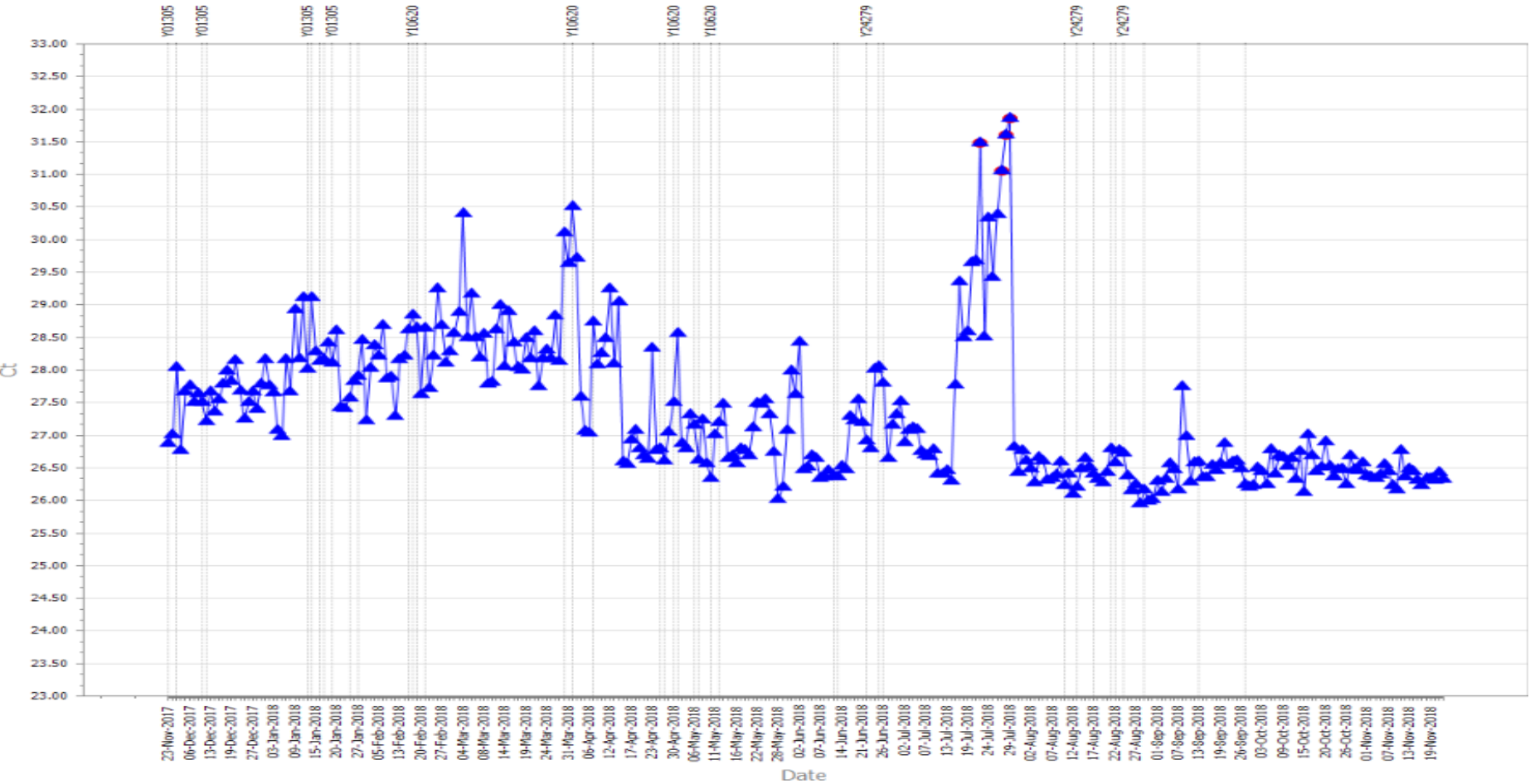
# Scenario 2

***Answer:***

Nothing to see here folks

# Scenario 3

# Scenario 3



HPV NAT on Roche COBAS HPV Assay

# What is the cause of the change

A - Change not significant **A**

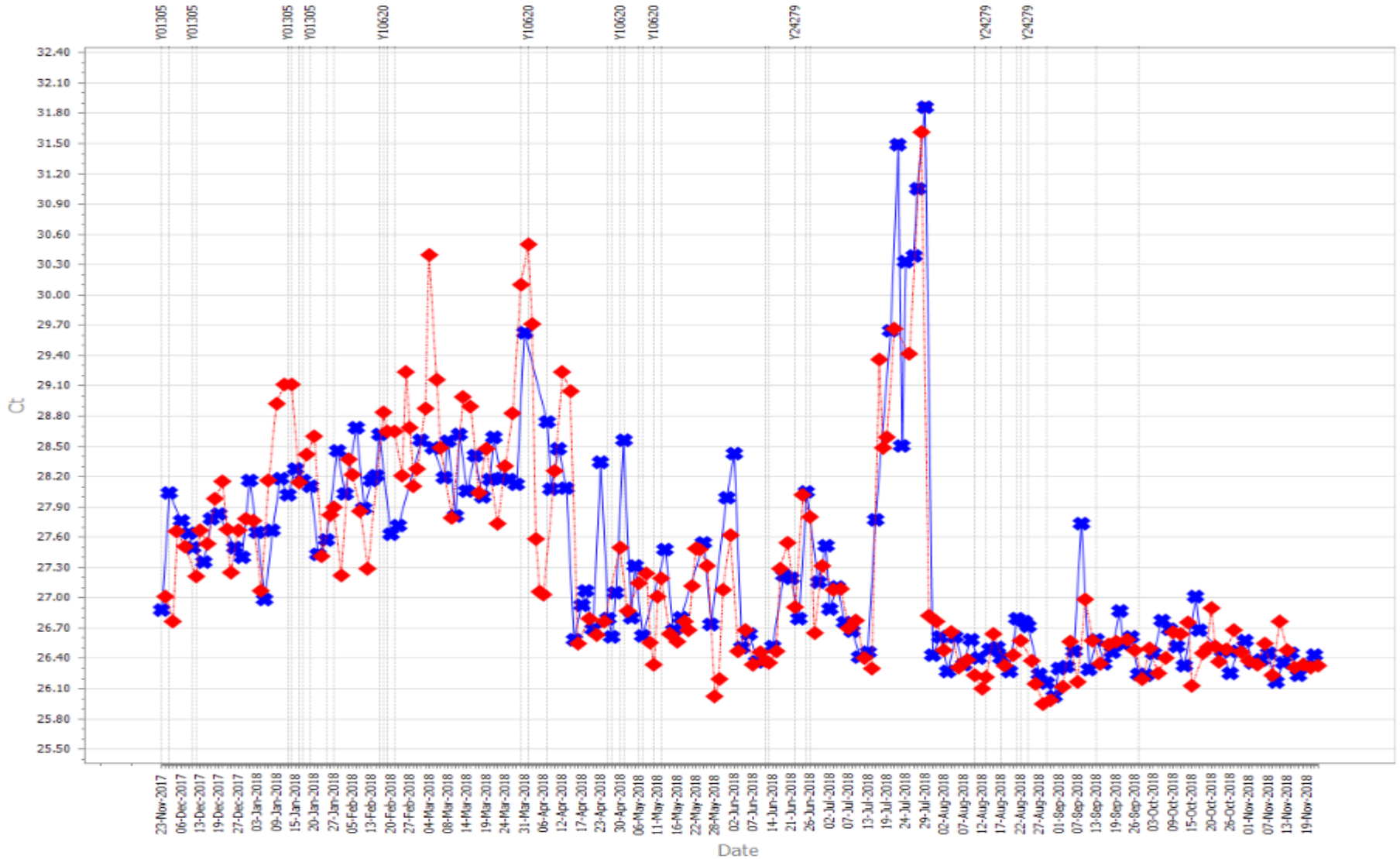
B - Reagent lot number **B**

C - Instrument **C**

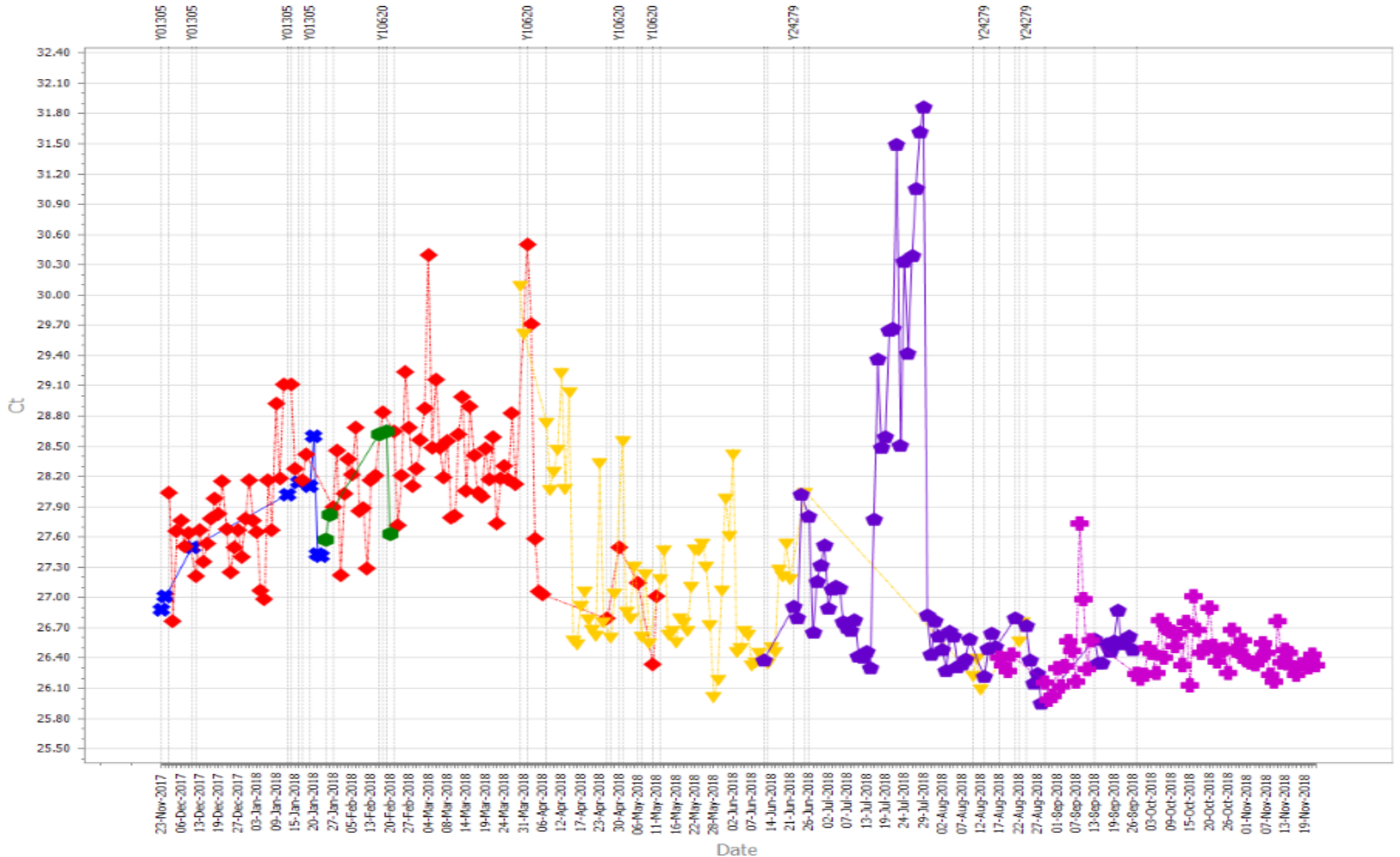
D - Process/People **D**



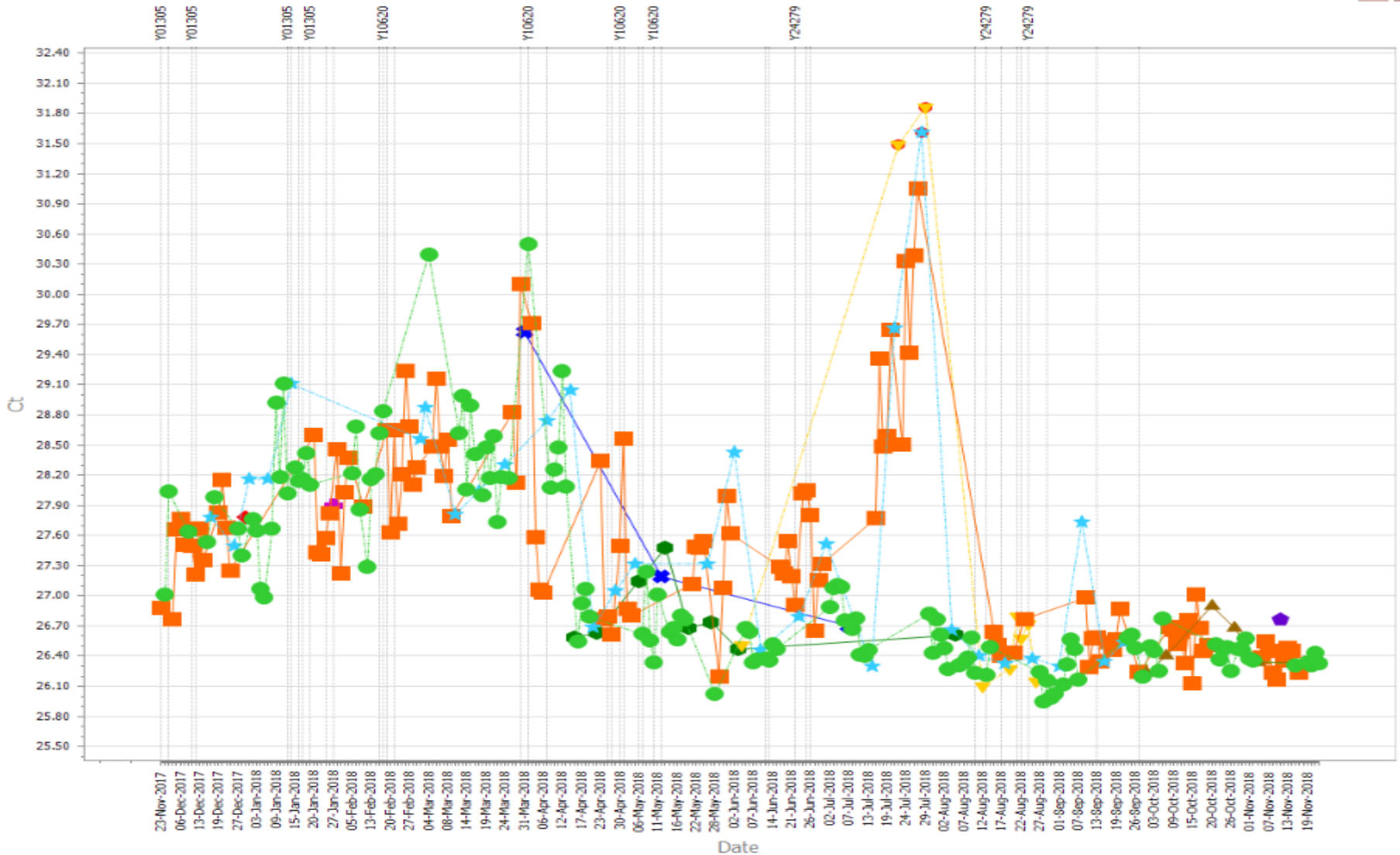
# Trending by Instrument



# Trending by Reagent Lot



# Trending by Operator



# Scenario 3

- Investigation initiated
- Observed processing of samples by each operator
- Discrepancies in processes detected
- Re-trained all operators
- Results returned to expected level

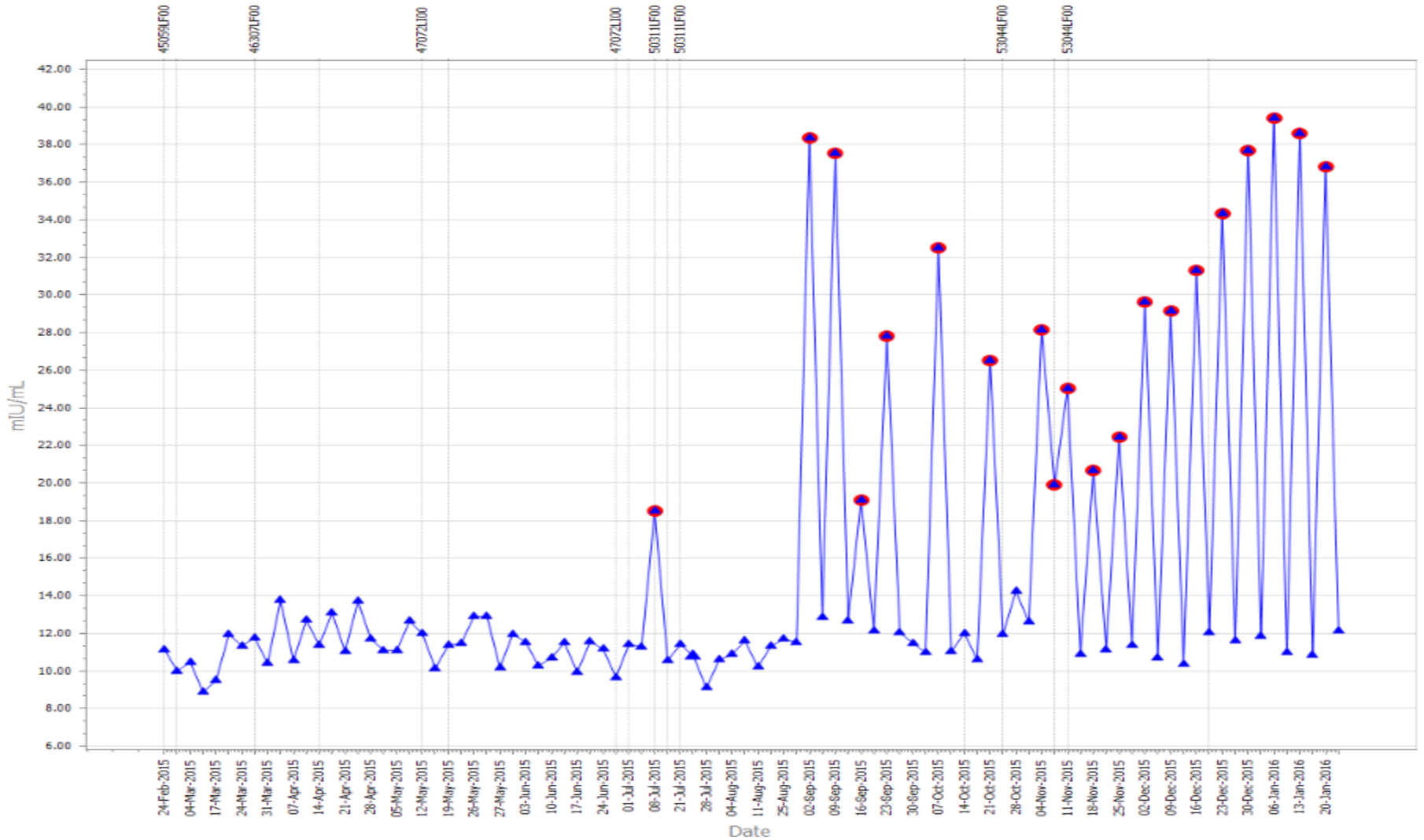
# Scenario 3

*Answer:*

Process issue

# Scenario 4

# Anti-HBs testing on Abbott Architect



# What is the cause of the change

A - Change not significant **A**

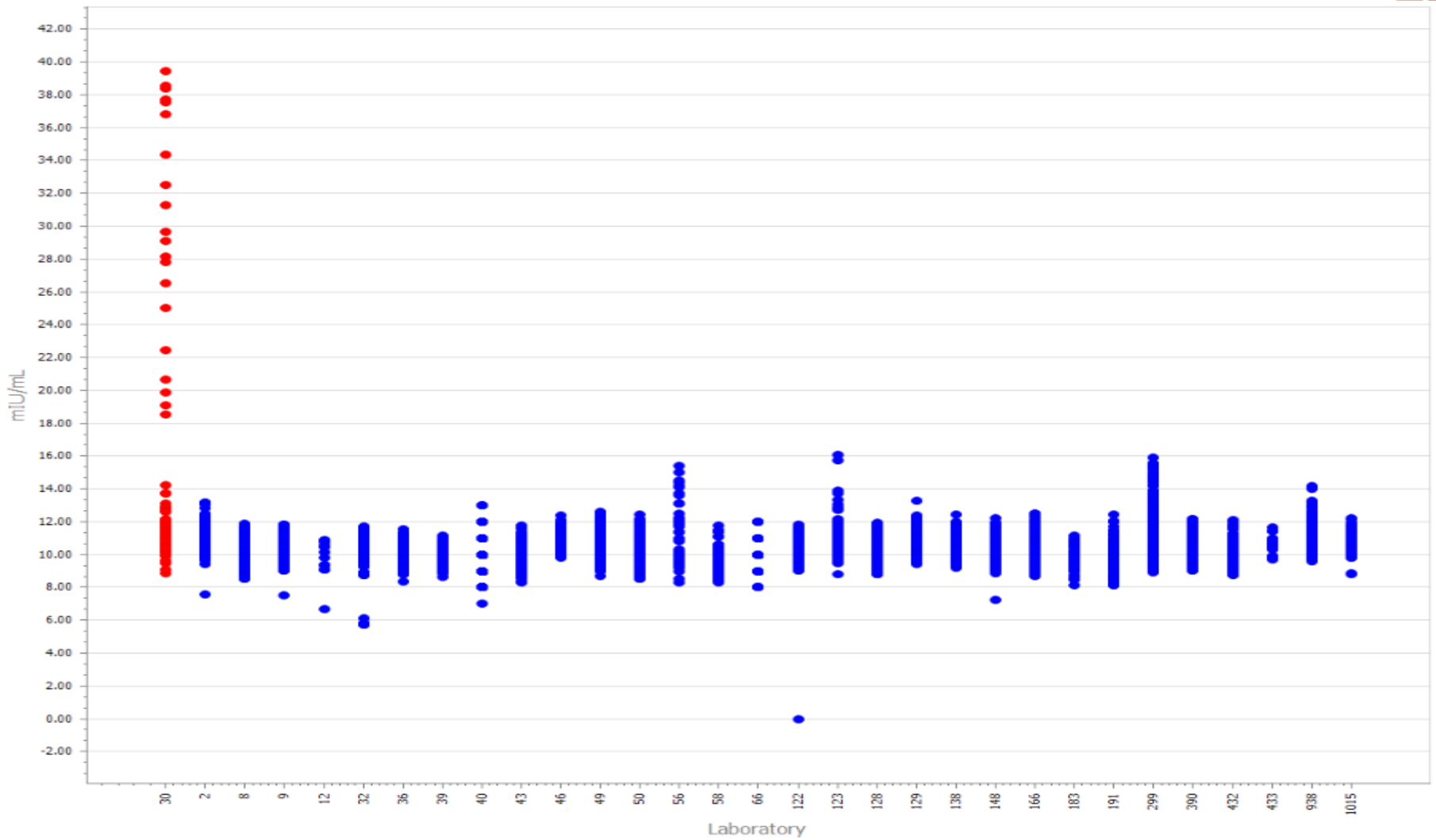
B - Reagent lot number **B**

C - Instrument **C**

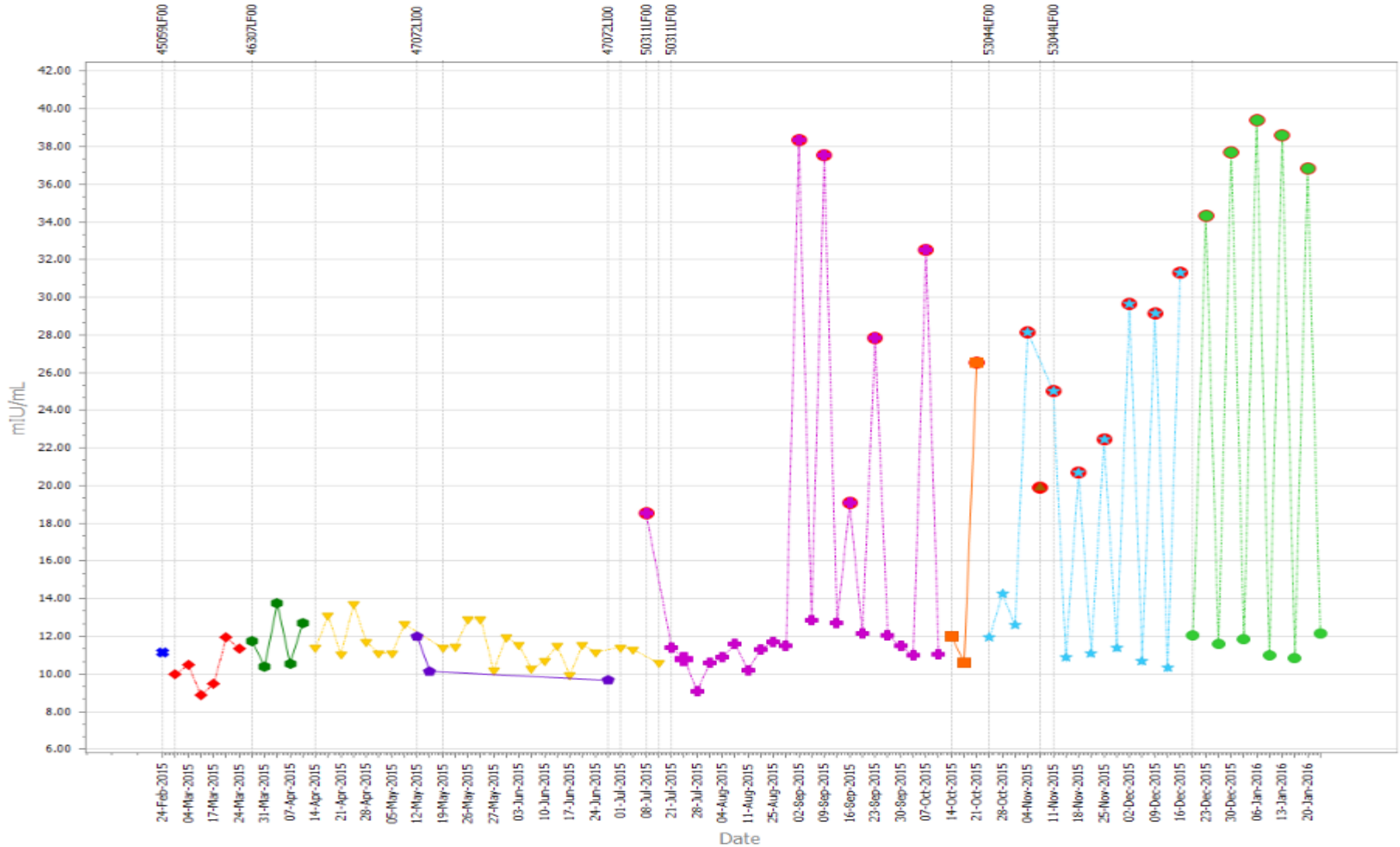
D - Process/People **D**



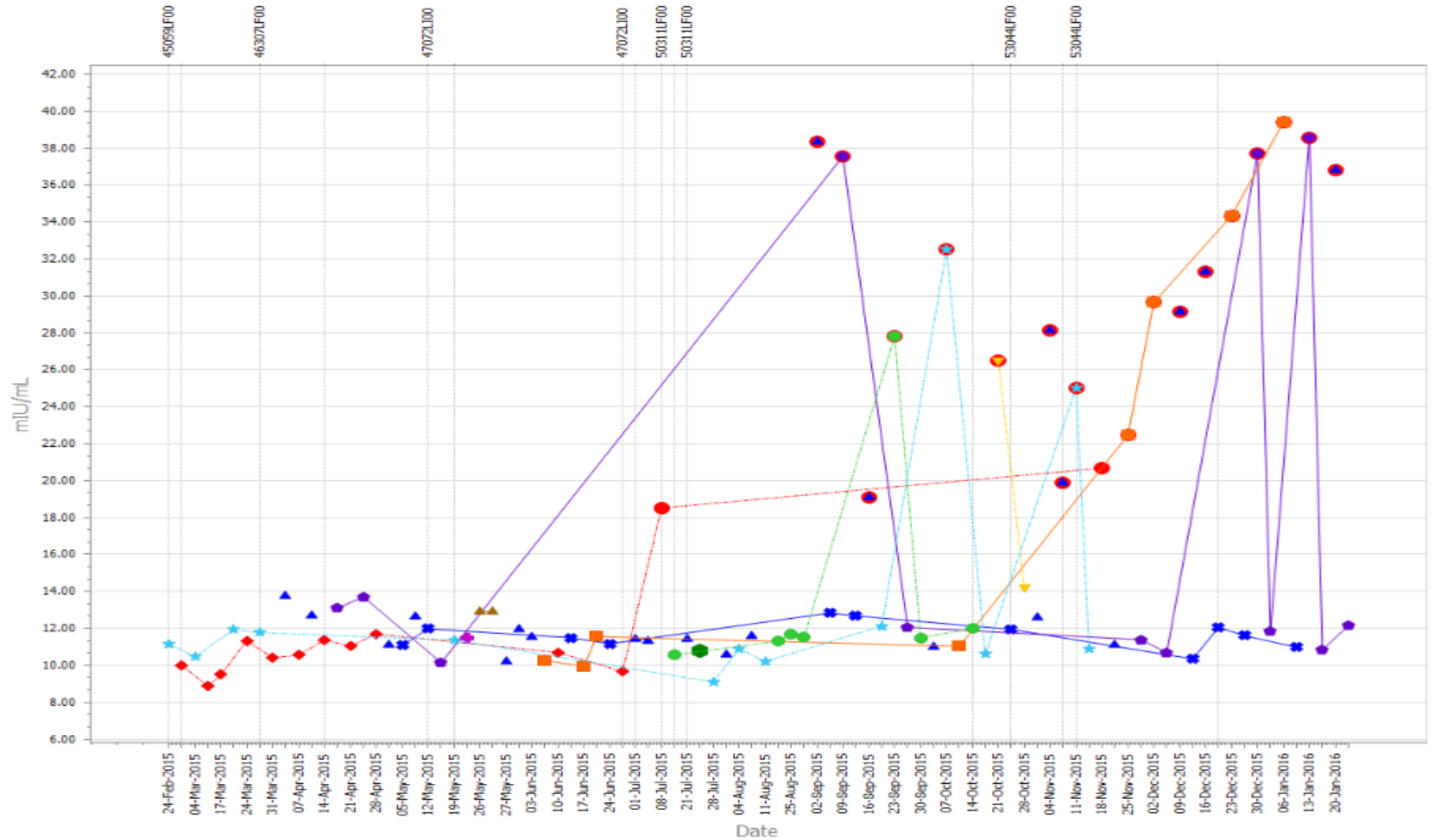
# Comparison with Peers



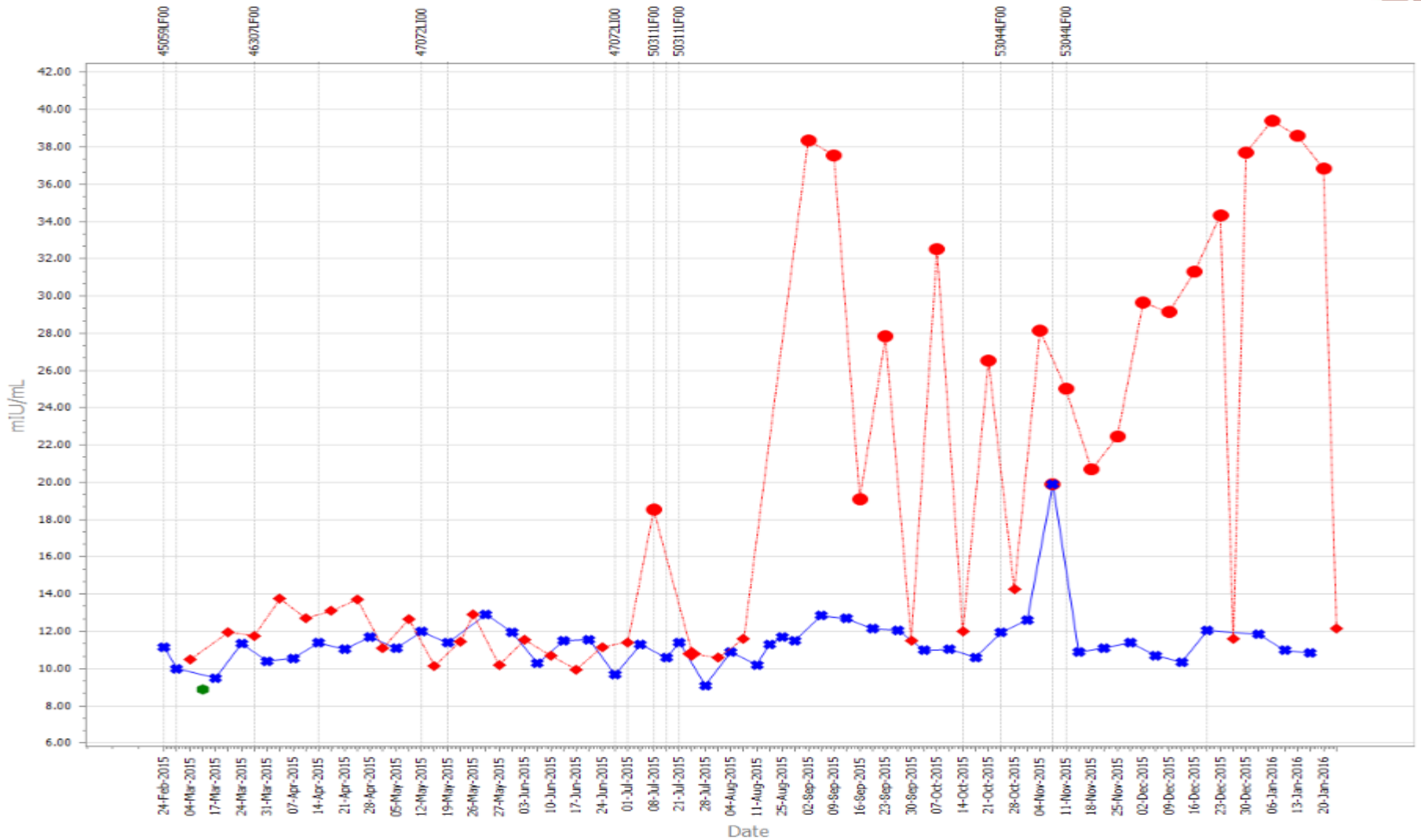
# Trending by Reagent Lot



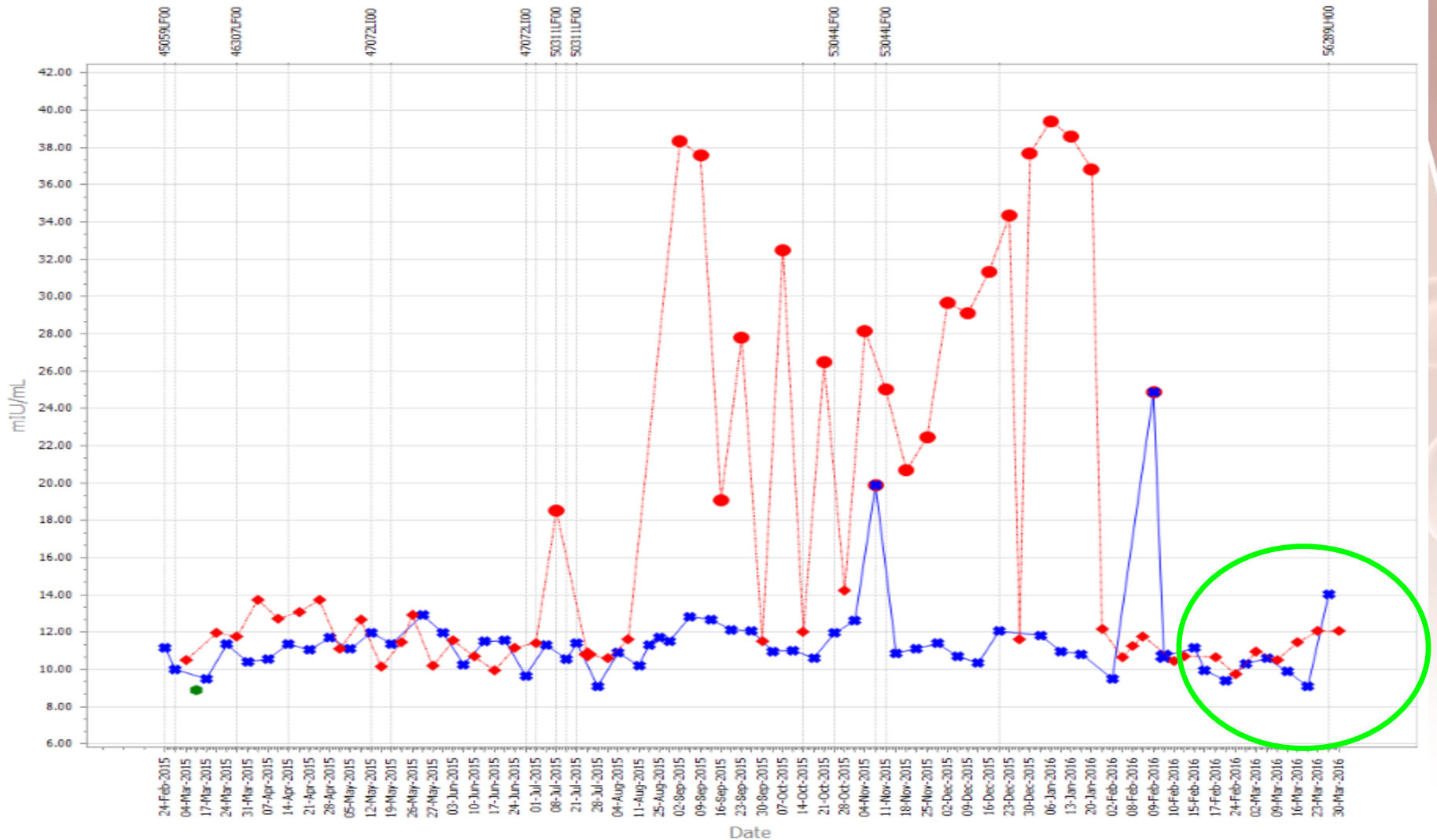
# Trending by Operator



# Trending by Instrument



# Resolution of Issue



# Scenario 4

***Answer:***

Instrument Issue

# Conclusions

# Conclusions

- Measuring systems must be fit for purpose
- Measurement systems have normal variation
- Variation is derived from people, process, components and equipment
- Dynamic range of serology assays is not always linear
- Choose QC sample that reacts at the linear portion of the dilution curve



# Conclusions

- Fundamental differences between testing for an inert chemical and a functional, biological analyte
- Recommend QC at start of day for automated platforms or every microtiter plate
- Monitoring QC results without reference to a peer-group only monitors precision
- Traditional QC approach assumptions are not true for infectious disease serology

# Conclusions

- Commutability of serology QC samples and patient samples should not be assumed
- Using a diluted sample to mimic early infection is a flawed concept
- Reagent lot variation is the major contributor to normal variation of serology assays

# Conclusions

- There are usually insufficient runs to re-set limits before reagent lot is exhausted
- By re-setting limits, you are accepting that the variation is normal
- Viral Load Testing has similarities with clinical chemistry and serology
- Promotion of methods for establishing acceptance criteria for viral load QC needs to be based on real data

# Conclusions

- Collect metadata with QC results
  - Date
  - Instrument(s) identification
  - Reagent lot number(s)
  - Operators
  - QC lot number
  - Calibration and maintenance data

# Conclusions

- Monitoring Quality Control in a systematic manner can identify unexpected variation that may, in time, contribute to incorrect patient results
- QC is done in addition to kit controls and EQA and QMS
- Do not blindly follow without evidence –  
**you are scientist**

# Thank-you!

Think different

